

**Proteomics Informatics -
Protein identification III:
de novo sequencing (Week 6)**

De Novo Sequencing of MS Spectra

Only a manually confirmed spectrum
is a correct spectrum

Beatrix Ueberheide

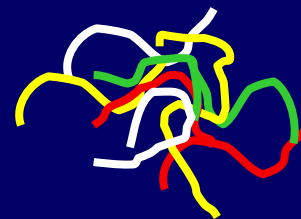
March 12th 2013

Biological Mass Spectrometry



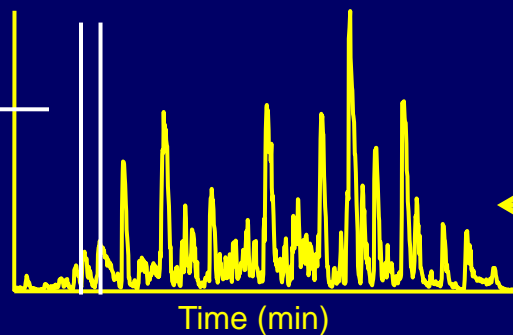
Protein(s)

Proteolytic digestion

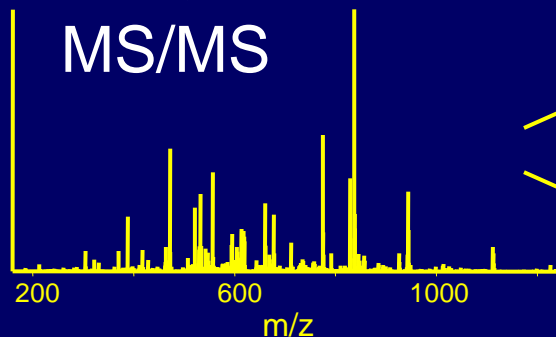
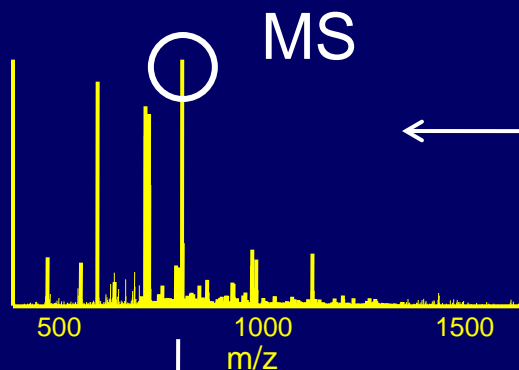


Peptides

Base Peak Chromatogram



Mass Spectrometer



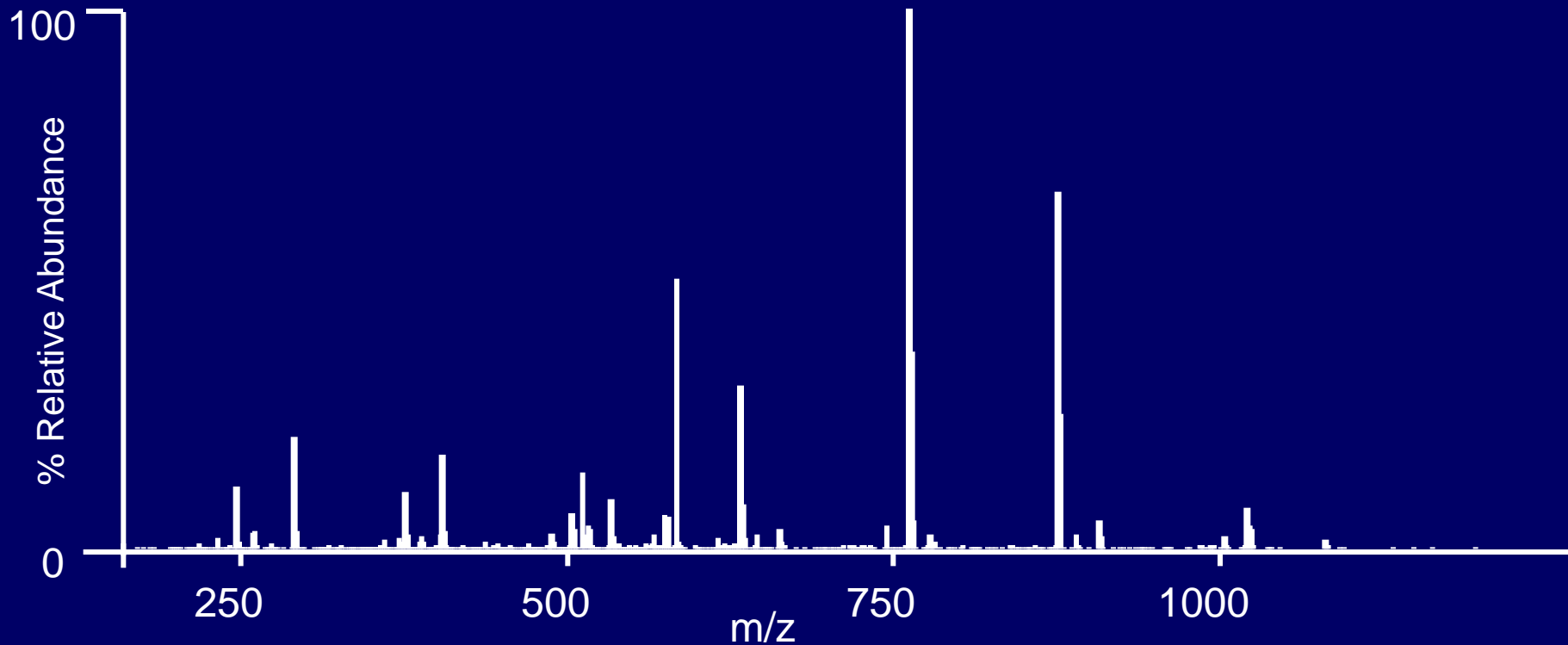
Database Search

Manual Interpretation



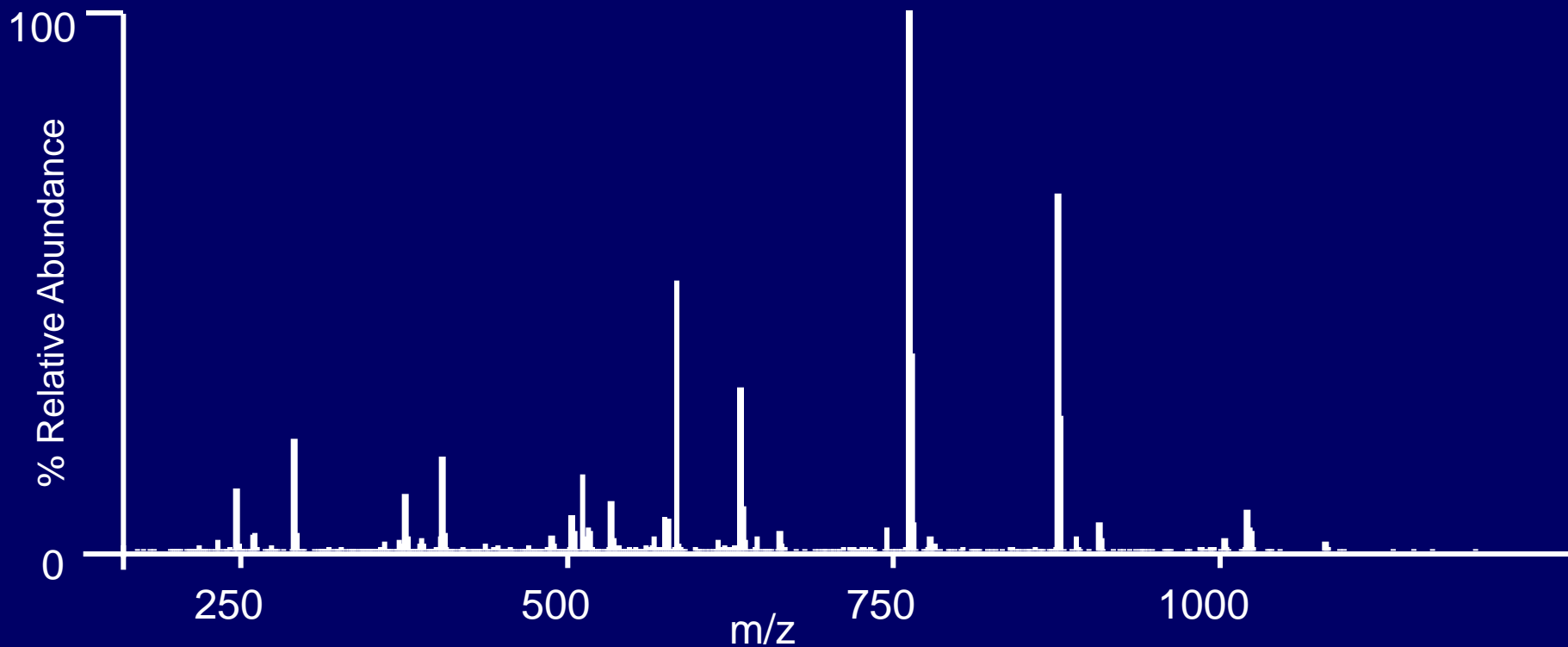
Peptide Sequencing using Mass Spectrometry

S G F L E E D E L K



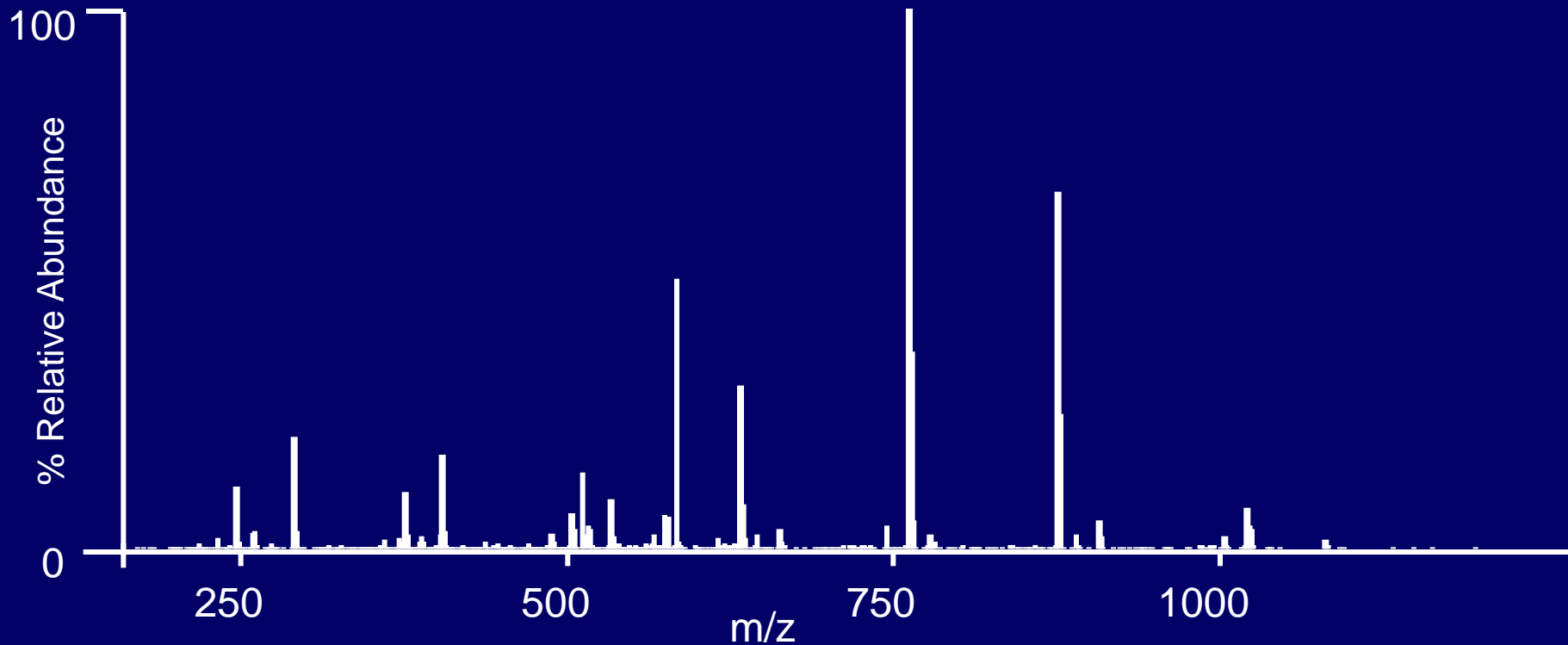
Peptide Sequencing using Mass Spectrometry

88 145 292 405 534 663 778 907 1020 1166 b ions
S G F L E E D E L K



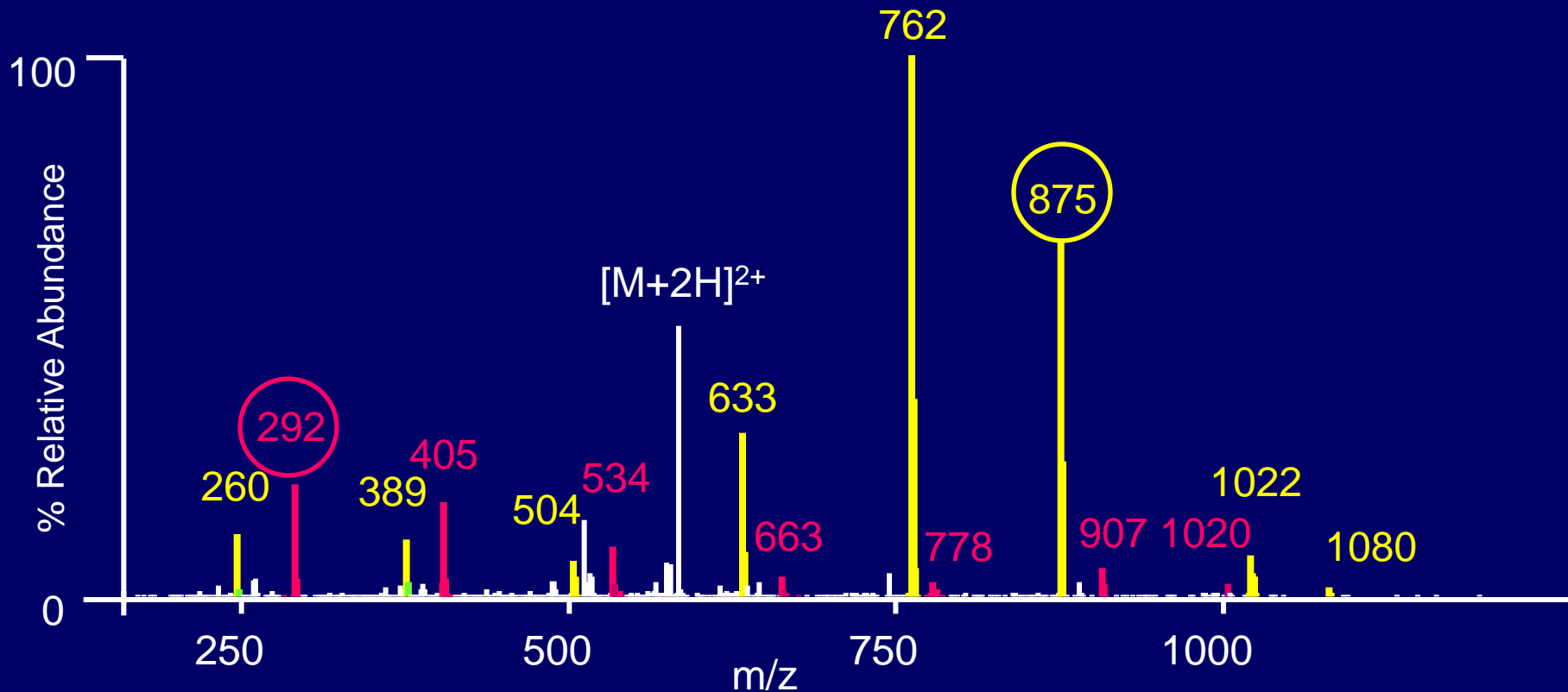
Peptide Sequencing using Mass Spectrometry

S G F L E E D E L K
1166 1080 1022 875 762 633 504 389 260 147 y ions



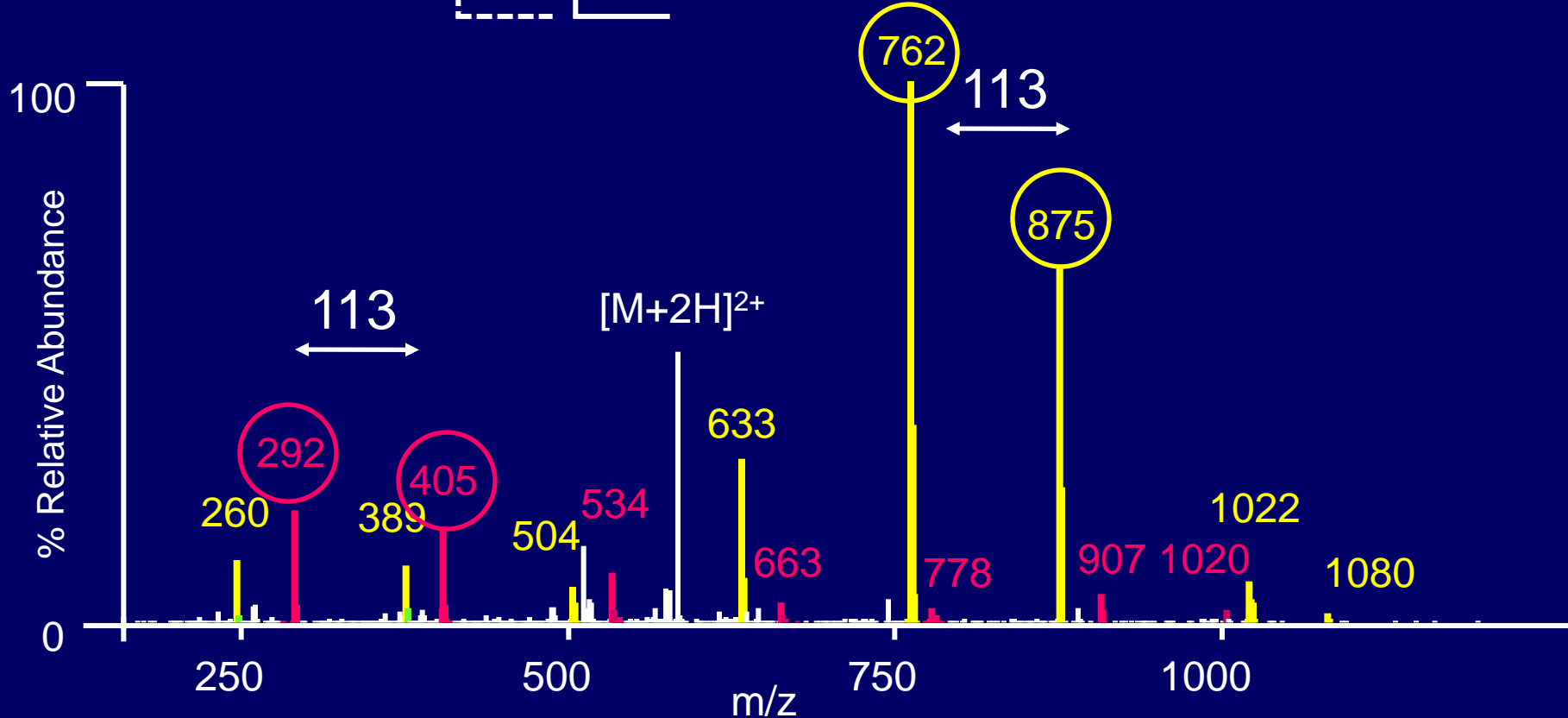
Peptide Sequencing using Mass Spectrometry

88	145	292	405	534	663	778	907	1020	1166	b ions
S	G	F	L	E	E	D	E	L	K	
1166	1080	1022	875	762	633	504	389	260	147	y ions



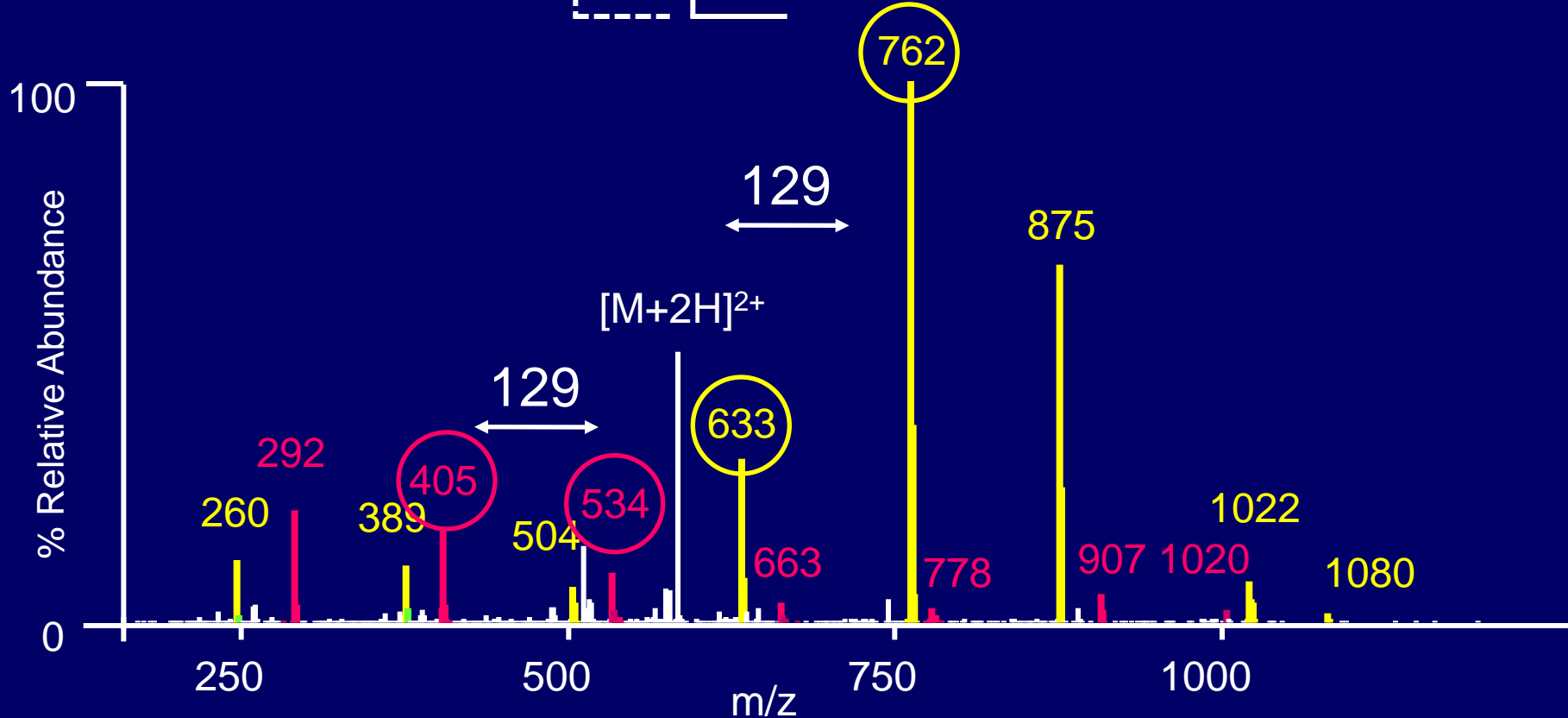
Peptide Sequencing using Mass Spectrometry

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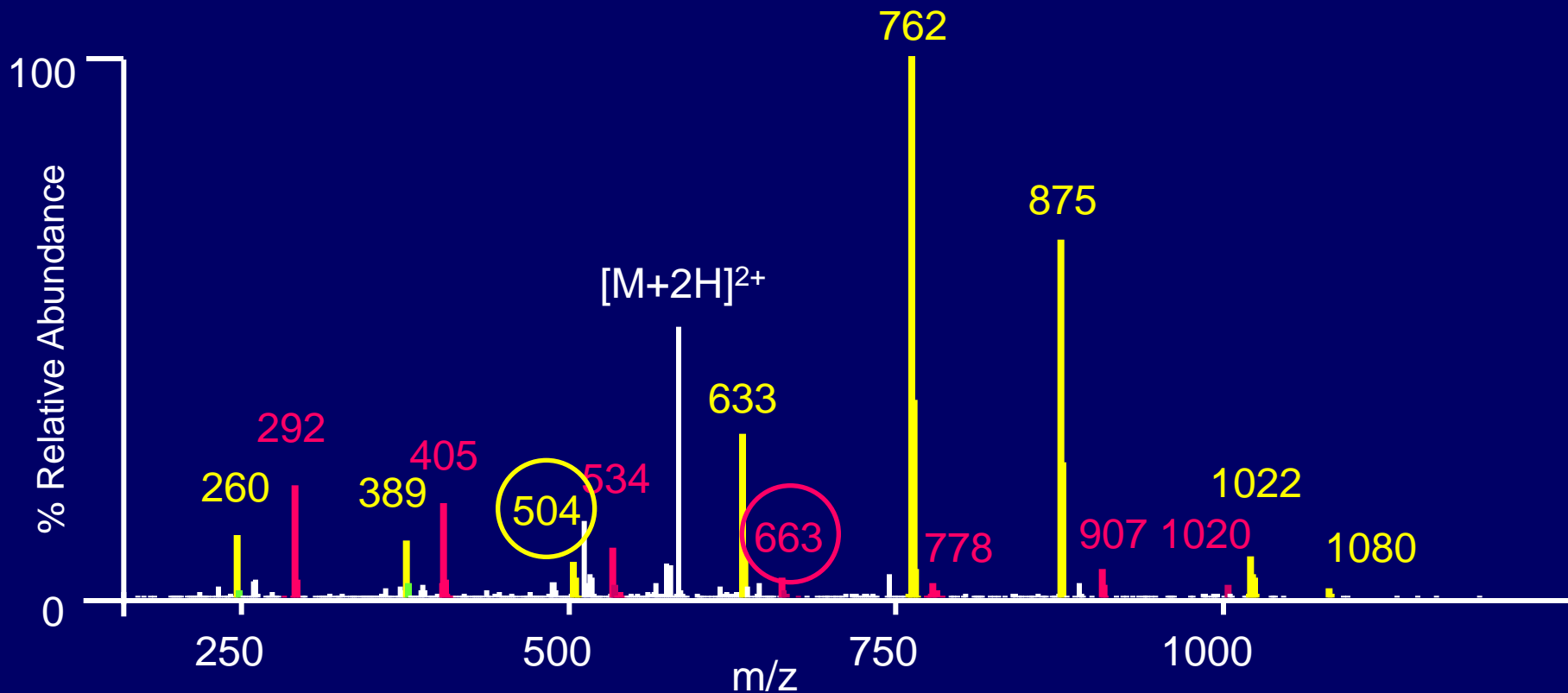
Peptide Sequencing using Mass Spectrometry

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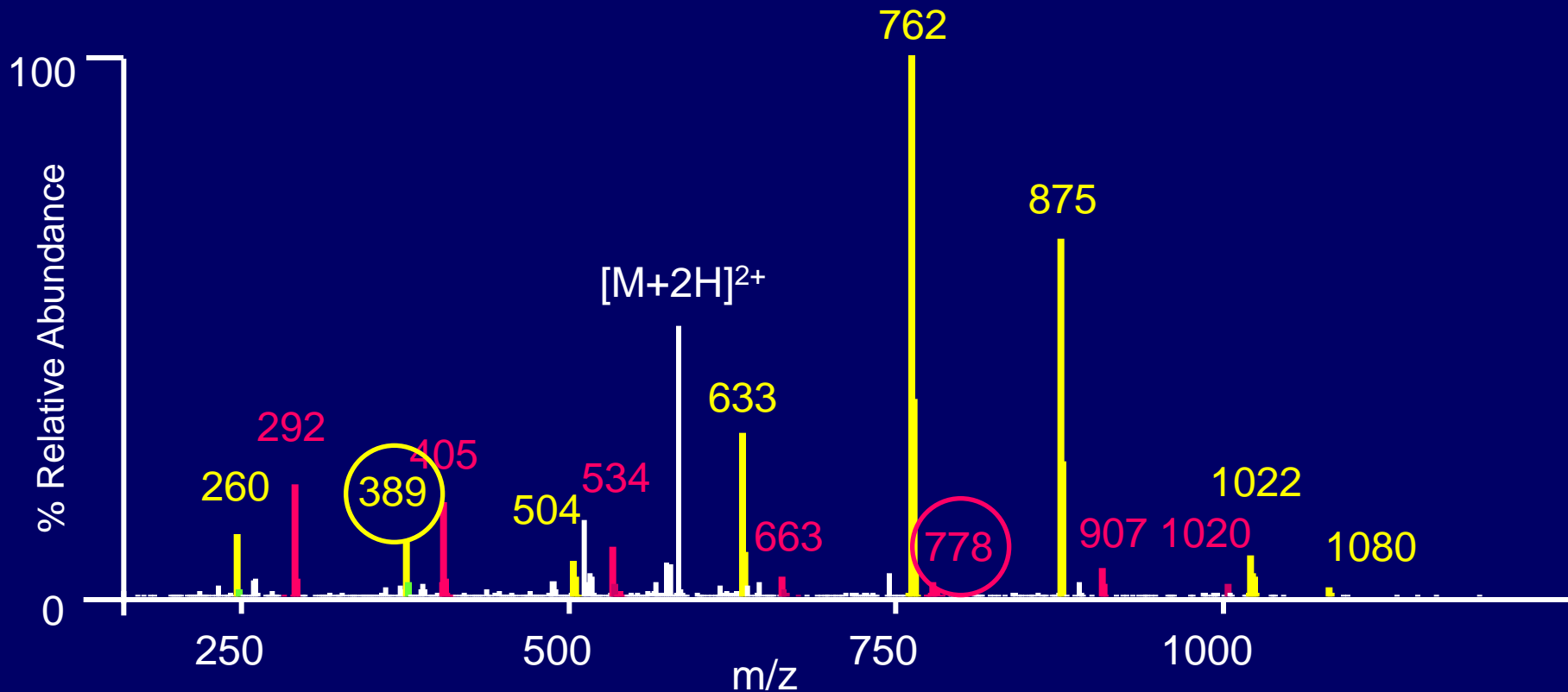
Peptide Sequencing using Mass Spectrometry

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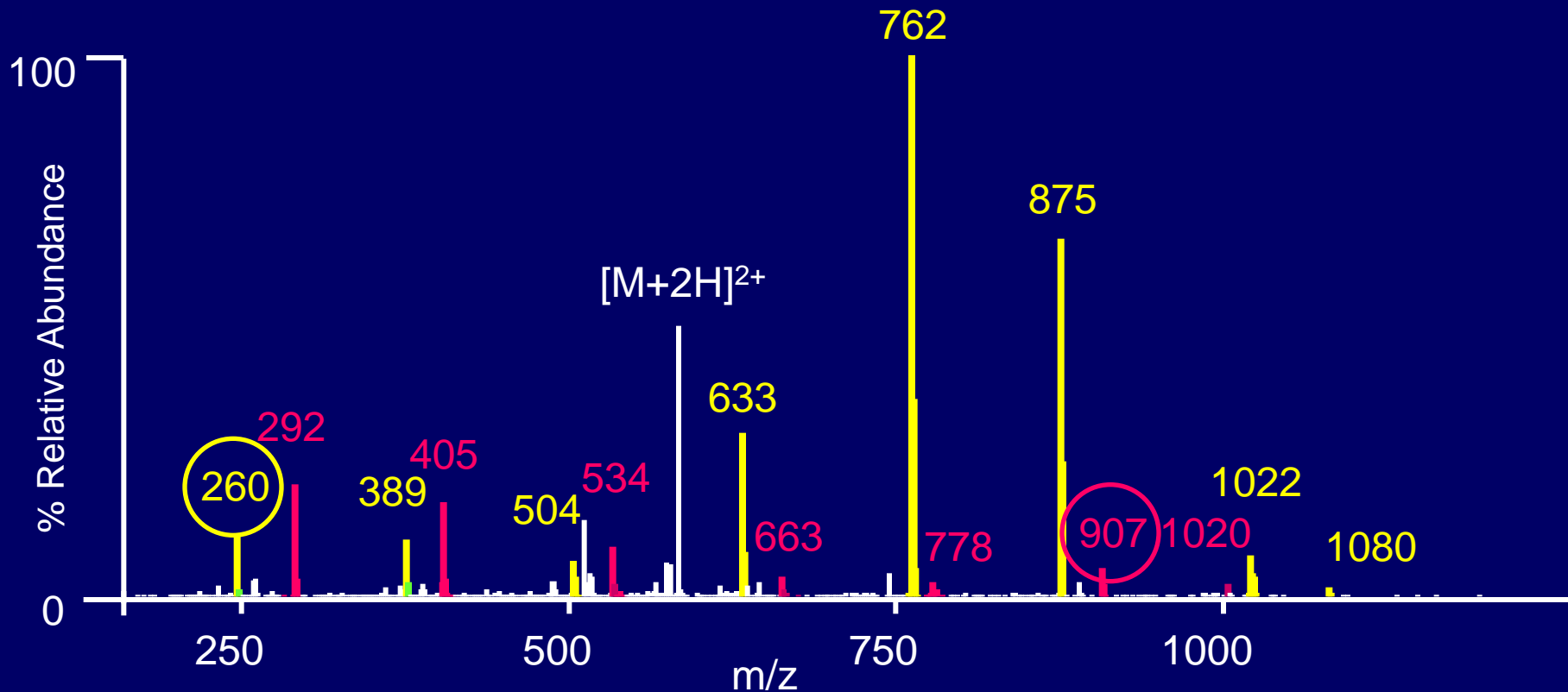
Peptide Sequencing using Mass Spectrometry

88	145	292	405	534	663	778	907	1020	1166	b ions
S	G	F	L	E	E	D	E	L	K	
1166	1080	1022	875	762	633	504	389	260	147	y ions

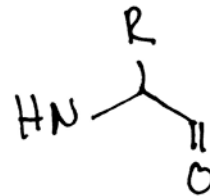
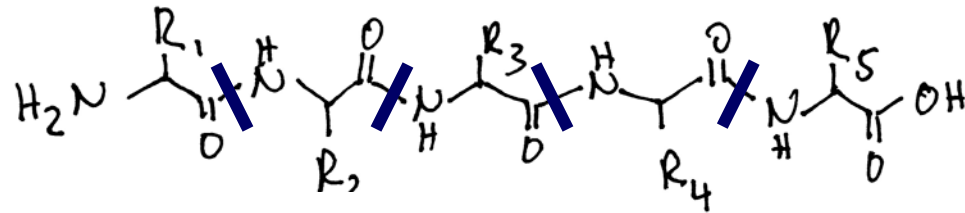


Peptide Sequencing using Mass Spectrometry

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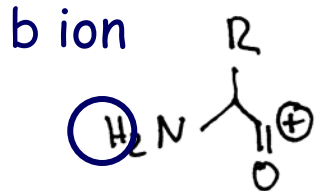


How to Sequence: CAD

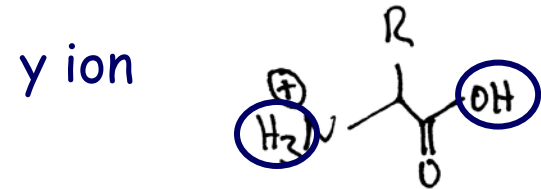


Residue Mass (RM)

The very first N- and C-terminal fragment ions are not just their corresponding residue masses. The peptides N or C-terminus has to be taken into account.



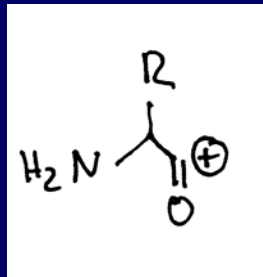
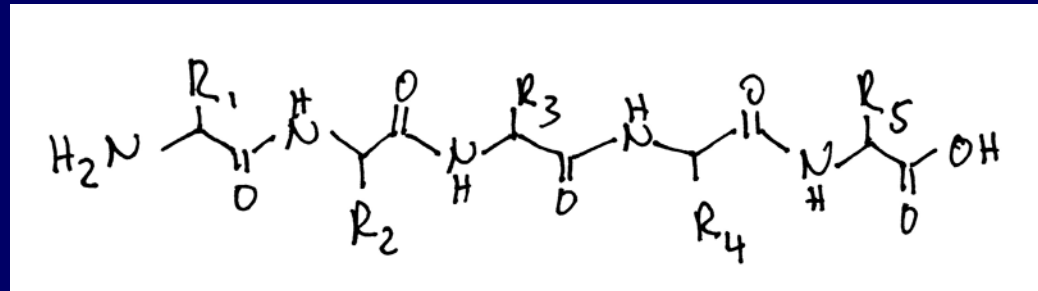
$$b_1 = RM + 1$$



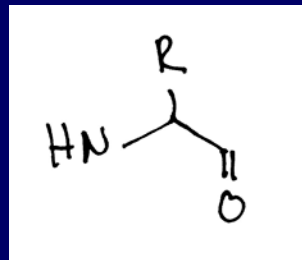
$$y_1 = RM + 19$$

Example of how to calculate theoretical fragment ions

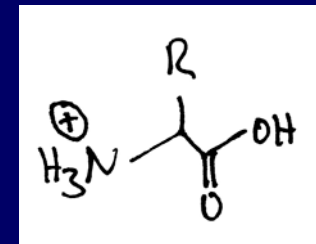
88	159	290	387	500	629	803
S	A	M	P	L	E	R
803	716	645	514	417	304	175



The first b ion



Residue Mass



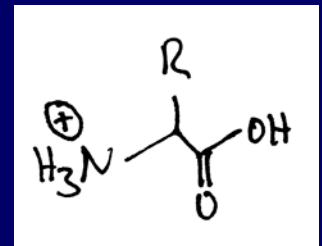
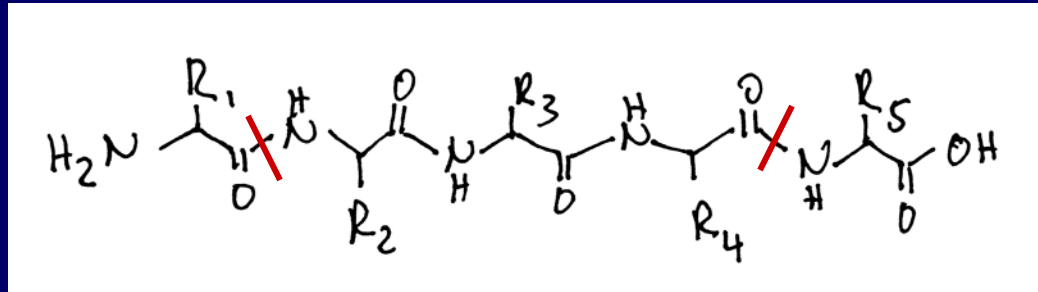
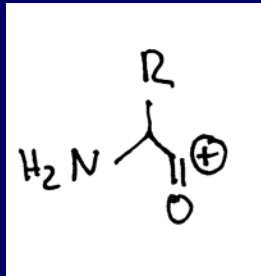
The first y ion

How to calculate theoretical fragment ions

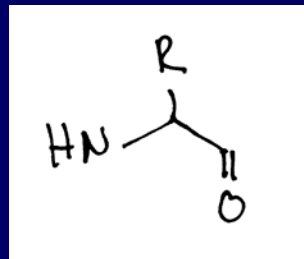
→

RM+1 + RM							+ RM	+ RM	+ RM	+ RM	+ RM	+RM+18
88	159	290	387	500	629	803						
S	A	M	P	L	E	R						
803	716	645	514	417	304	175						
+ RM							+ RM	+ RM	+ RM	+ RM	+ RM	RM+19

←



The first b ion



Residue Mass The first y ion

Finding 'pairs' and 'biggest' ions

$$\begin{aligned} \text{largest b ion} &= (n+H)^+ - (H + \text{HN} \begin{array}{c} \text{R} \\ | \\ \text{C} \\ // \\ \text{O} \end{array} + \text{OH}) \\ &= (n+H)^+ - (Rn + 18) \end{aligned}$$

$$\begin{aligned} \text{largest y ion} &= (n+H)^+ - (\text{HN} \begin{array}{c} \text{R} \\ | \\ \text{C} \\ // \\ \text{O} \end{array}) \\ &= (n+H)^+ - Rn \end{aligned}$$

If trypsin was used for digestion, one can assume that the peptide terminates in K or R. Therefore the biggest observable b ion should be:

Mass of peptide $[M+H]^+ - 128$ (K) - 18

Mass of peptide $[M+H]^+ - 156$ (K) - 18

y ions are truncated peptides. Therefore subtract a residue mass from the parent ion $[M+H]^+$. The highest possible ion could be at $[M+H]^+ - 57$ (G) and the lowest possible ion at $[M+H]^+ - 186$ (W)

b and y ion pairs:

Complementary b and y ions should add up and result in the mass of the intact peptide, except since both b and y ion carry $1H^+$ the peptide mass will be by $1H^+$ too high

therefore: $b (m/z) + y (m/z) - 1 = [M+H]^+$

Check the SAMPLER example

How to start sequencing

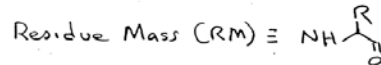
- Know the charge of the peptide
- Know the sample treatment (i.e. alkylation, other derivatizations that could change the mass of amino acids)
- Know what enzyme was used for digestion
- Calculate the $[M+1H]^+$ charge state of the peptide
- Find and exclude non sequence type ions (i.e. unreacted precursor, neutral loss from the parent ion, neutral loss from fragment ions)
- Try to see if you can find the biggest y or b ion in the spectrum. Note, if you used trypsin your C-terminal ion should end in lysine or arginine
- Try to find sequence ions by finding b/y pairs
- You usually can conclude you found the correct sequence if you can explain the major ions in a spectrum

Common observed neutral losses and mass additions:

Table 1. Mass Values for Peptide Fragment Ions

One Let Code	Amino Acid	Tfree Let Code	Mono Isotopic Integer Residue Mass		Neutral Mass Lost		Mono Isotopic Methyl Ester Residue Mass		Average Residue Mass	
			Mass	Change	Mass	Mass	Mass	Mass		
G	Glycine	Gly	57	Da	0	Da			57.02	57.05
A	Alanine	Ala	71	Da	14	Da			71.04	71.08
S	Serine	Ser	87	Da	16	Da	18	Da	87.03	87.08
P	Proline	Pro	97	Da	10	Da			97.05	97.12
V	Valine	Val	99	Da	2	Da			99.07	99.13
T	Threonine	Thr	101	Da	2	Da	18	Da	101.05	101.11
C	Cysteine	Cys	103	Da	2	Da	34	Da	103.01	103.14
L	Leucine	Leu	113	Da	10	Da			113.08	113.16
I	Isoleucine	Ile	113	Da					113.08	113.16
N	Asparagine	Asn	114	Da	1	Da	17	Da	114.04	114.1
D	Aspartic Acid	Asp	115	Da	1	Da	18	Da	115.03	115.09
K	Lysine	Lys	128	Da	13	Da	17	Da	128.09	128.17
Q	Glutamine	Gln	128	Da			17	Da	128.06	128.13
E	Glutamic Acid	Glu	129	Da	1	Da	18	Da	129.04	129.12
M	Methionine	Met	131	Da	2	Da	48	Da	131.04	131.19
H	Histidine	His	137	Da	6	Da			137.06	137.14
F	Phenylalanine	Phe	147	Da	10	Da			147.07	147.18
R	Arginine	Arg	156	Da	9	Da	17	Da	156.1	156.19
Y	Tyrosine	Tyr	163	Da	7	Da			163.06	163.18
W	Tryptophan	Trp	186	Da	23	Da			186.08	186.21

Hydrogen	H	1	1.008	1.008
Carbon	C	12	12	12.011
Nitrogen	N	14	14.003	14.007
Oxygen	O	16	15.995	15.999
Sulfur	S	32	31.972	32.06
Methyl		15	15.02	15.04
Water		18	18.01	18.02
Acetyl		43	43	43.05



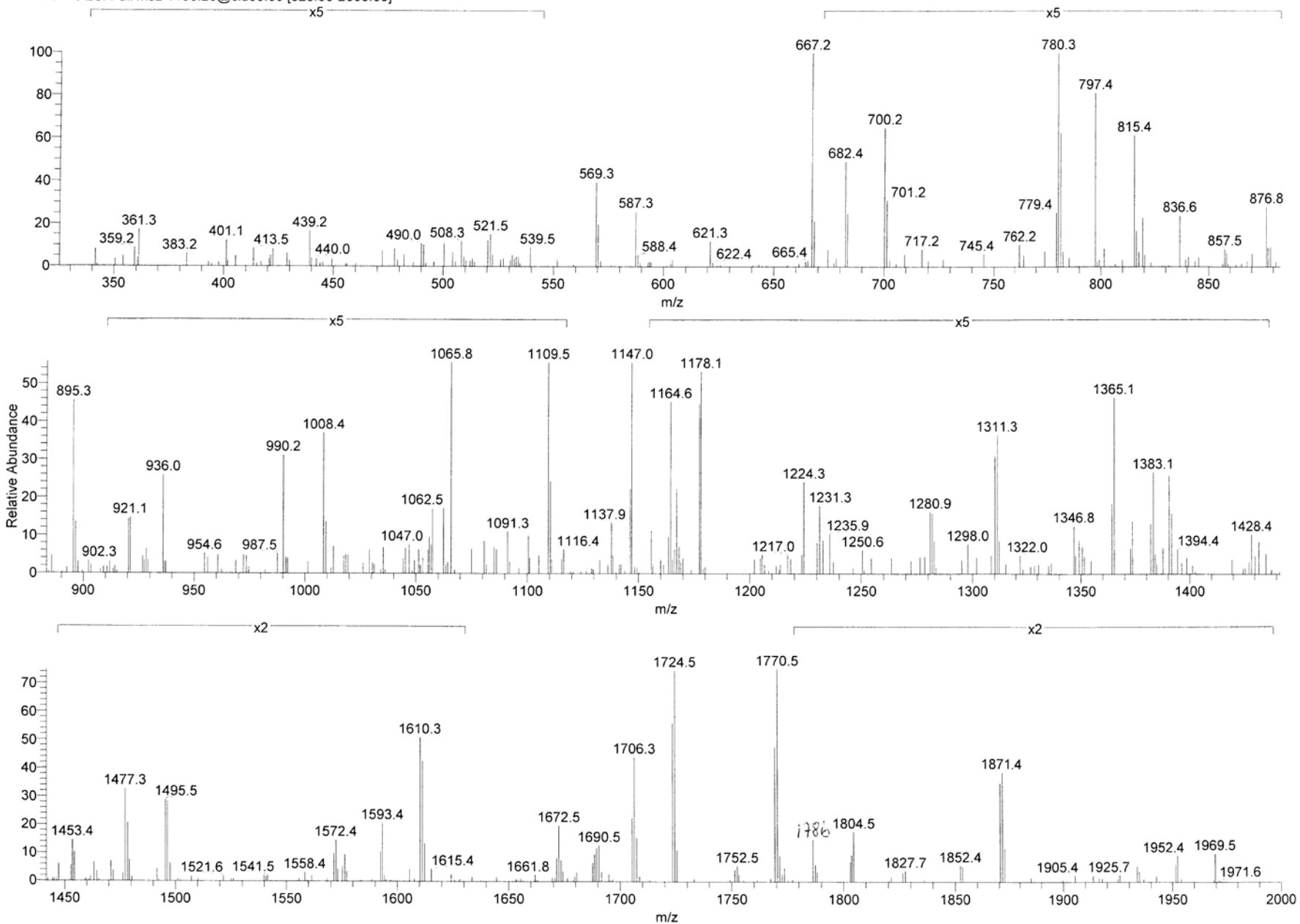
- Ammonia -17
- Water -18
- Carbon Monoxide from b ions -28
- Phosphoric acid from phosphorylated serine and threonine -98
- Carbamidomethyl modification on cysteines upon alkylation with iodoacetamide +57
- Oxidation of methionine +18

Calculate with nominal mass during sequencing, but use the monoisotopic masses to check if the parent mass fits. For high res. MS/MS check that the residue mass difference is correct.

loaded 3 ul (total ~12ul) of gel digested Stat3

Stat3_cytosolic_a #8004 RT: 63.24 AV: 1 NL: 1.15E3

T: ITMS + c ESI Full ms2 1196.20@cid35.00 [325.00-2000.00]

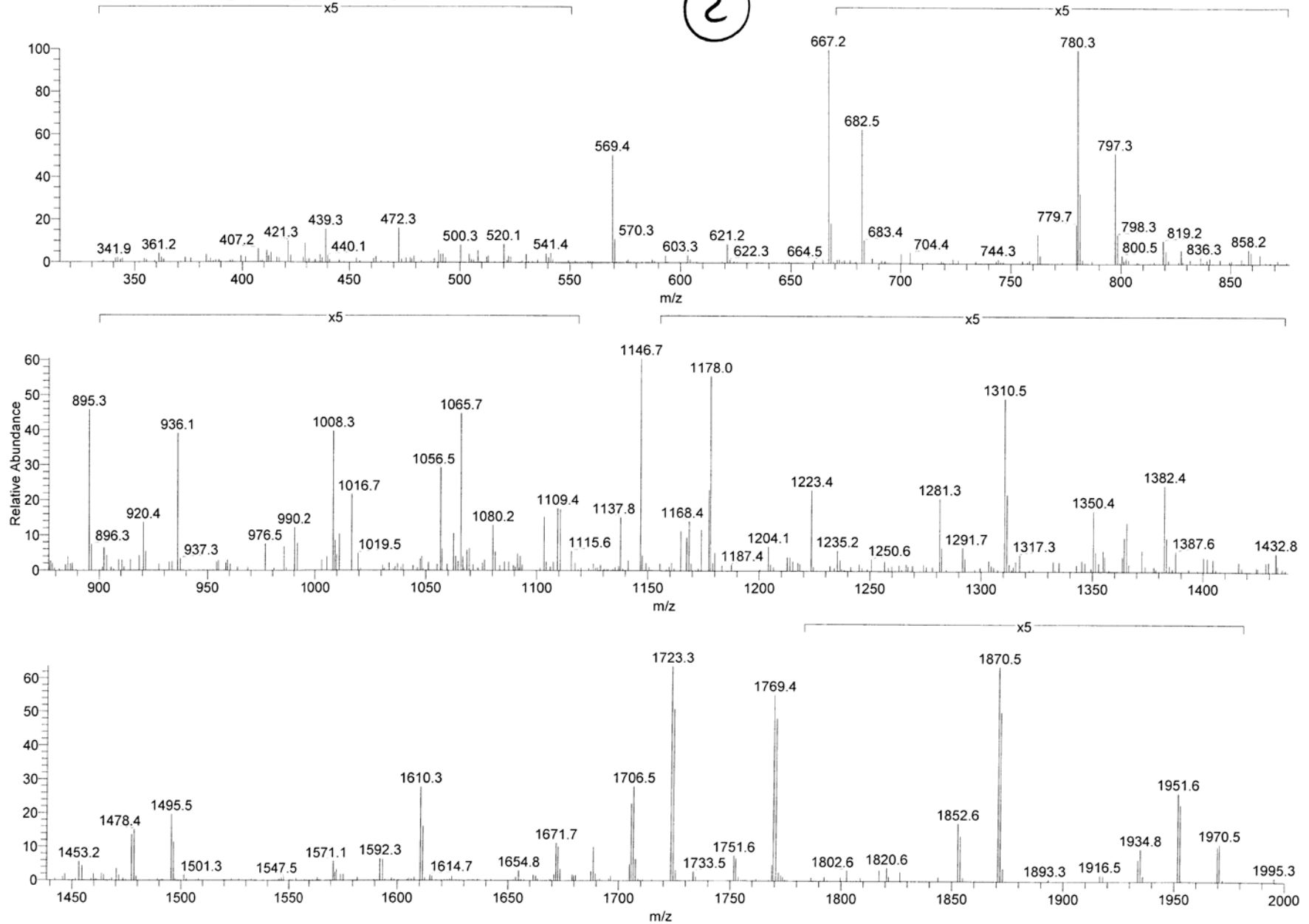


loaded 3 ul (total ~12ul) of gel digested Stat3

Stat3_cytosolic_a #7952 RT: 62.88 AV: 1 NL: 6.00E3

T: ITMS + c ESI d Full ms2 1196.04@cid35.00 [315.00-2000.00]

2



Mixed Phospho spectra

<http://prospector.ucsf.edu>

FIC(Carbamidomethyl)VTPTTC(Carbamidomethyl)SNTIDLPMSPR

User AA Formula 1: **C2 H3 N1 O1**
 Elemental Composition: C98 H161 N26 O32 S3

$[MH^+]$	$[MH^+(mono)]$	$[MH^2]$	$[MH^2(mono)]$
2311.3894	2110.0927	1156.1984	1155.5500

[-] Main Sequence Ions

k	—	1	F	20	—	y	y ²
2011598	2	I	19	2242.9906	1121.9989		
4211904	3	C(Carbamidomethyl)	18	2287.9802	1023.4737		
5202388	4	V	17	1809.8096	945.4384		
6213065	5	T	16	1740.8411	895.3542		
7183310	6	P	15	1689.7935	845.4404		
8194469	7	T	14	1592.7867	796.8780		
9204546	8	T	13	1491.6930	746.3302		
10004853	9	C(Carbamidomethyl)	12	1396.6453	695.8203		
11075173	10	S	11	1296.6147	615.8100		
12015602	11	N	10	1143.5827	572.2390		
13024079	12	T	9	1024.5307	515.2795		
14094920	13	I	8	928.4921	464.7697		
16107189	14	D	7	815.4080	408.2076		
17238030	15	L	6	700.3813	350.6942		
18208557	16	P	5	587.2970	294.1321		
1918862	17	M	4	490.2442	245.6238		
20084292	18	S	3	359.2037	186.3095		
21335809	19	P	2	272.1717	136.5895		
—	20	R	1	175.1390	88.0631		

[-] All Sequence Ions

unmodified

<http://prospector.ucsf.edu>

FIC(Carbamidomethyl)VTPTTC(Carbamidomethyl)SNTIDLPMSPR

User AA Formula 1: **C2 H3 N1 O1**
 Elemental Composition: C98 H162 N26 O35 S3 P1

$[MH^+]$	$[MH^+(mono)]$	$[MH^2]$	$[MH^2(mono)]$
2391.3693	2390.0590	1196.1884	1195.5331

[-] Main Sequence Ions

k	—	1	F	20	—	y	y ²
2011598	2	I	19	2242.9906	1121.9989		
4211904	3	C(Carbamidomethyl)	18	2129.9605	1065.4369		
5202388	4	V	17	1760.8739	983.4116		
7012728	5	T	16	1678.8675	933.9678		
7983236	6	P	15	1609.7935	845.4384		
8993733	7	T	14	1592.7867	796.8780		
10004209	8	T	13	1491.6930	746.3302		
11044936	9	C(Carbamidomethyl)	12	1396.6453	695.8203		
12574830	10	S	11	1230.6147	615.8100		
13615206	11	N	10	1143.5827	572.2390		
14625742	12	T	9	1024.5307	515.2795		
16786683	13	I	8	928.4921	464.7697		
18989832	14	D	7	815.4080	408.2076		
18033039	15	L	6	700.3811	350.6942		
19408221	16	P	5	587.2970	294.1321		
20918625	17	M	4	490.2442	245.6238		
21188946	18	S	3	359.2037	186.3095		
22181473	19	P	2	272.1717	136.5895		
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1 Phospho site

<http://prospector.ucsf.edu>

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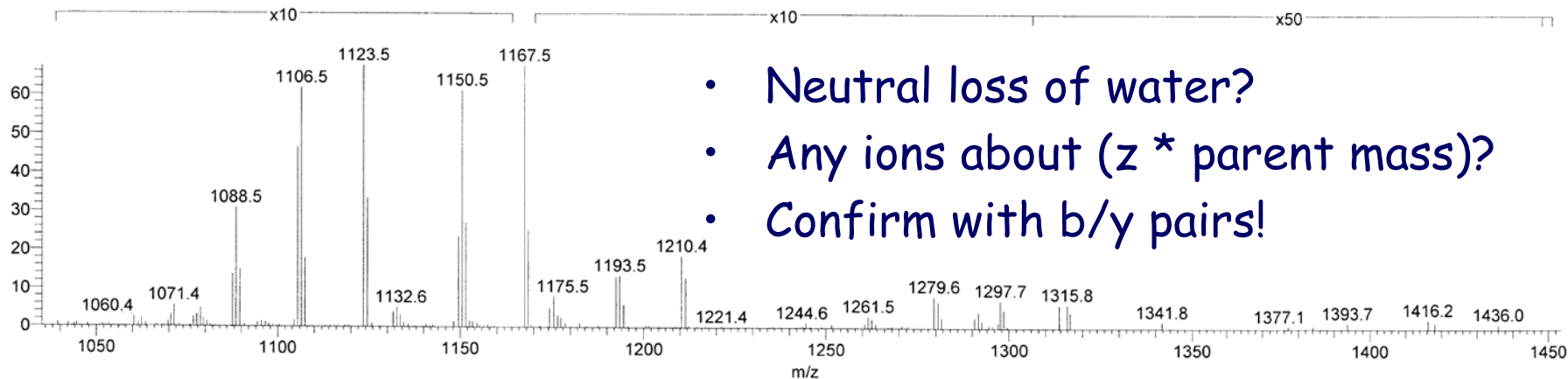
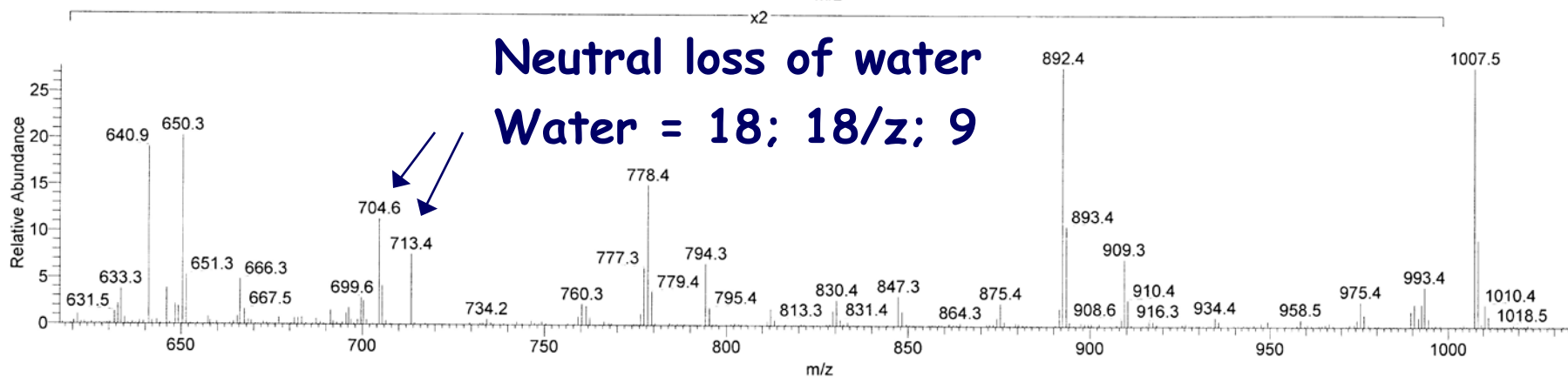
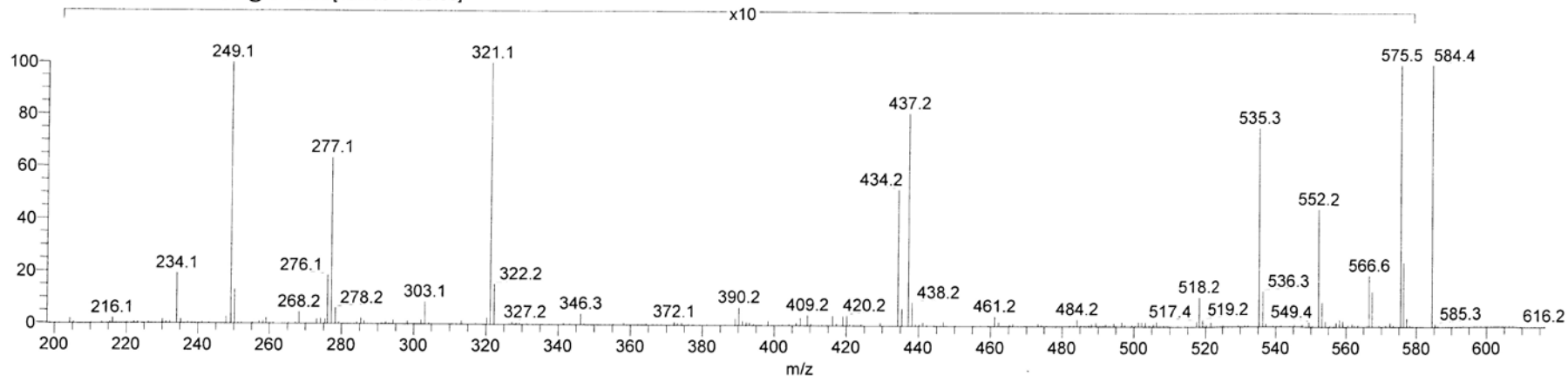
[-] All Sequence Ions

1 Phospho site

First 'on your own example'

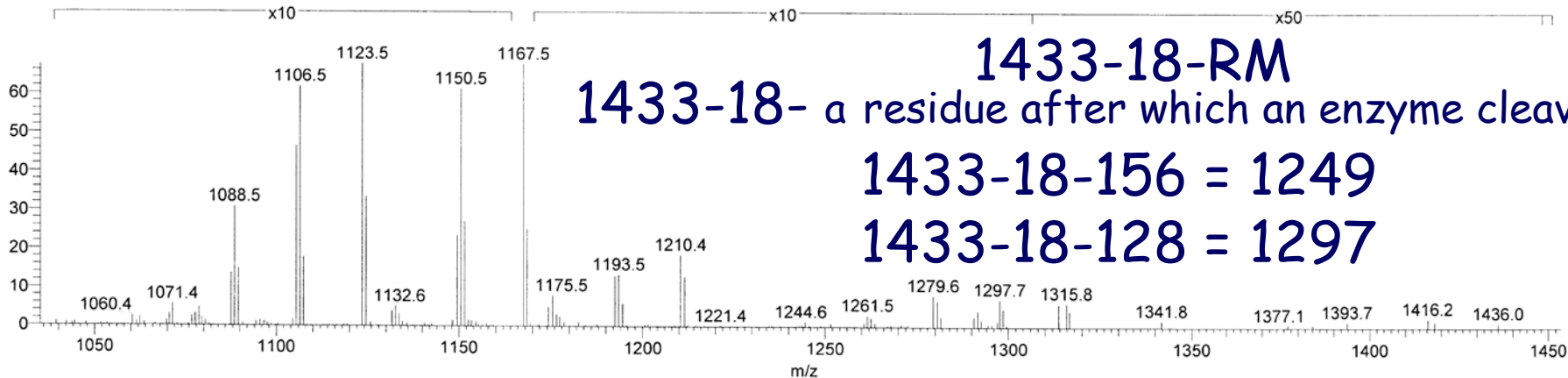
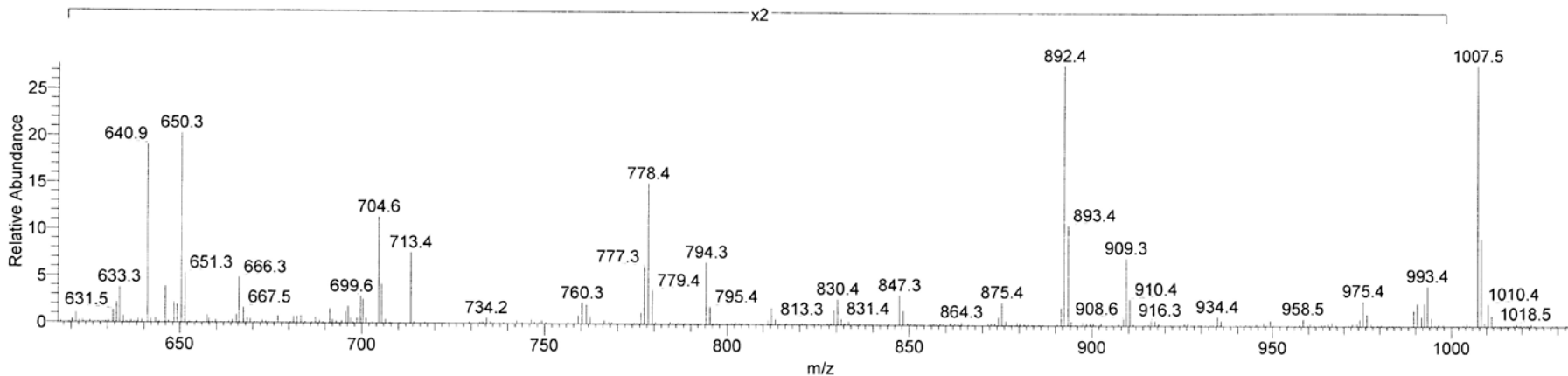
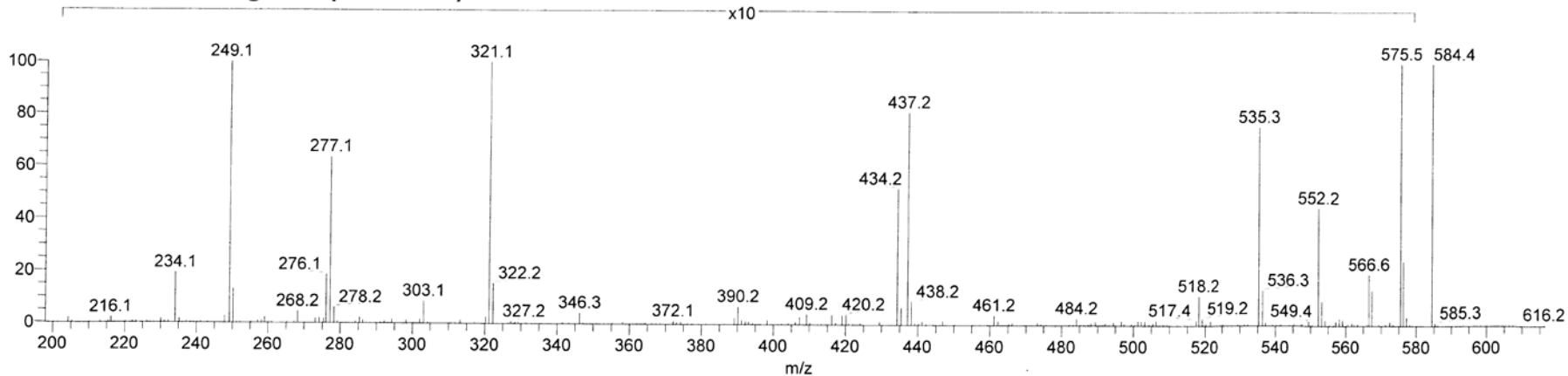
Remember what you need to know first!

What is the charge state?

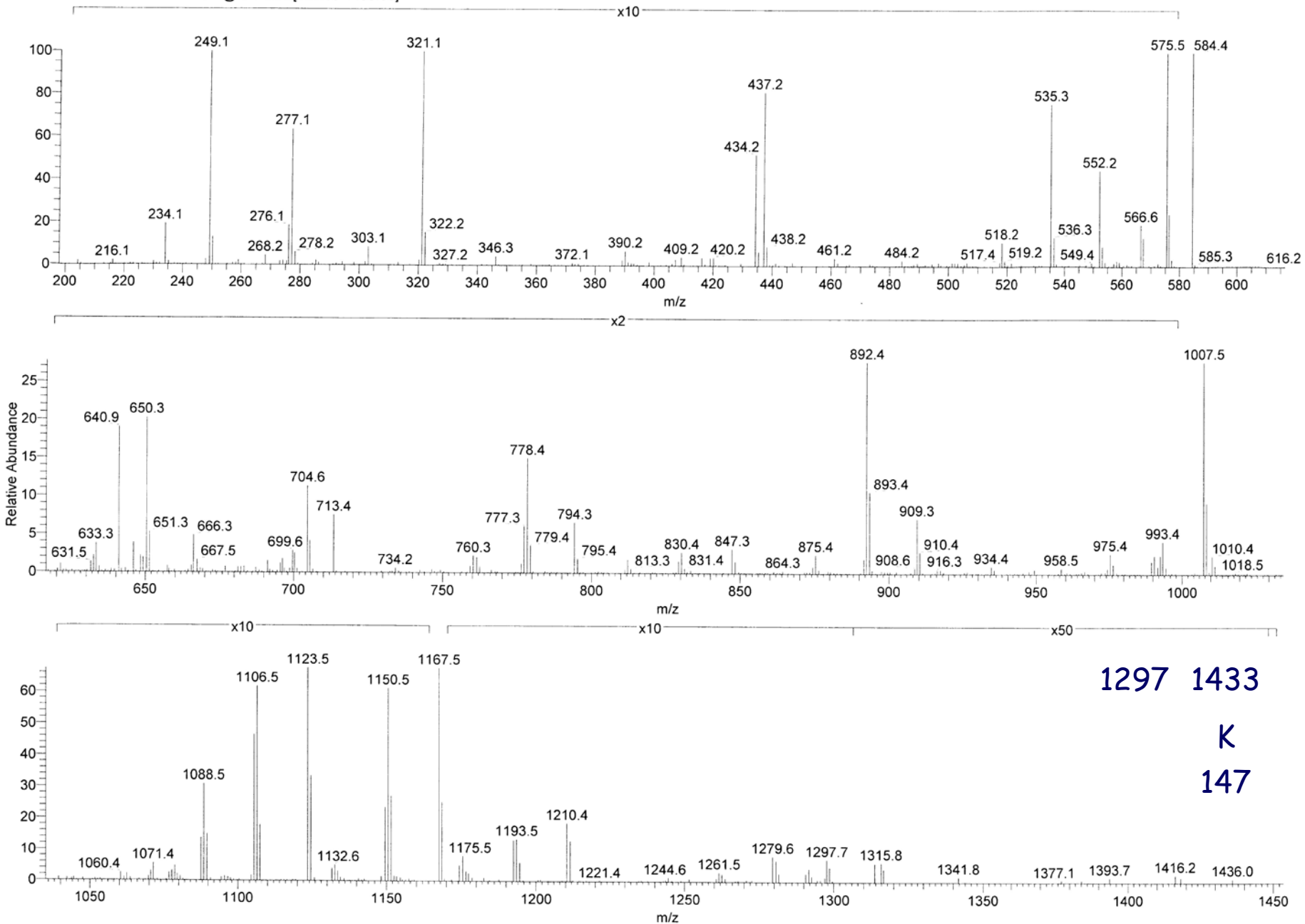


- Neutral loss of water?
- Any ions about ($z \times \text{parent mass}$)?
- Confirm with b/y pairs!

Search for 'biggest ion'



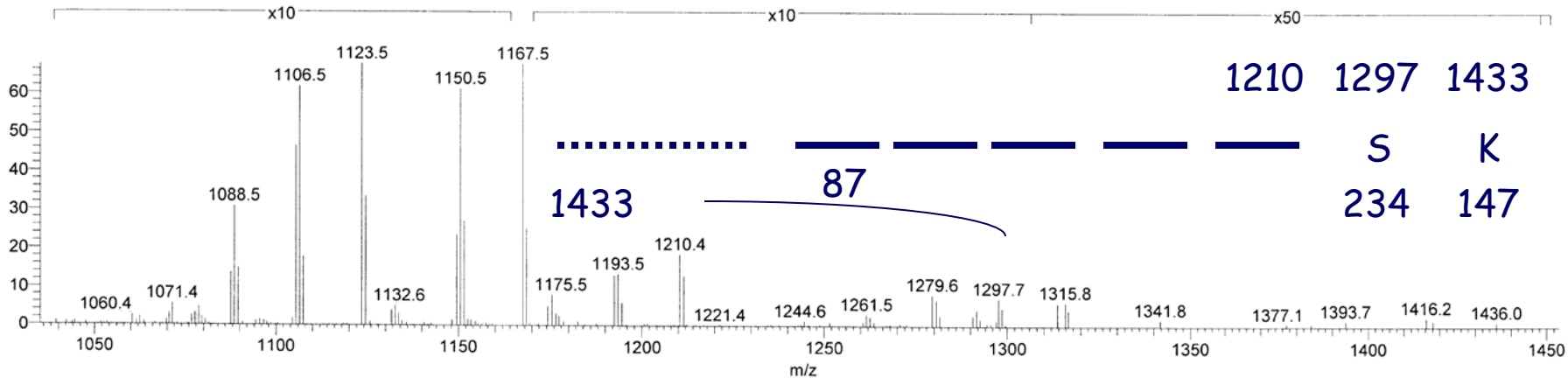
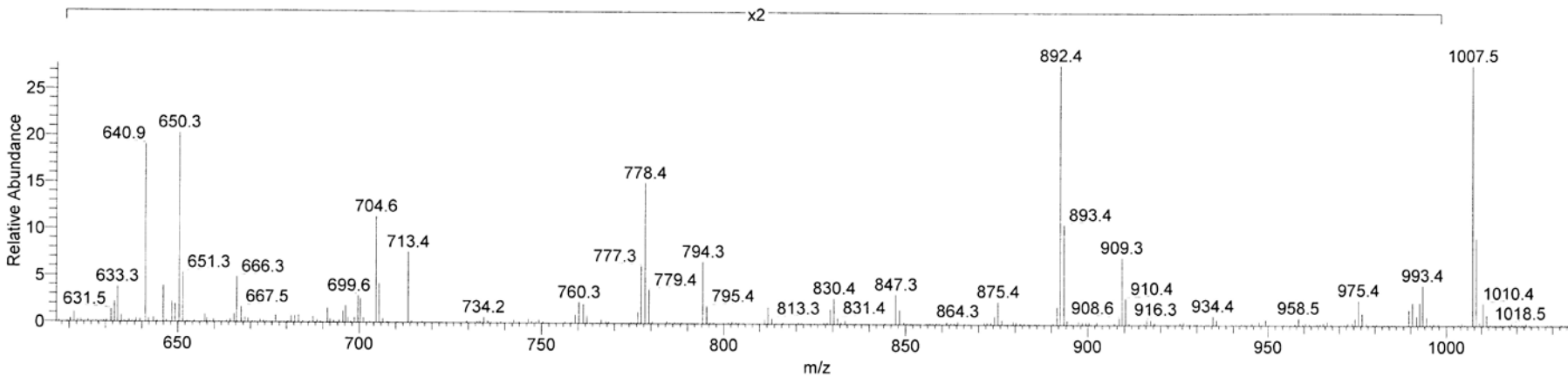
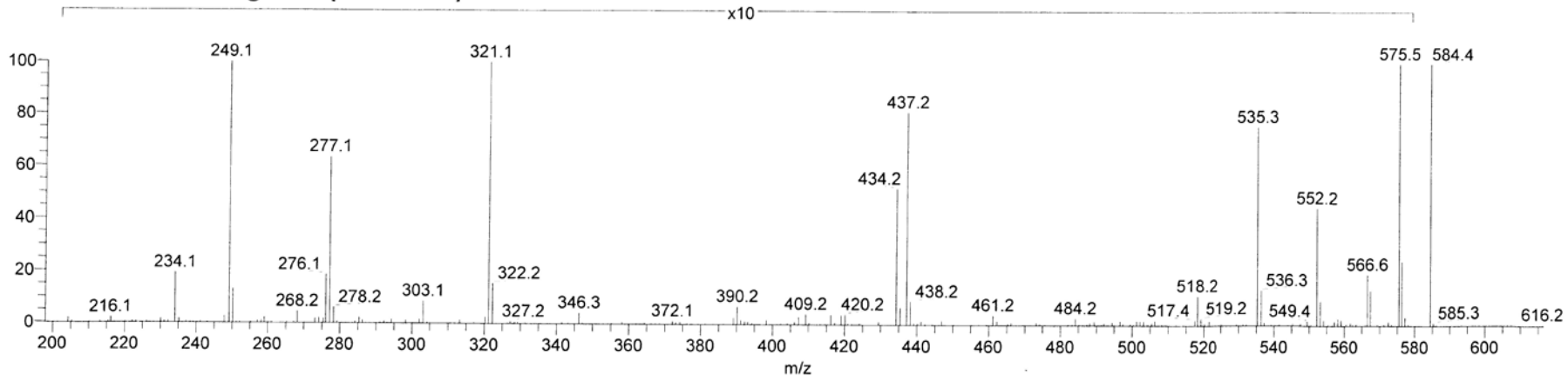
1433-18-RM
1433-18- a residue after which an enzyme cleaves
1433-18-156 = 1249
1433-18-128 = 1297



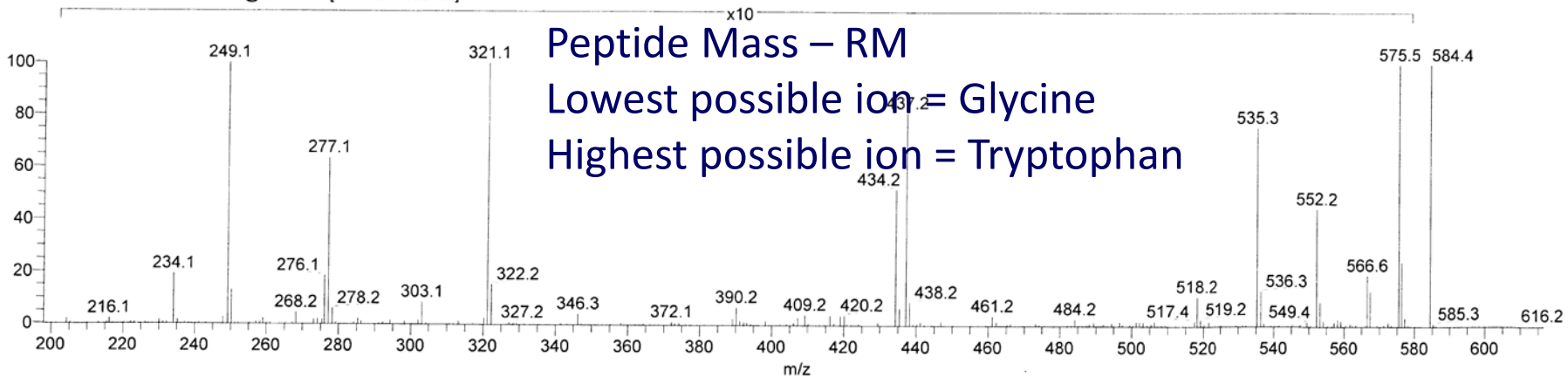
1297 1433

K

147



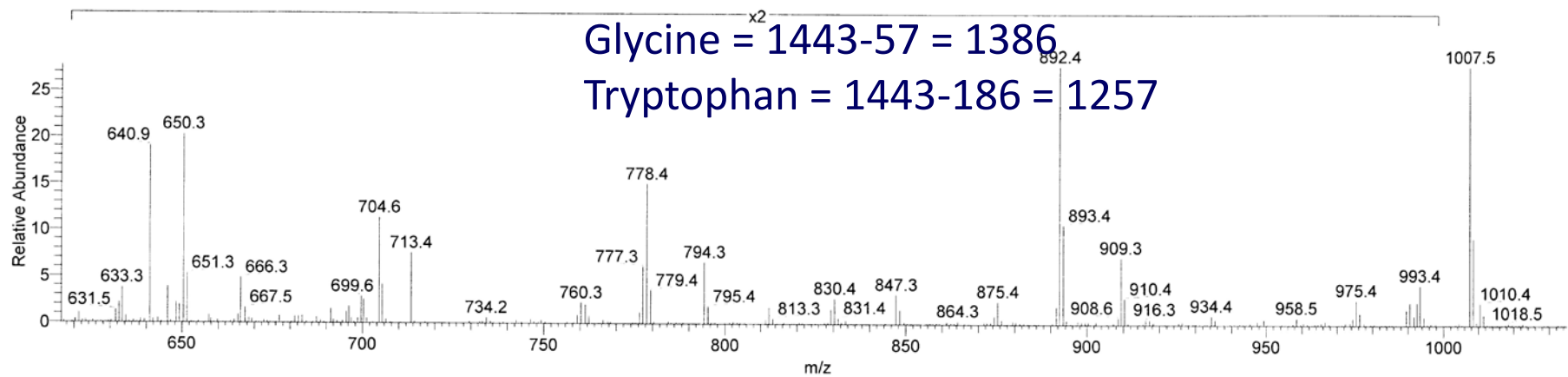
Find the biggest y ion!



Peptide Mass – RM

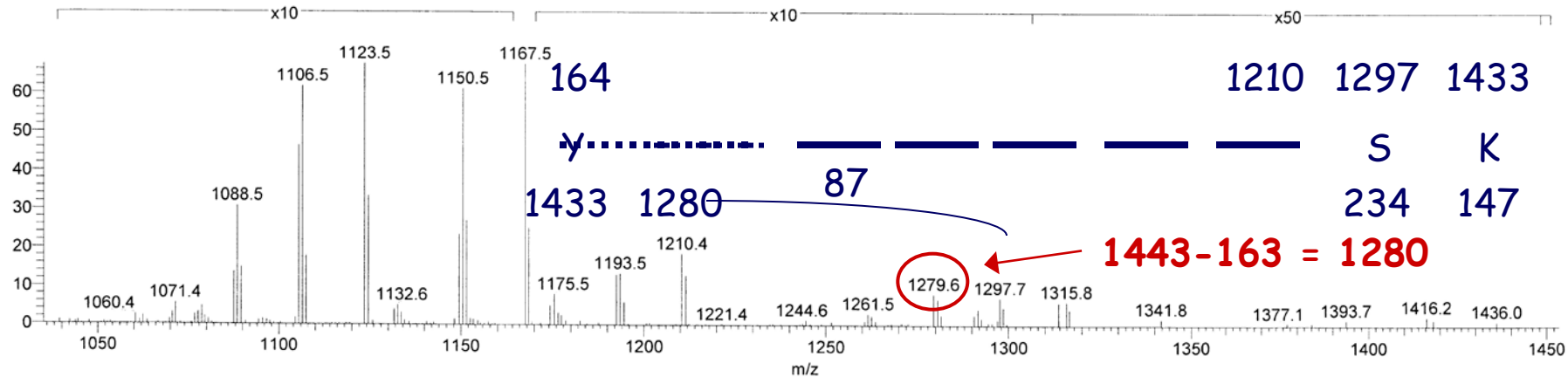
Lowest possible ion = Glycine

Highest possible ion = Tryptophan

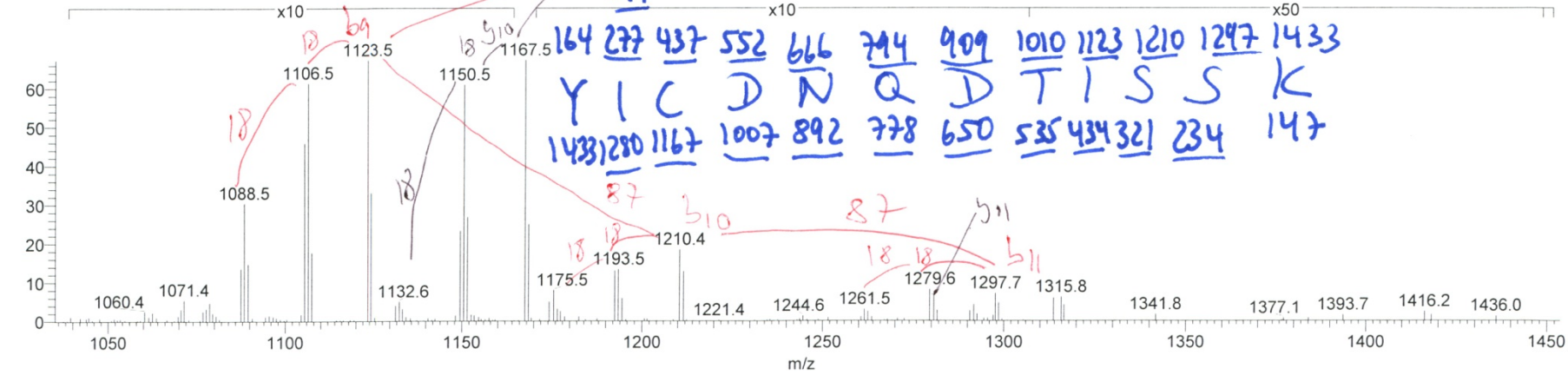
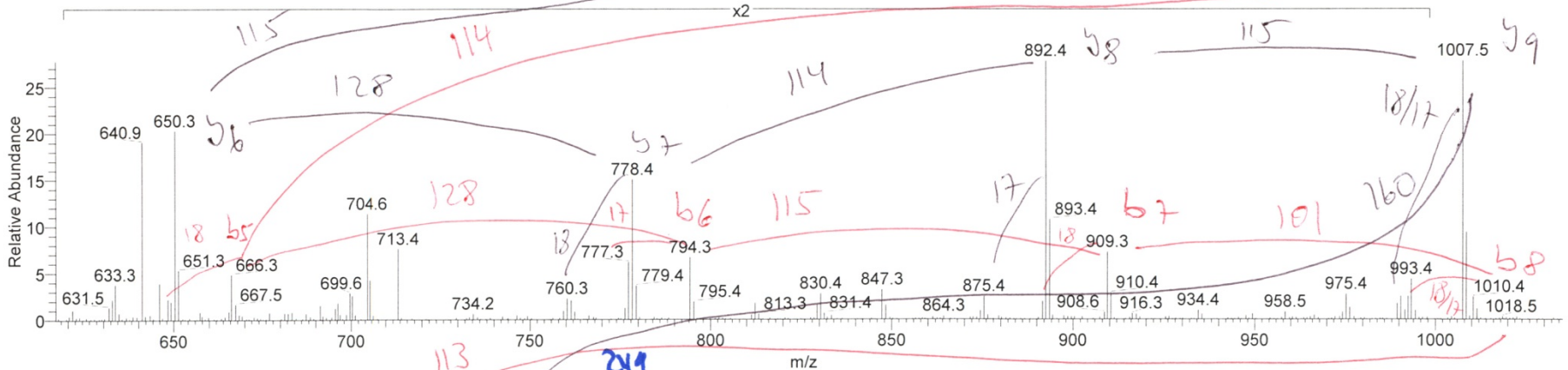
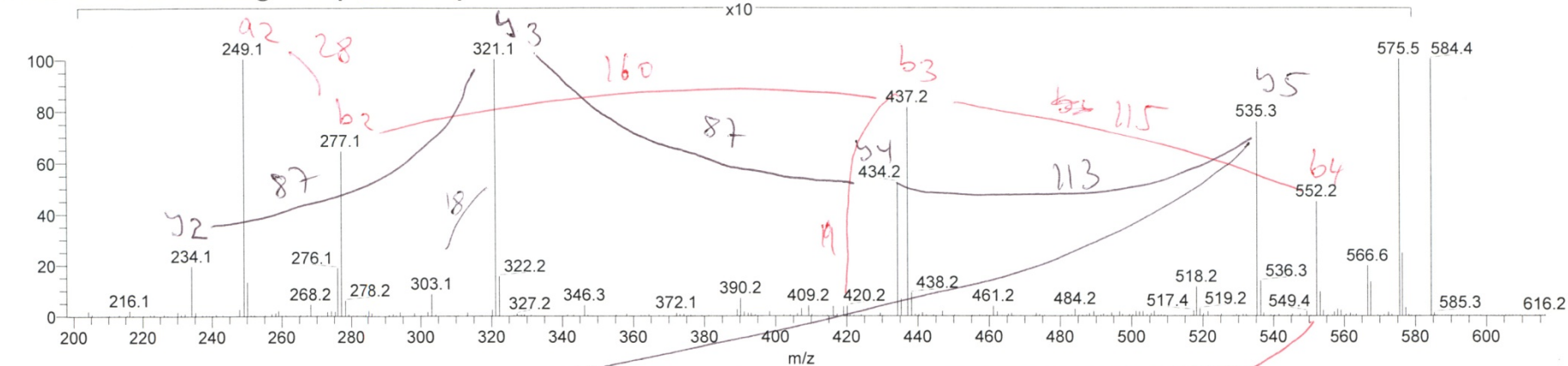


Glycine = $1443 - 57 = 1386$

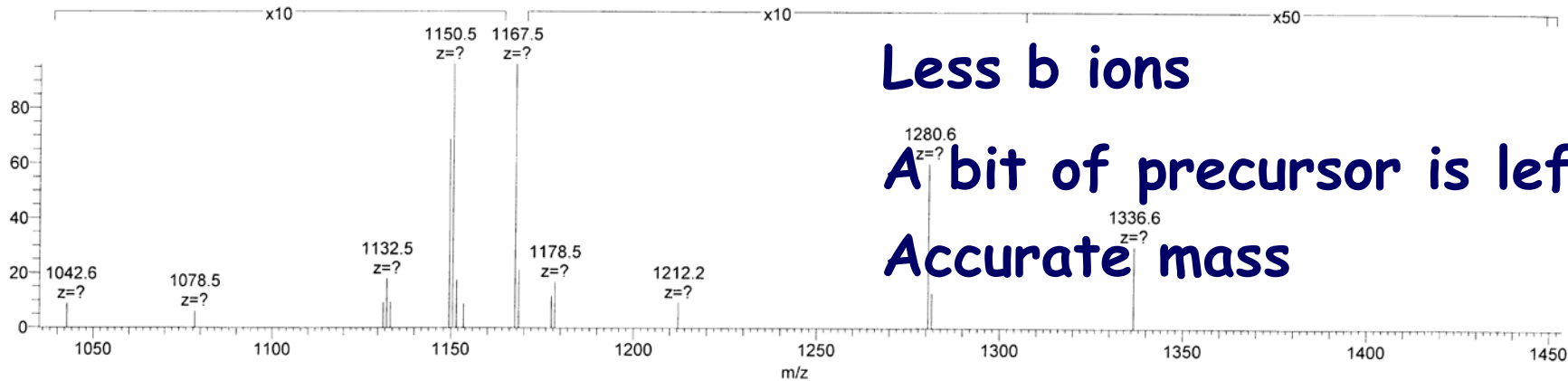
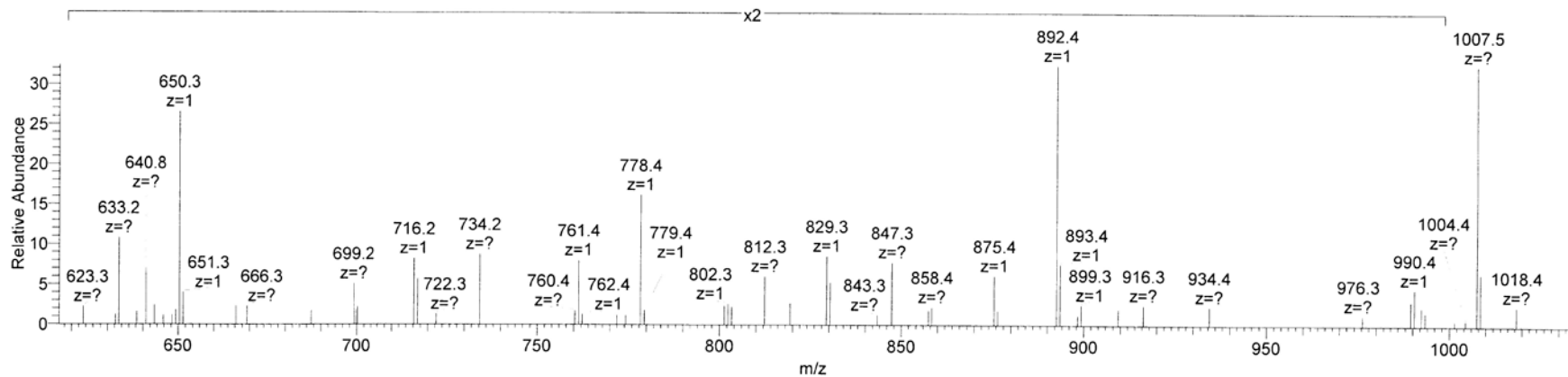
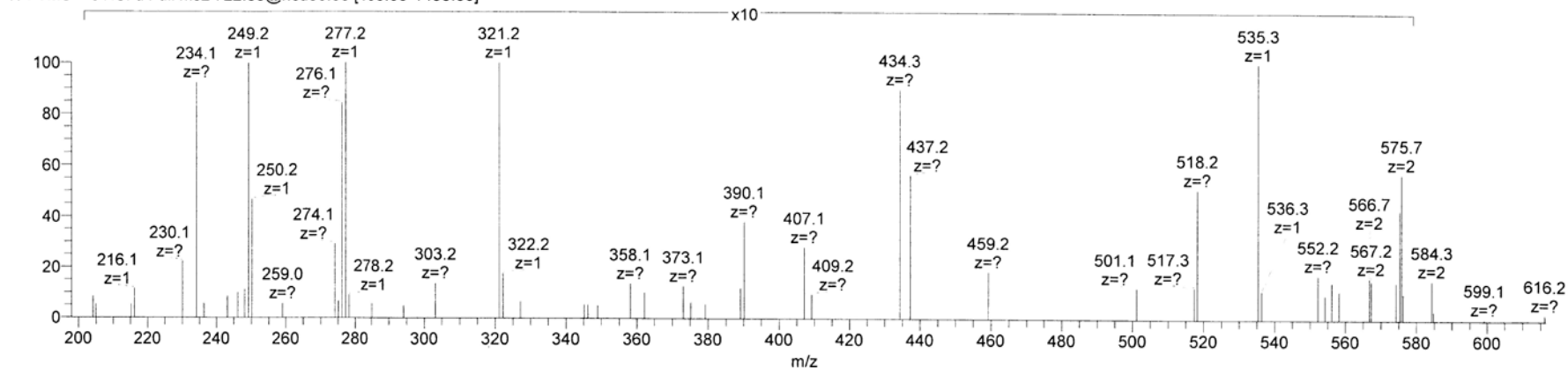
Tryptophan = $1443 - 186 = 1257$



And the sequence is.....



What is the difference ?

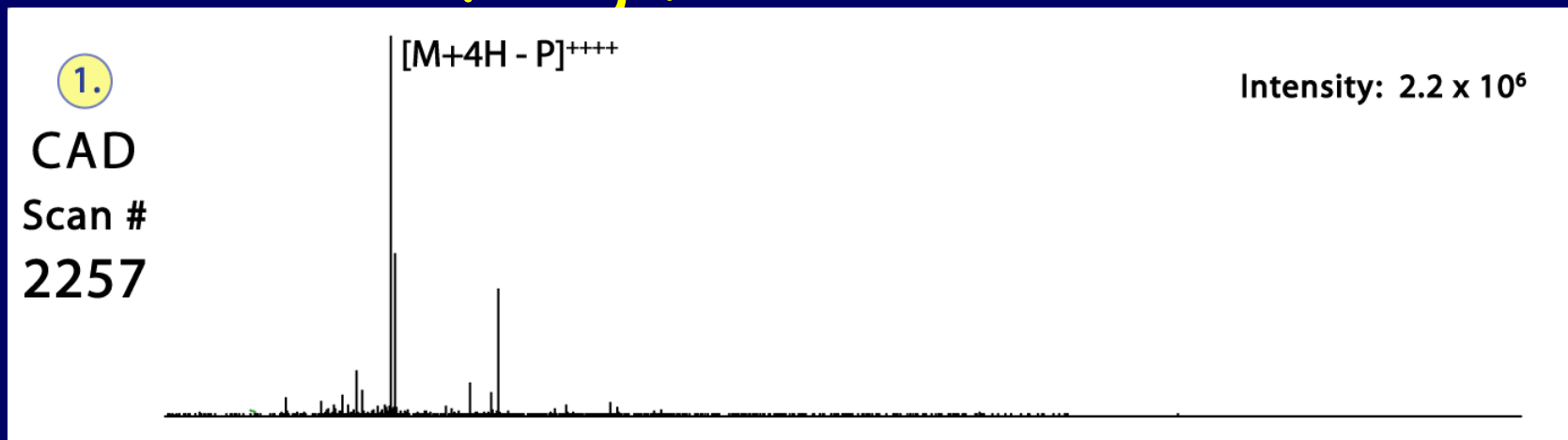


Less b ions

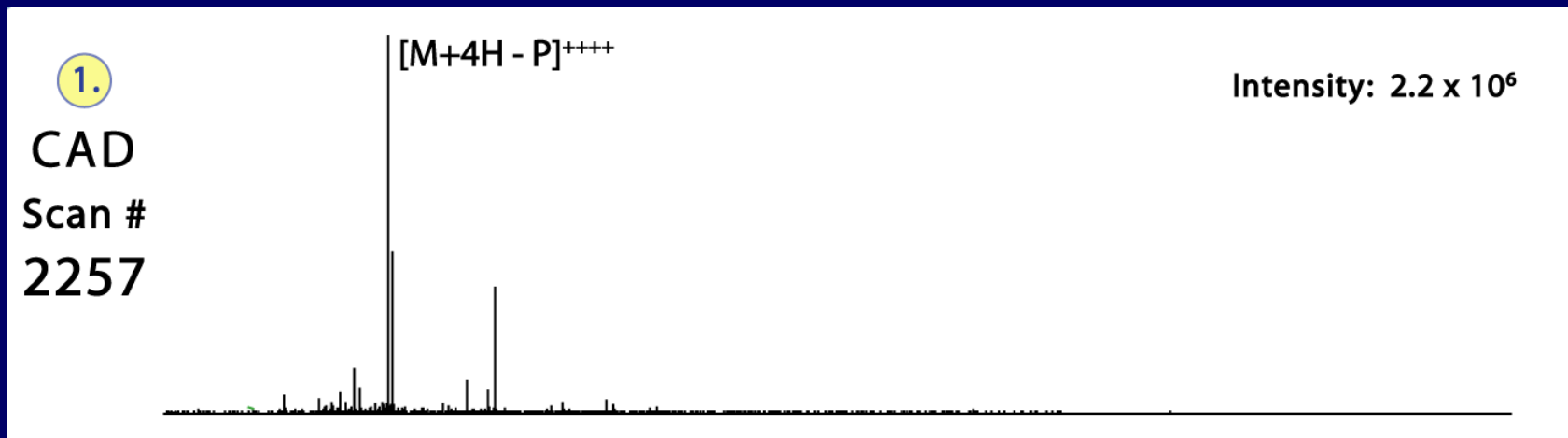
A bit of precursor is left

Accurate mass

What if we do not get good fragmentation?



Try a different mode of dissociation



ETD

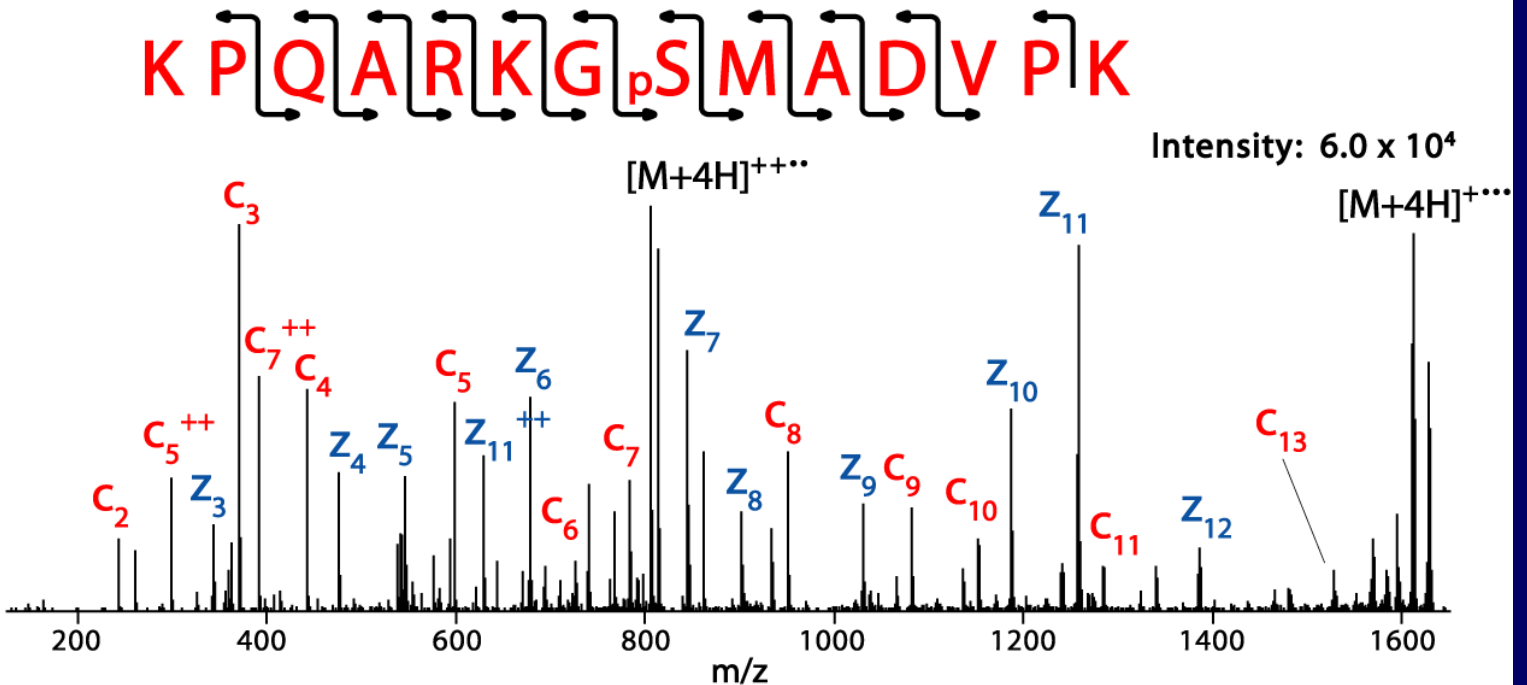
1.

CAD
Scan #
2257

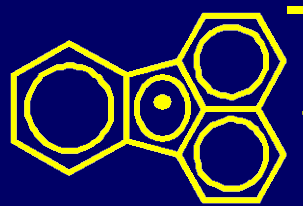


2.

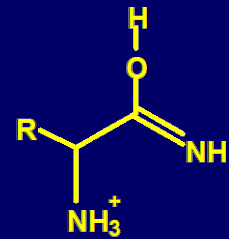
ETD
Scan #
2258



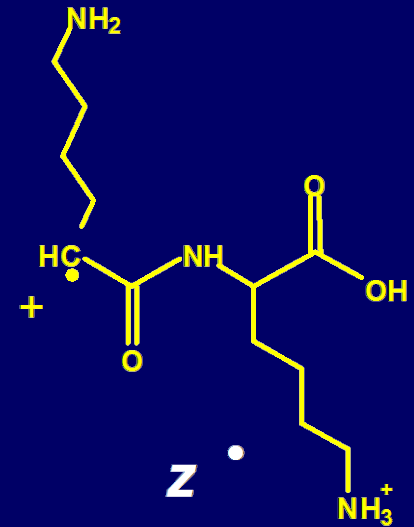
Electron Transfer Dissociation



+ peptide⁺

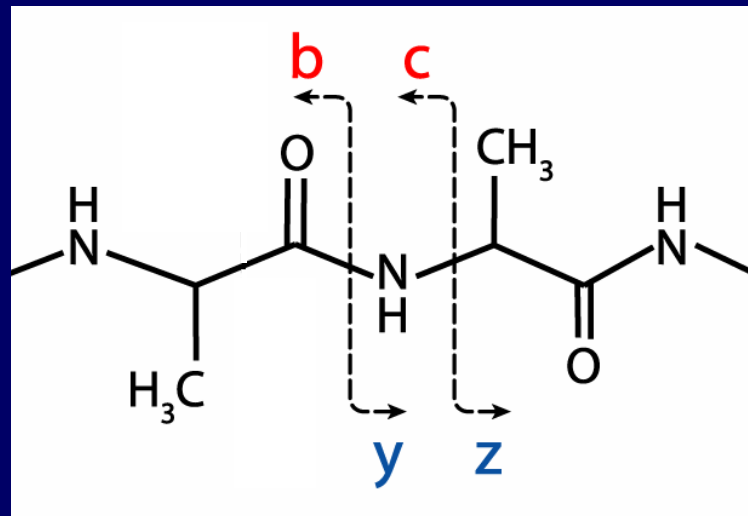


c



z

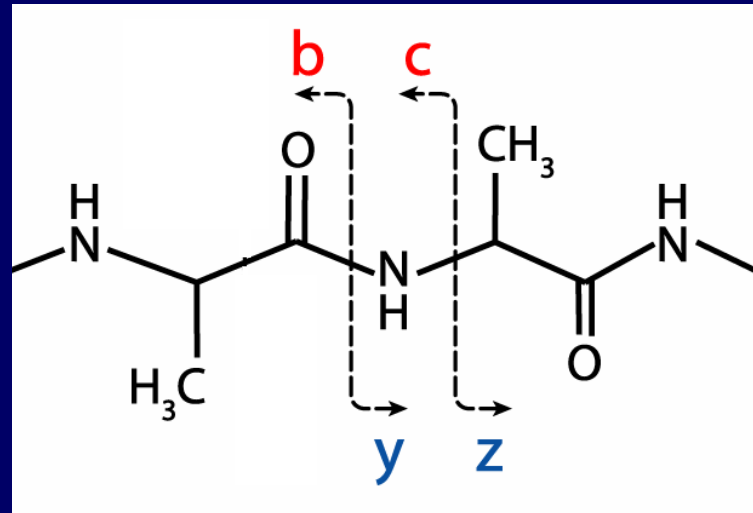
Fluoranthene



Tandem MS - Dissociation Techniques

CAD: Collision Activated Dissociation (b, y ions)

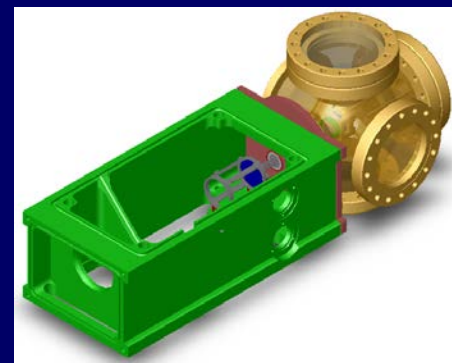
⇒ increase of internal energy through collisions



ETD: Electron Transfer Dissociation (c, z ions)

⇒ bombardment of peptides with electrons
(radical driven fragmentation)

The Prototype Instrument

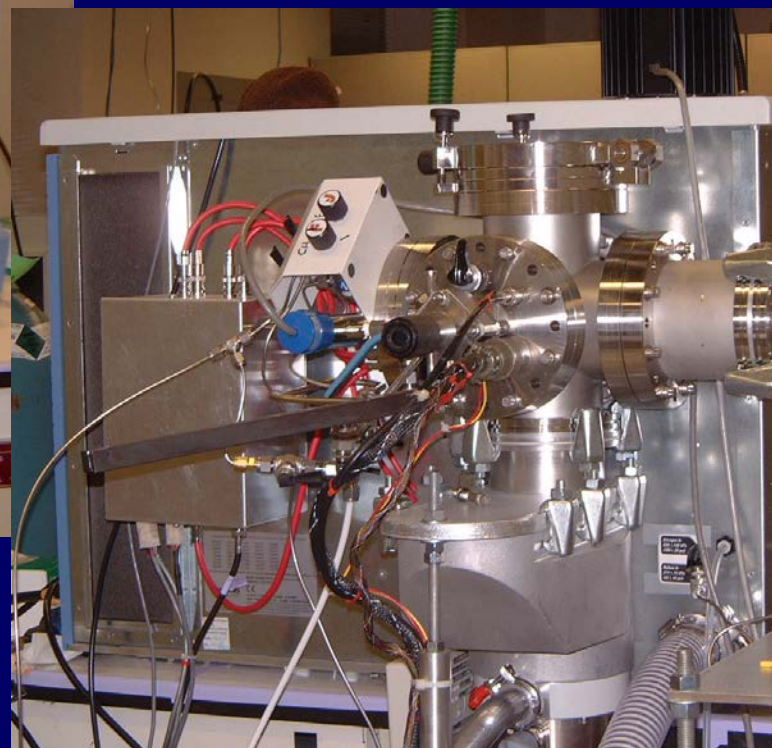


Modified rear / CI source

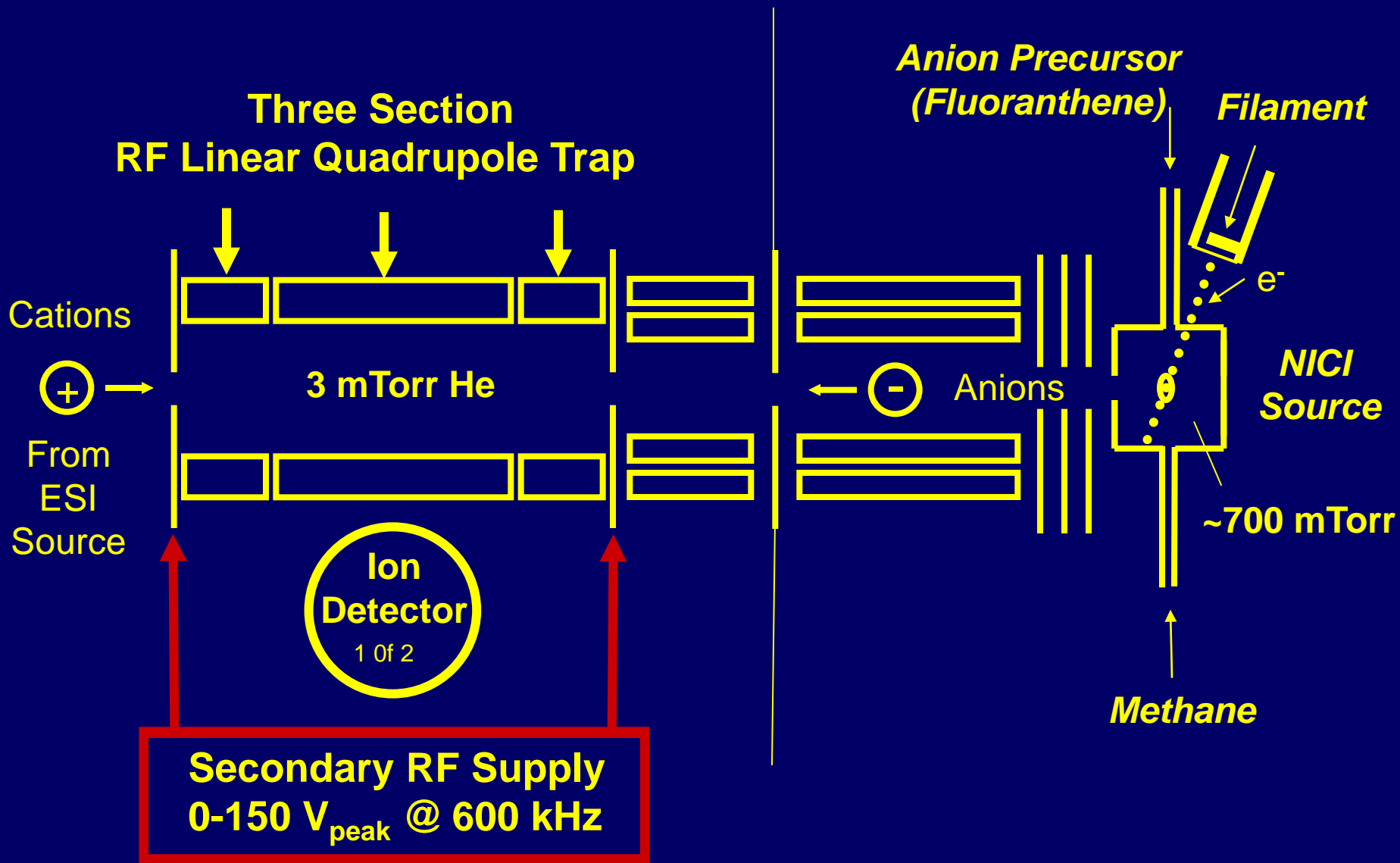


HPLC

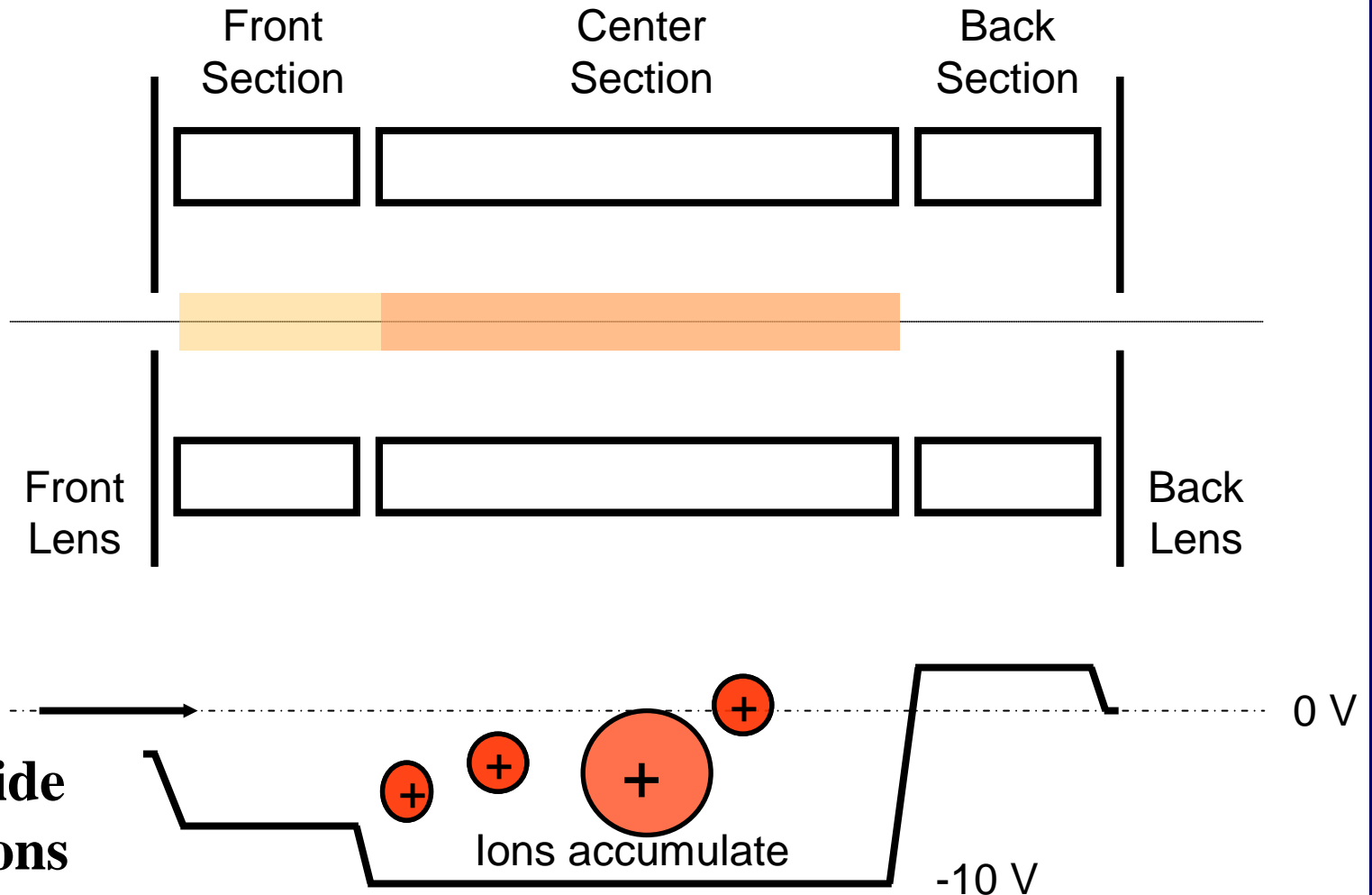
LTQ front



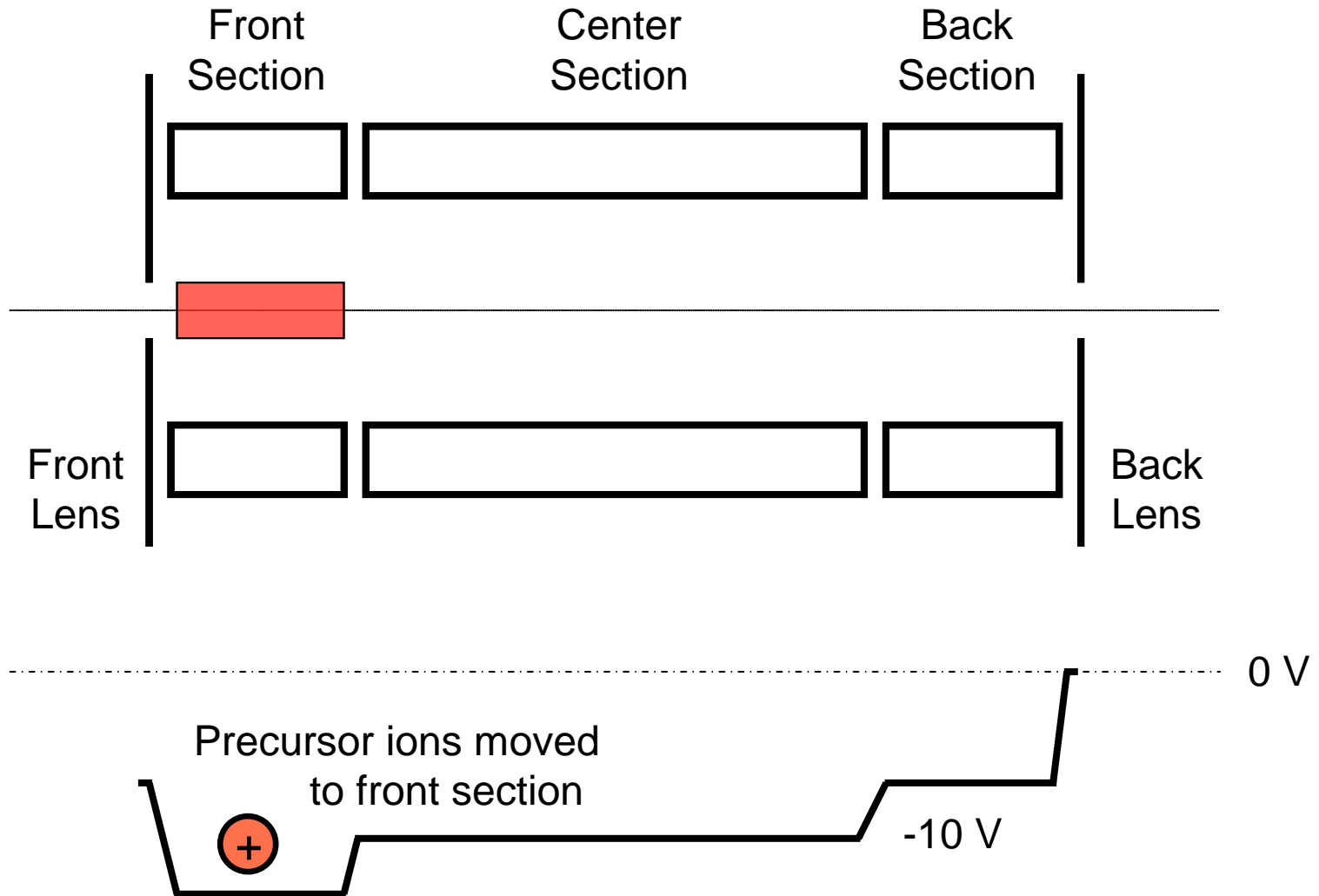
Modifications For Ion/Ion Experiments



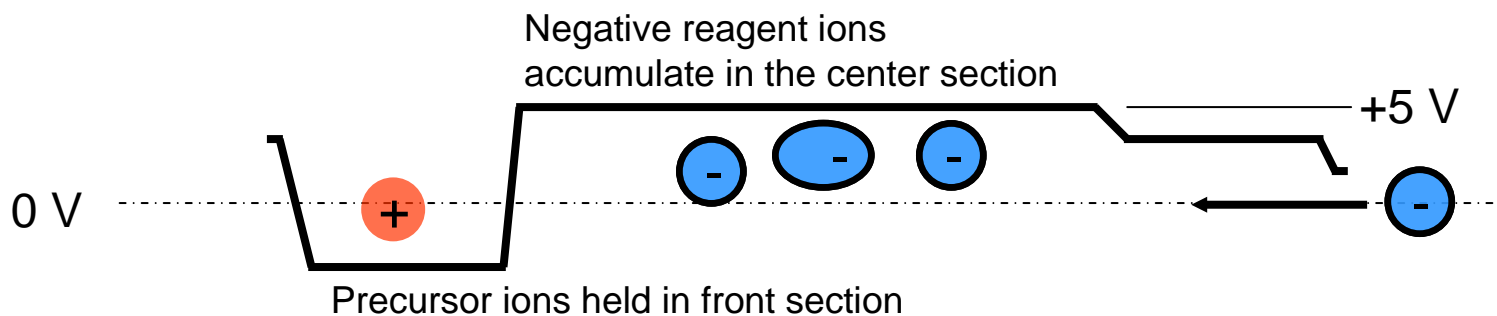
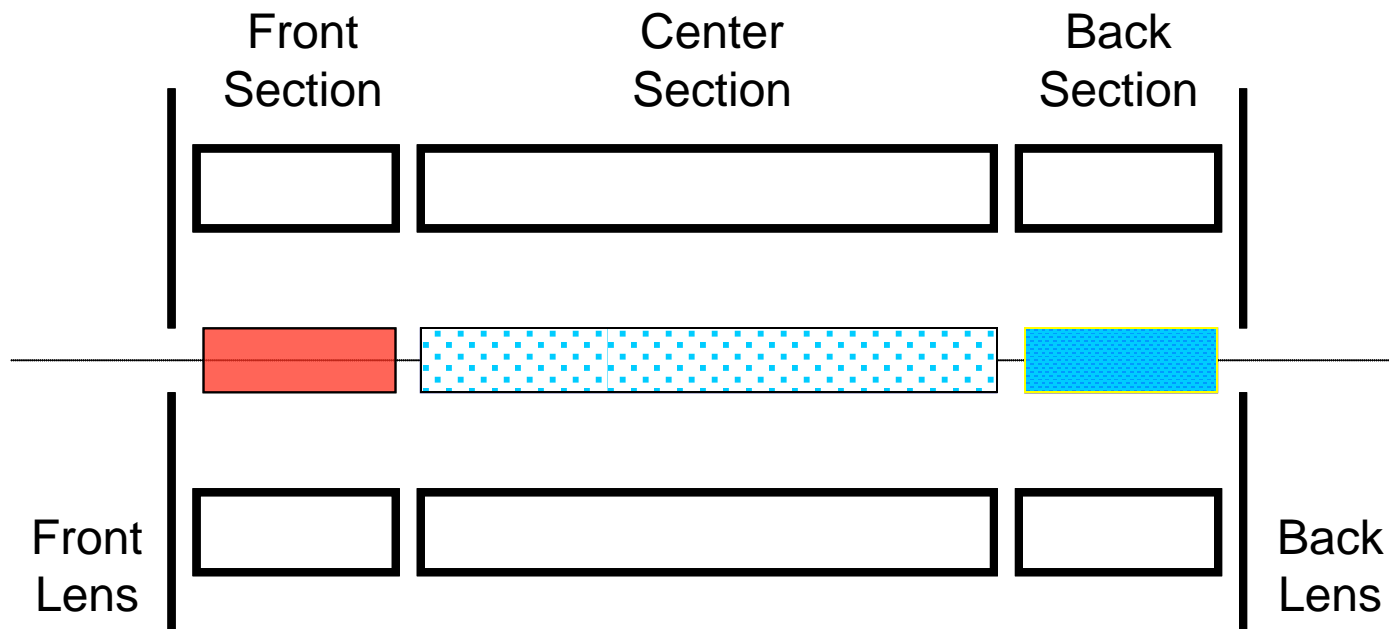
Injection of Positive Ions (ESI)



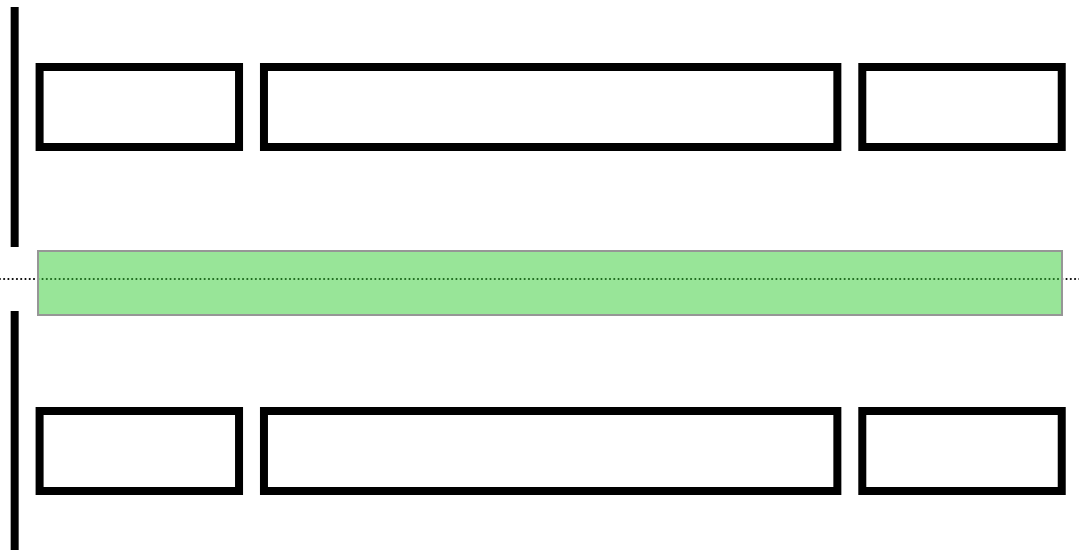
Precursor Storage in Front Section



Injection of Negative Ions (CI)

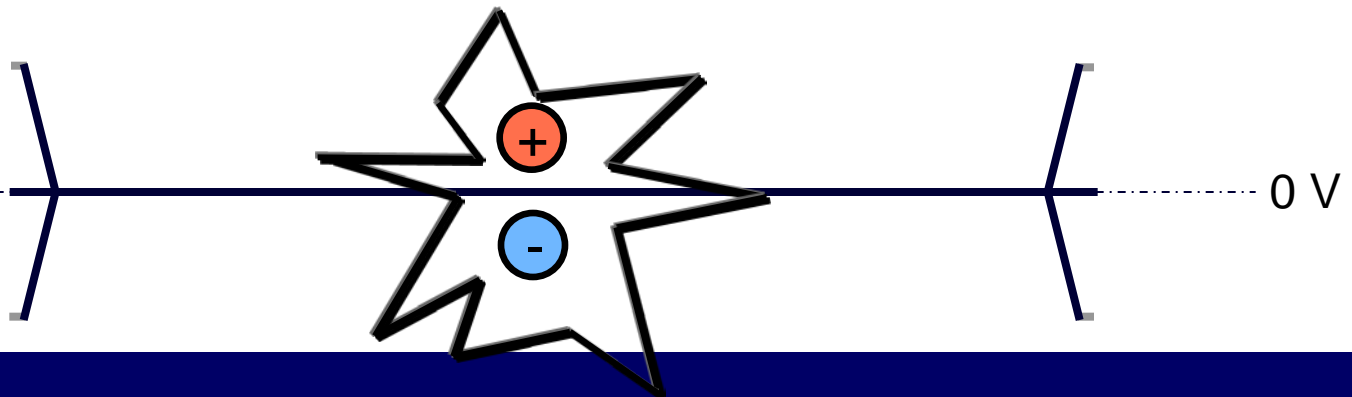


Charge-Sign Independent Trapping

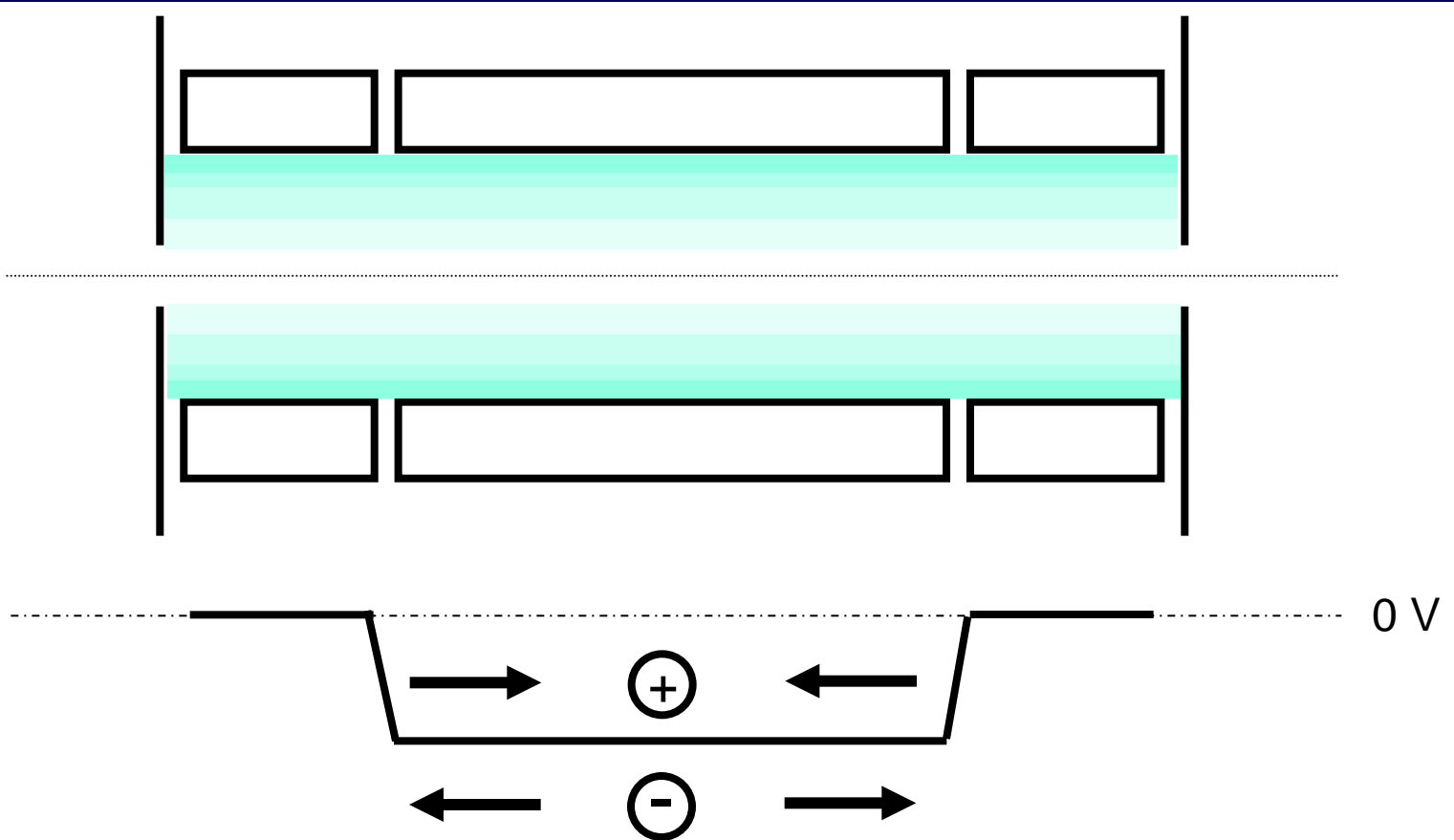


Positive and negative ions react while trapped in axial pseudo-potential

Pseudo-potential created by $+150 \text{ V}_p$ 600 kHz applied to lenses

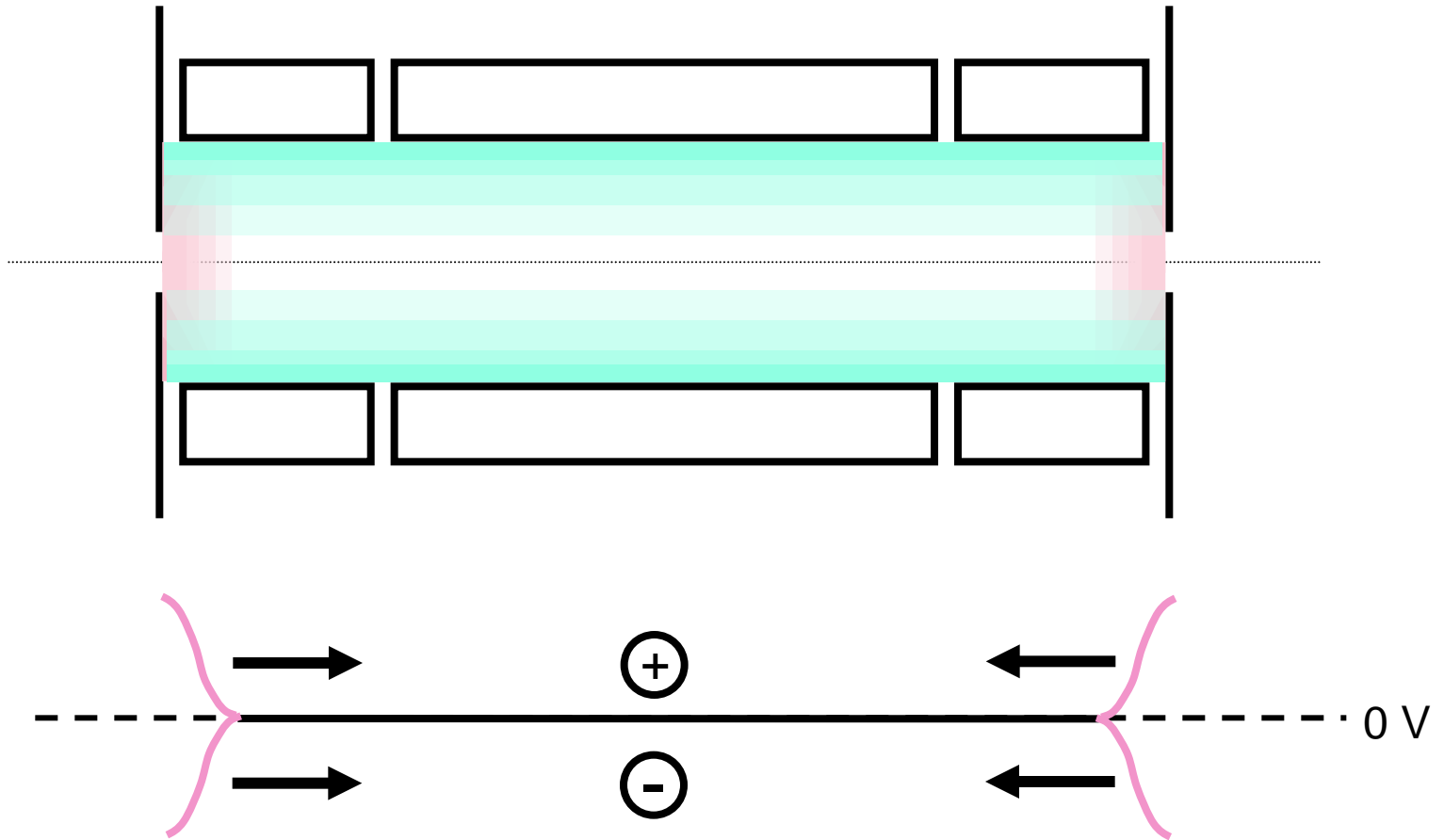


Charge sign independent radial confinement

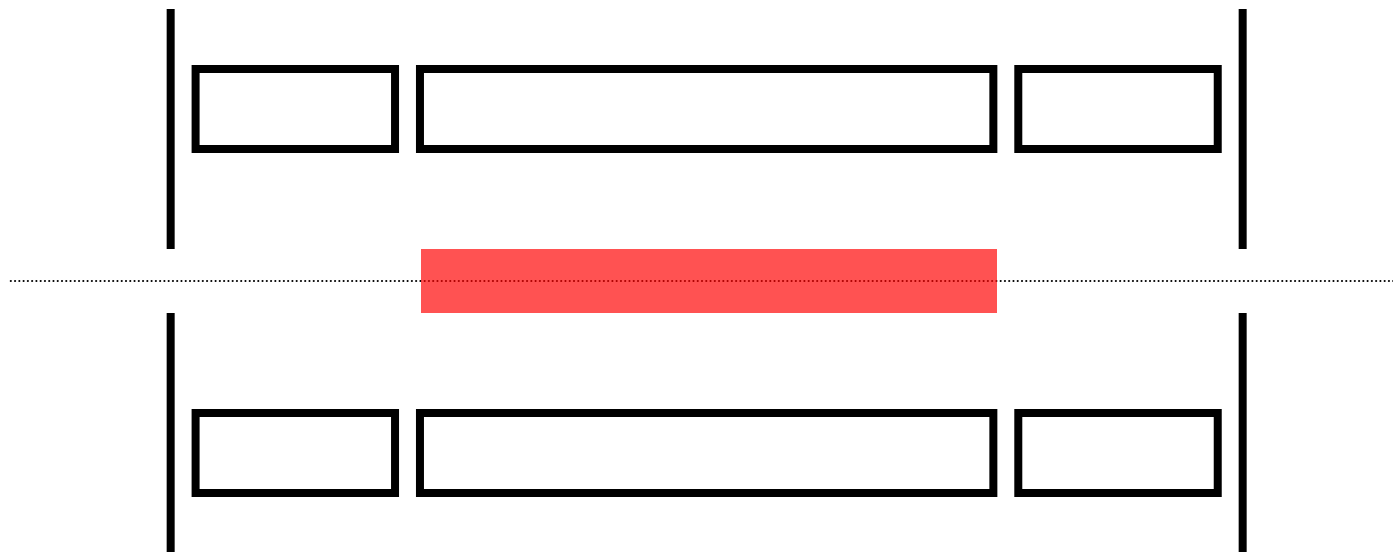


**Axial Confinement With DC Potentials
Trapping is Charge Sign Dependent**

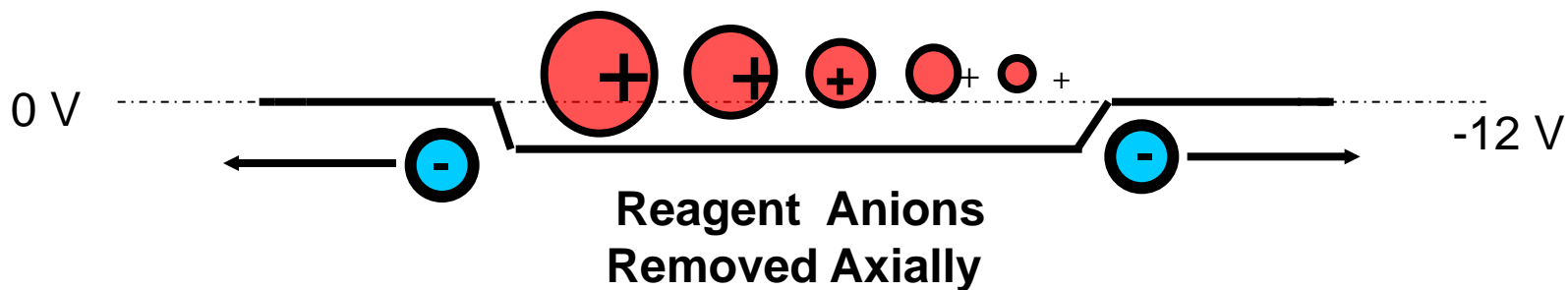
Charge sign independent axial confinement with combined RF Quadrupole and end lens RF pseudo-potentials



End ion/ion reactions prepare for product ion analysis

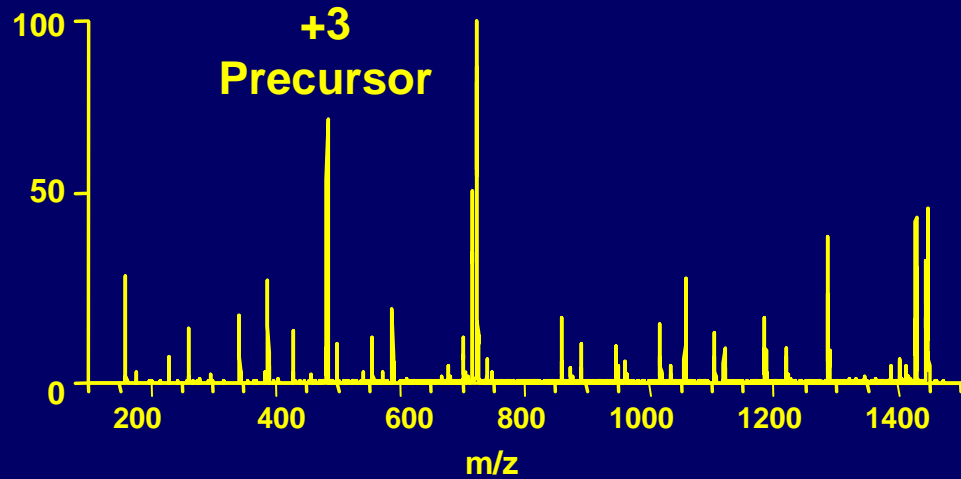
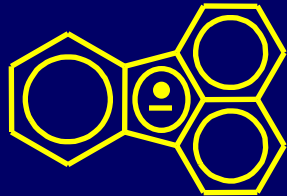


Product Cations Trapped in
Center Section For Scan Out



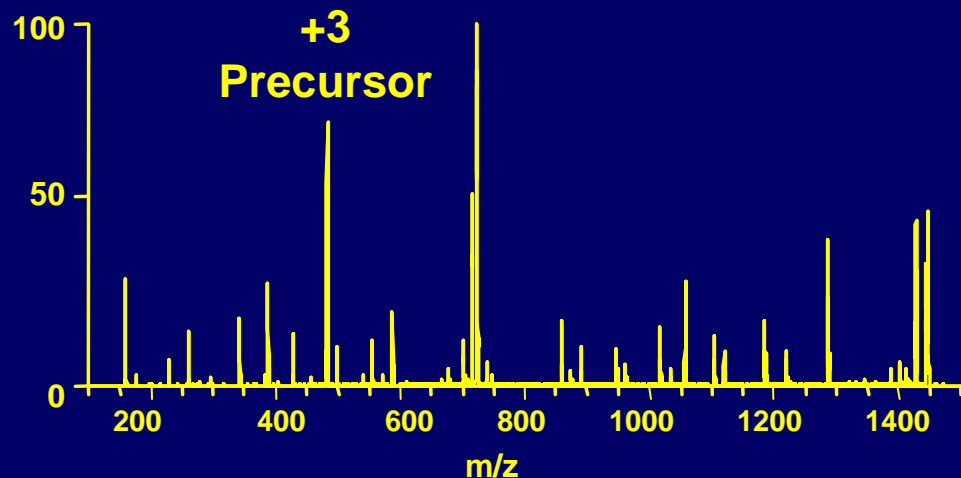
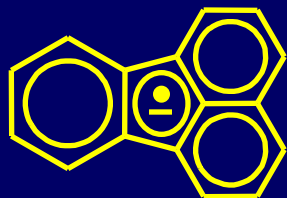
Electron Transfer - Proton Transfer

Fragmentation (ETD)

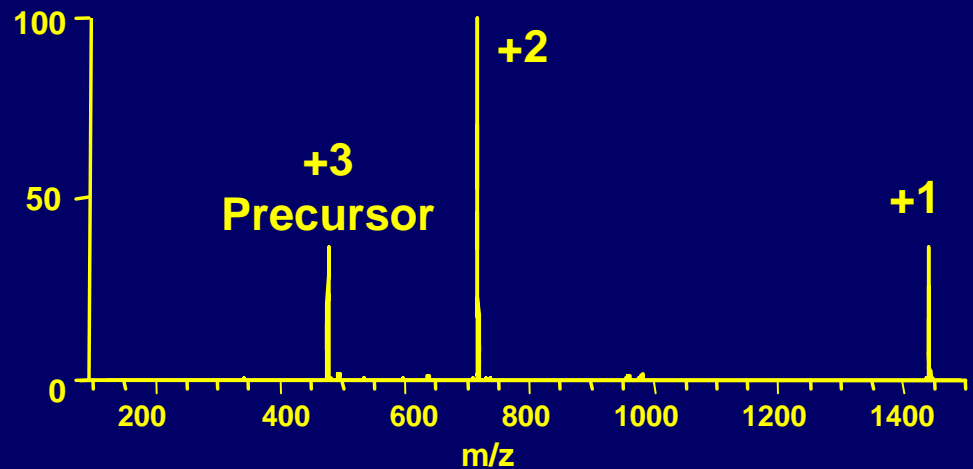
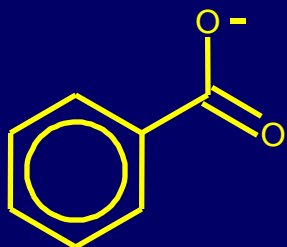


Electron Transfer - Proton Transfer

Fragmentation (ETD)



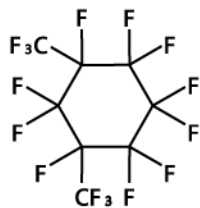
Charge Reduction (PTR)



The two types of ion reactions

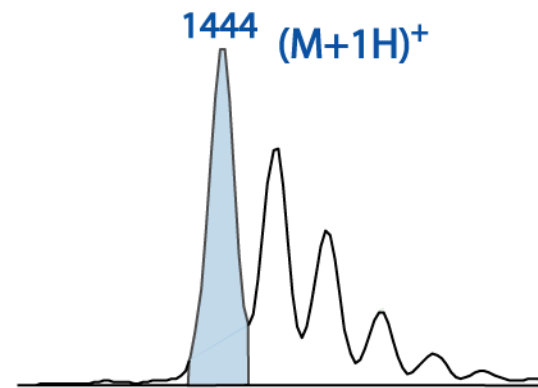
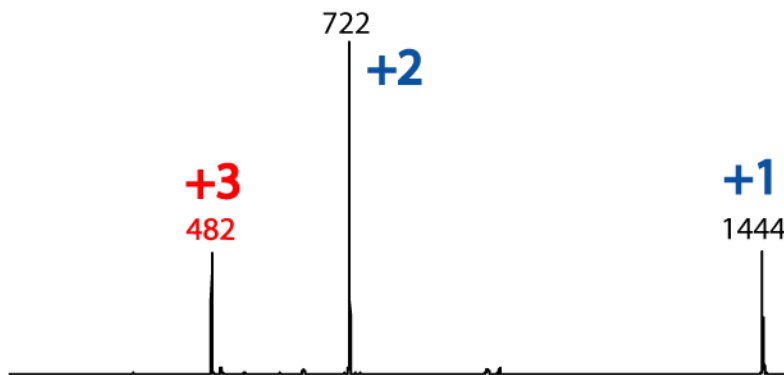
1.

PDCH



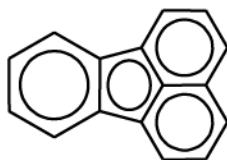
65 ms

Proton transfer (PTR)



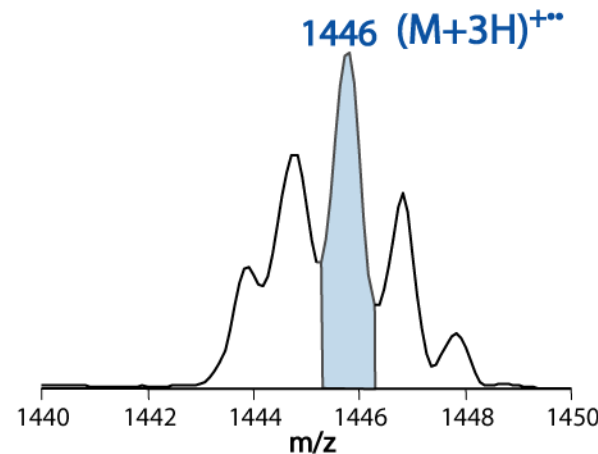
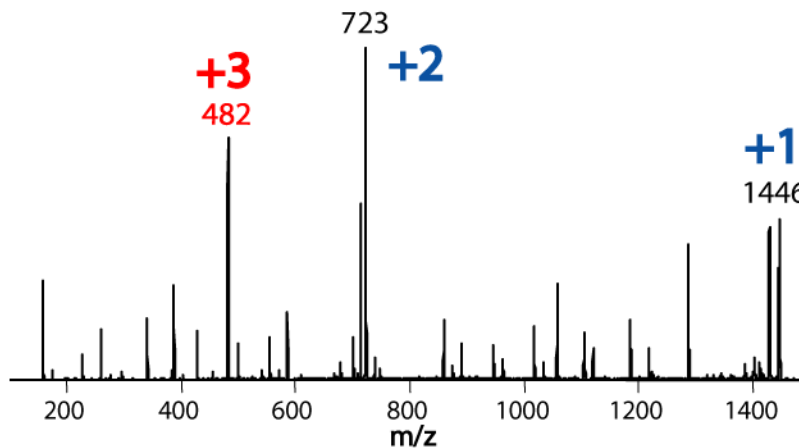
2.

fluoranthene

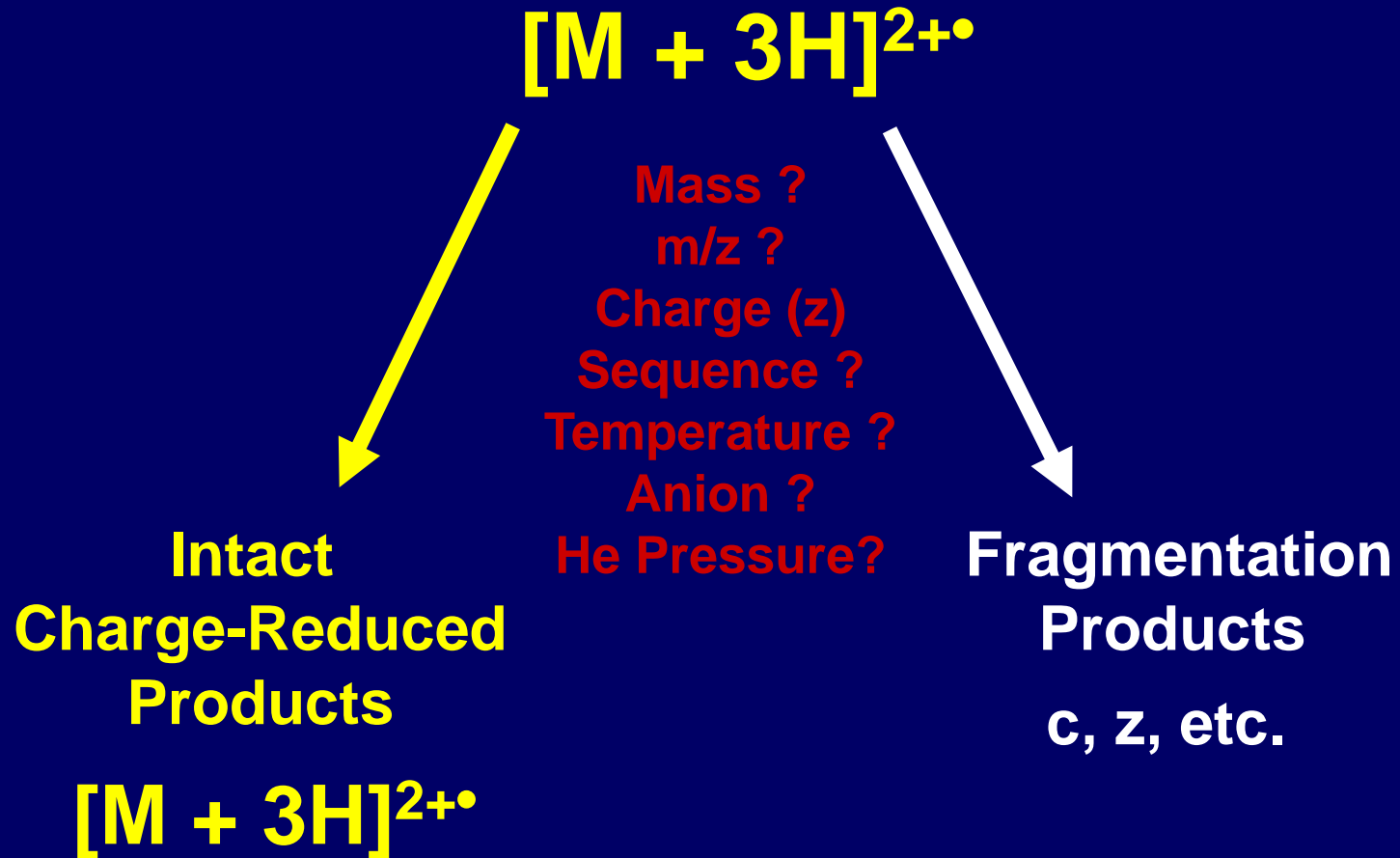


65 ms

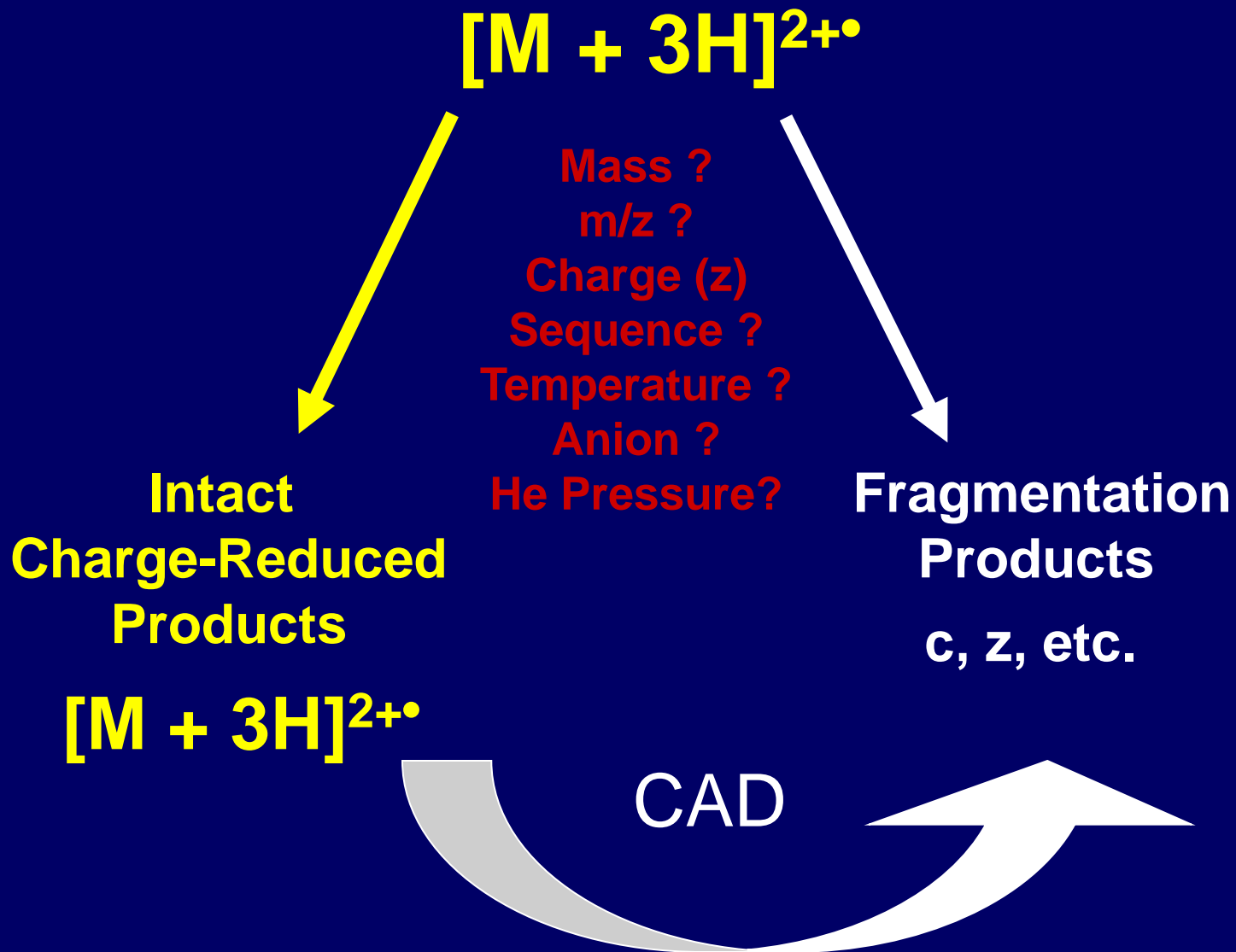
Electron transfer (ETD)



ET or ETD

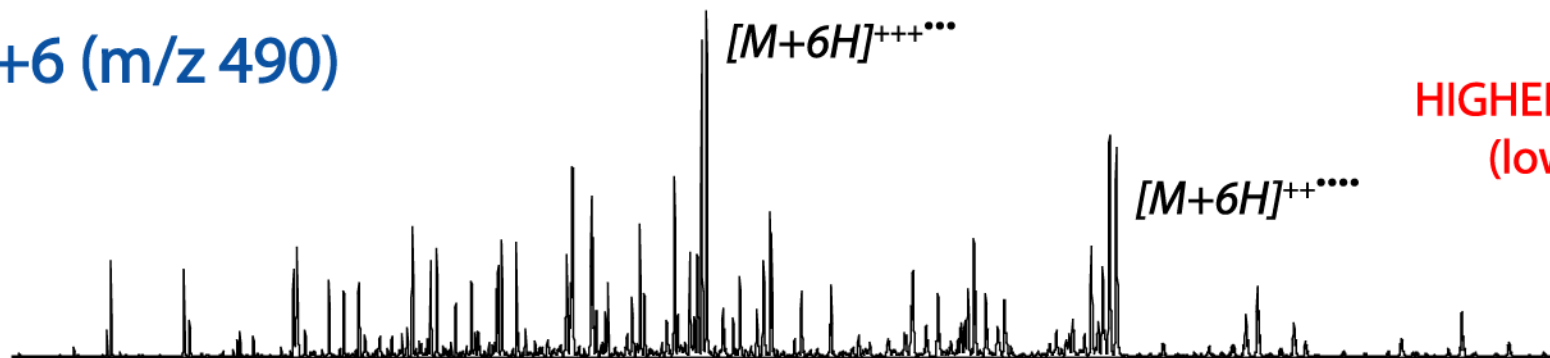


ET or ETD



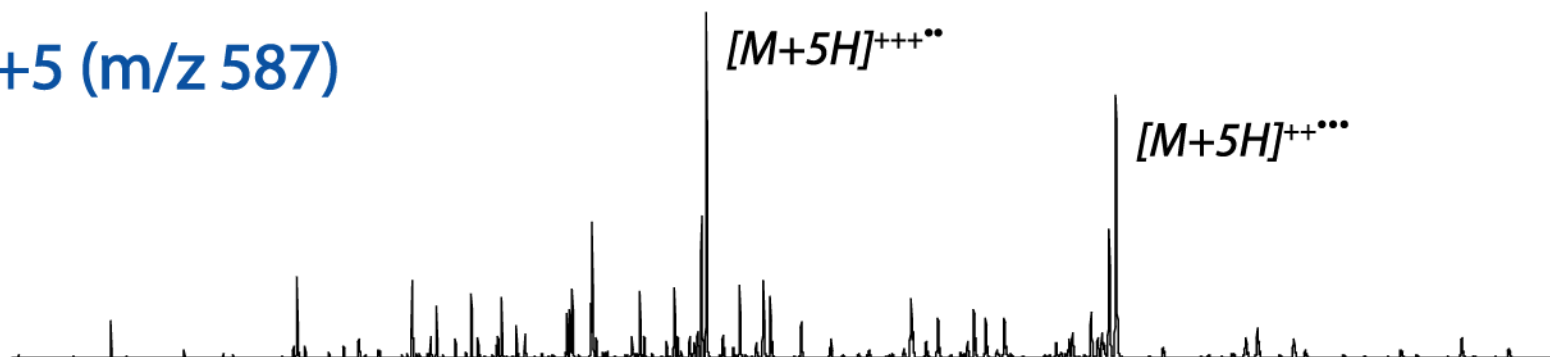
Charge dependence in fragmentation

+6 (m/z 490)



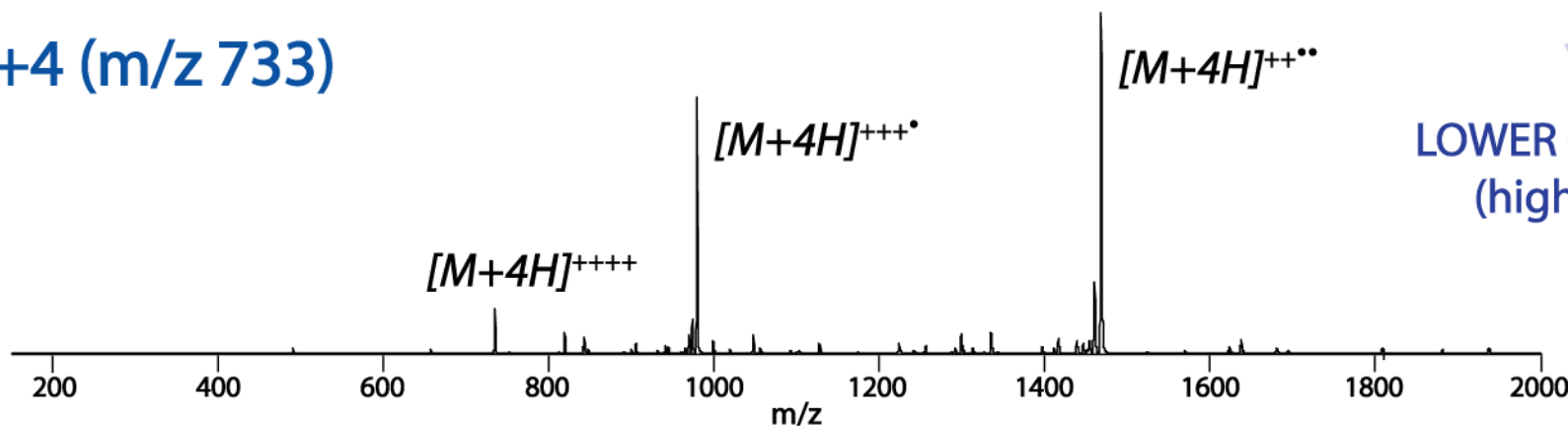
HIGHER CHARGE
(low m/z)

+5 (m/z 587)



decreasing c/z-product
yield

+4 (m/z 733)

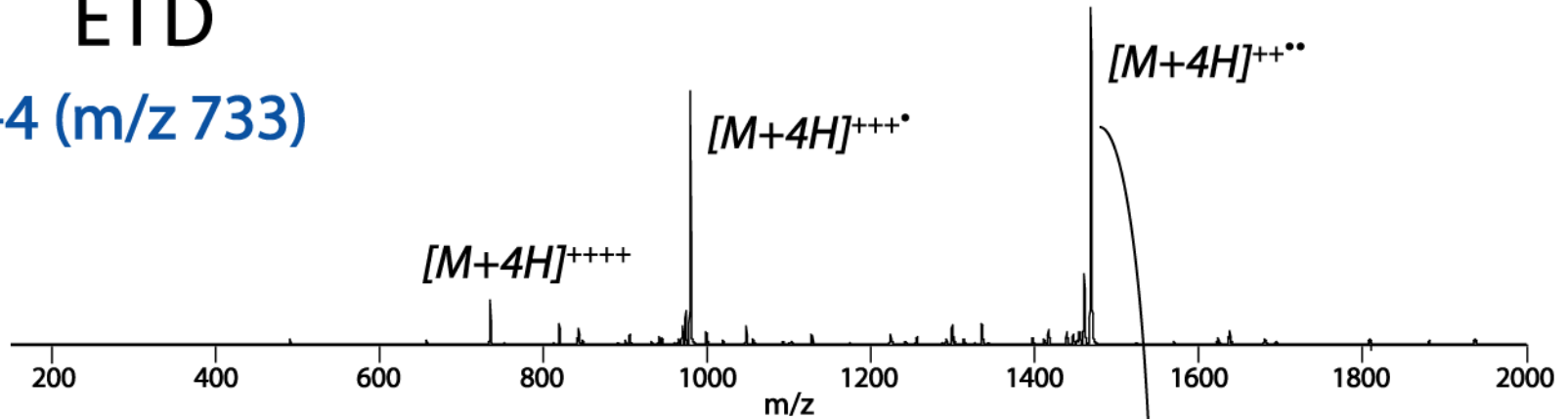


LOWER CHARGE
(high m/z)

Gentle off resonance activation

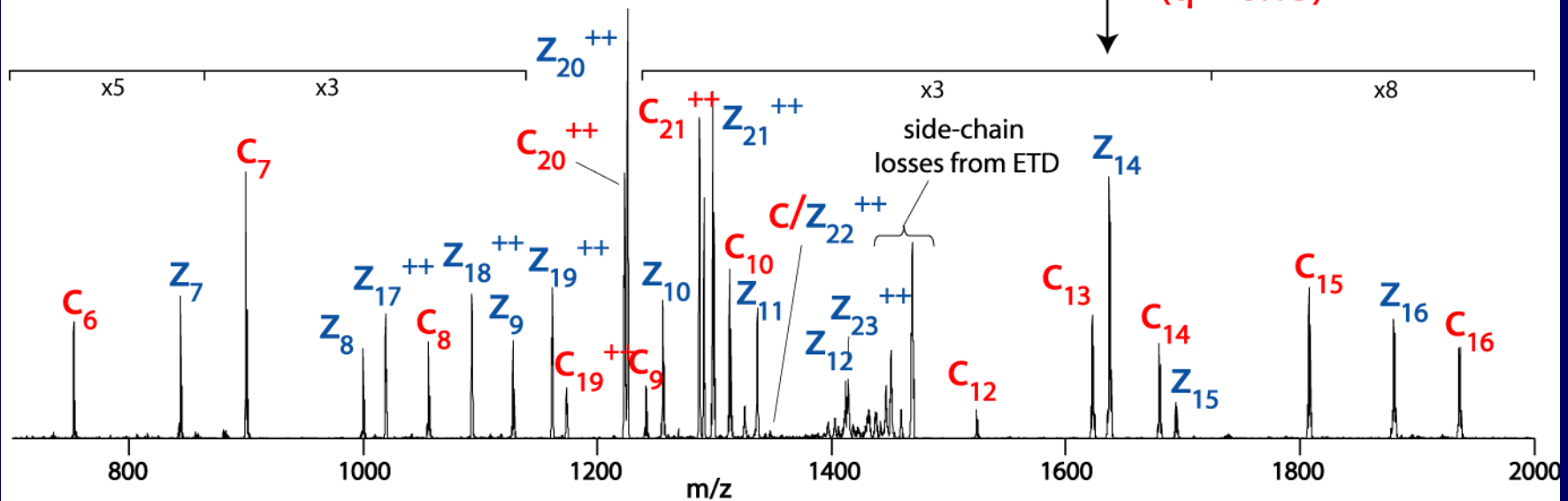
ETD

+4 (m/z 733)

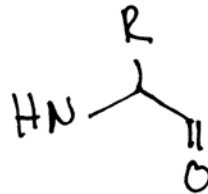
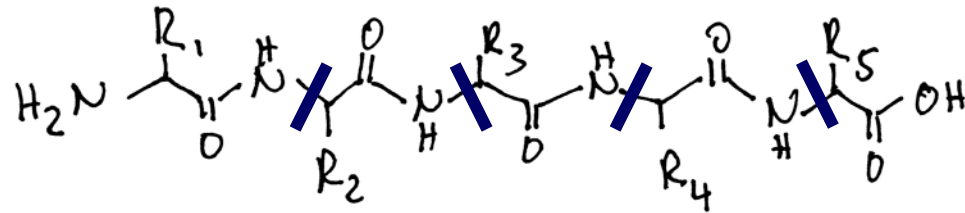


S|Y|S|M|E|H|F|R|W|G|K|P|V|G|K|K|R|R|P|V|R|V|Y|P

gentle
activation
($q = 0.15$)

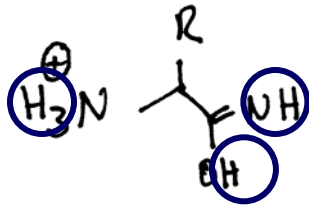


How to Sequence ETD



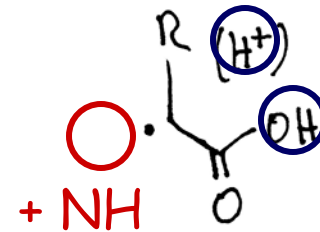
Residue Mass (RM)

c1 ion



$$c1 = RM + 18$$

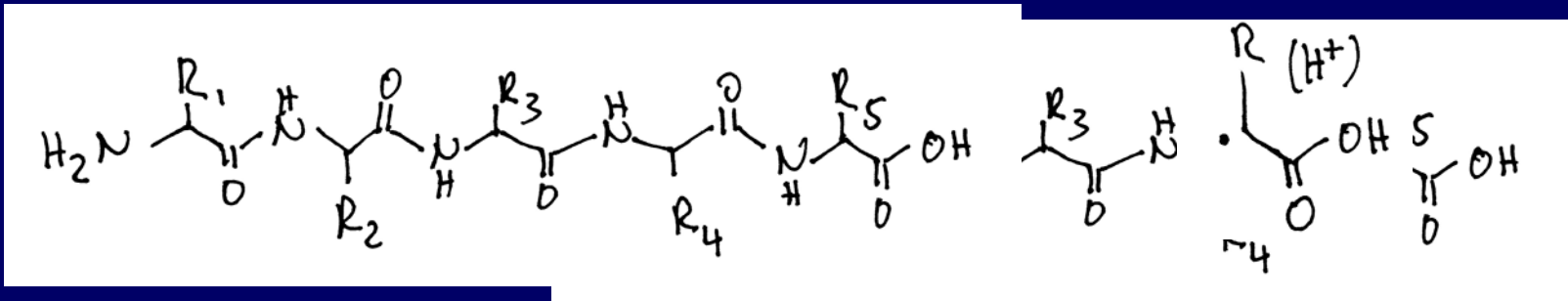
z1 ion



$$z1 = RM + 3$$

Example of how to calculate theoretical fragment ions

105	176	307	404	517	646	803
S	A	M	P	L	E	R
803	700	629	498	401	288	159

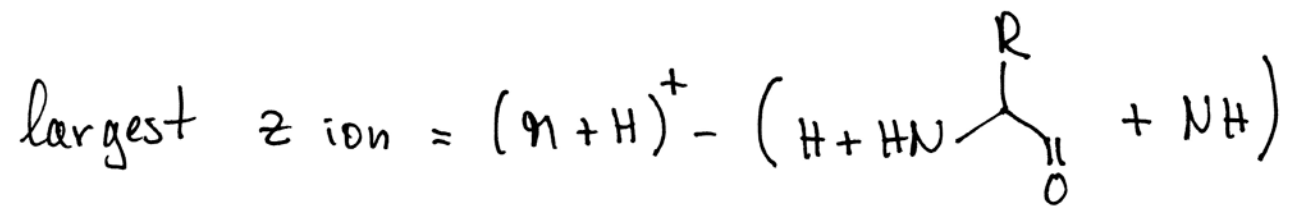


The first c ion

Residue Mass

The first z ion

Largest c and z ions



Background necessary to know

**Show a hand annotated spectra from
one of my toxin talks**

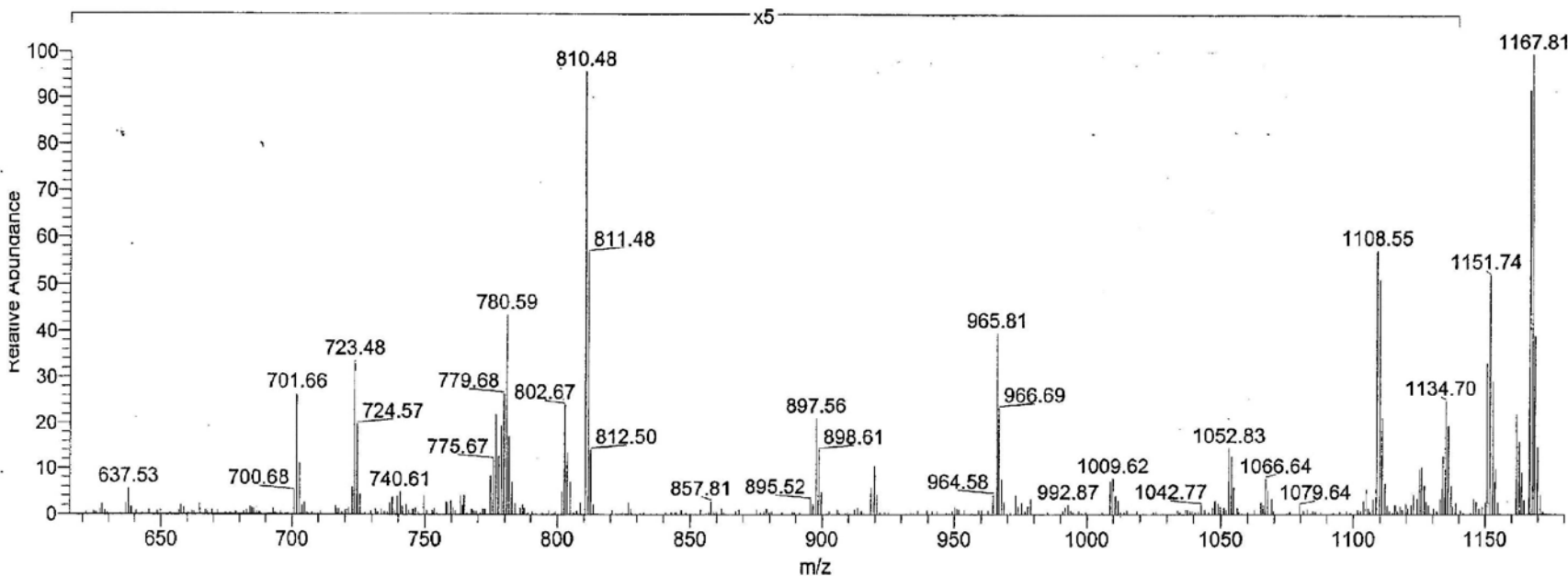
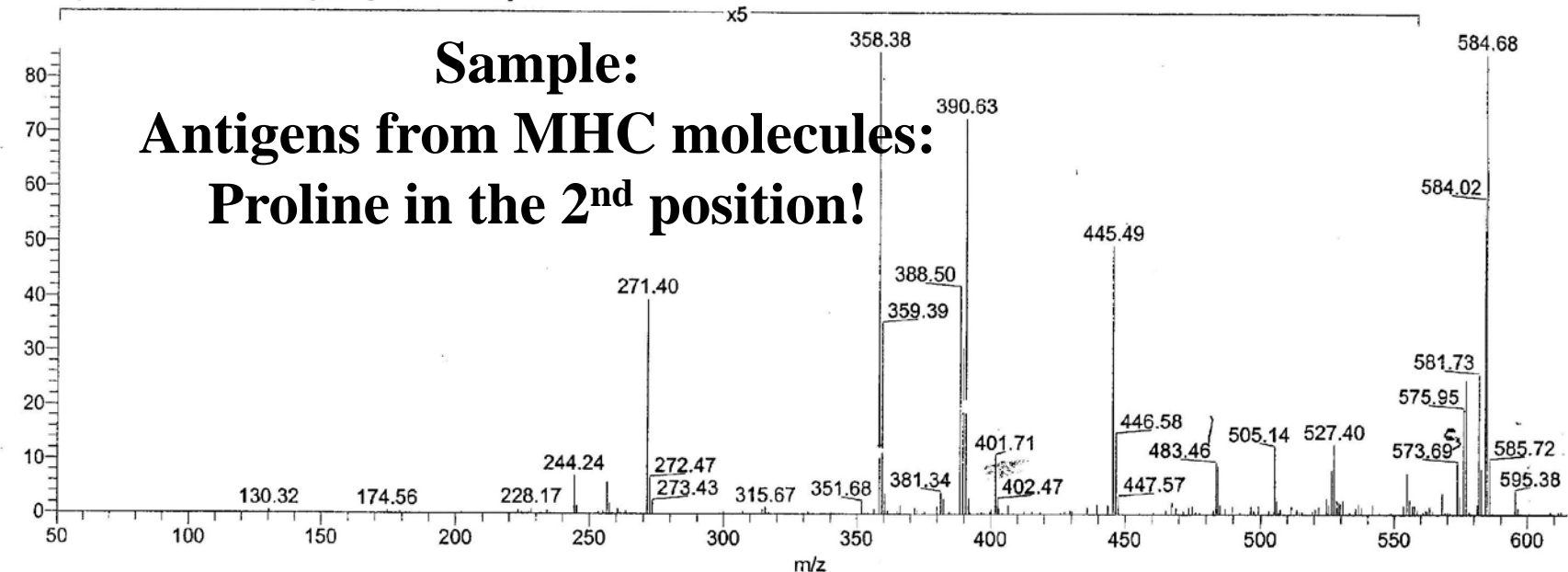
**Show how to calculate a charge state,
also in a spectrum, give a spectrum
where they should find out the charge
state for CAD and then for ETD and
how they confirm that by finding pairs
(must have mentioned that in the
previous slide for the example)**

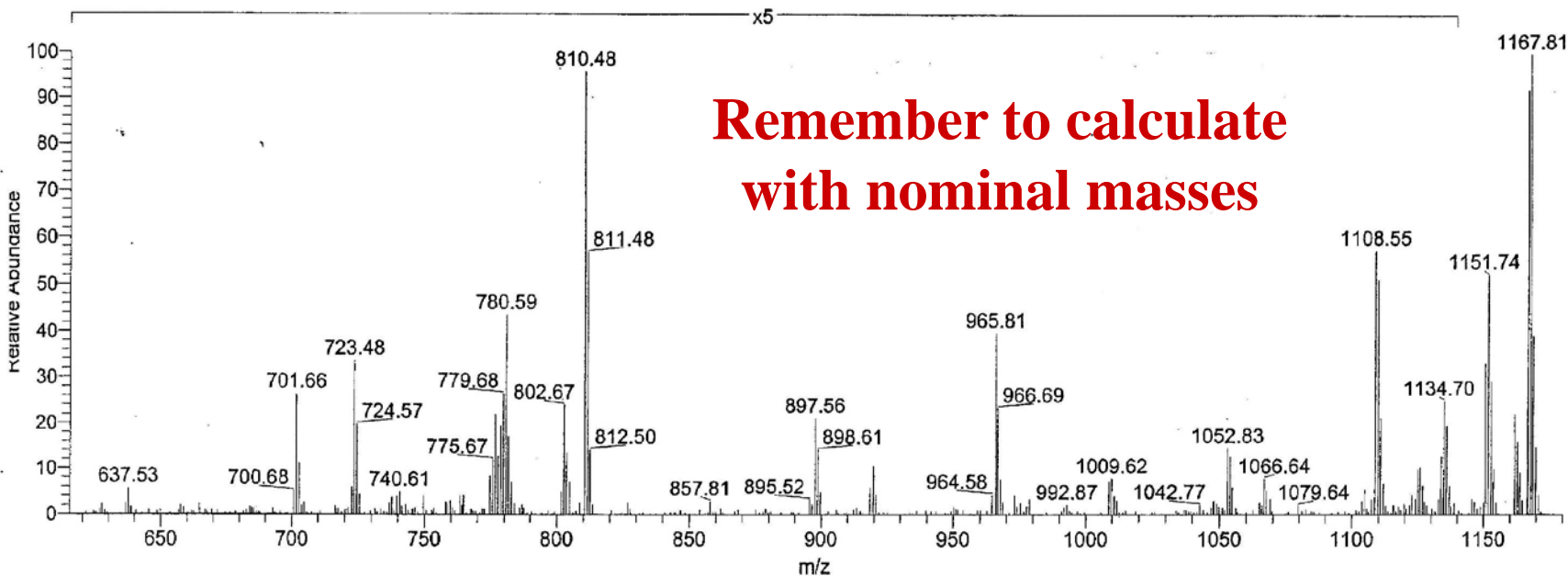
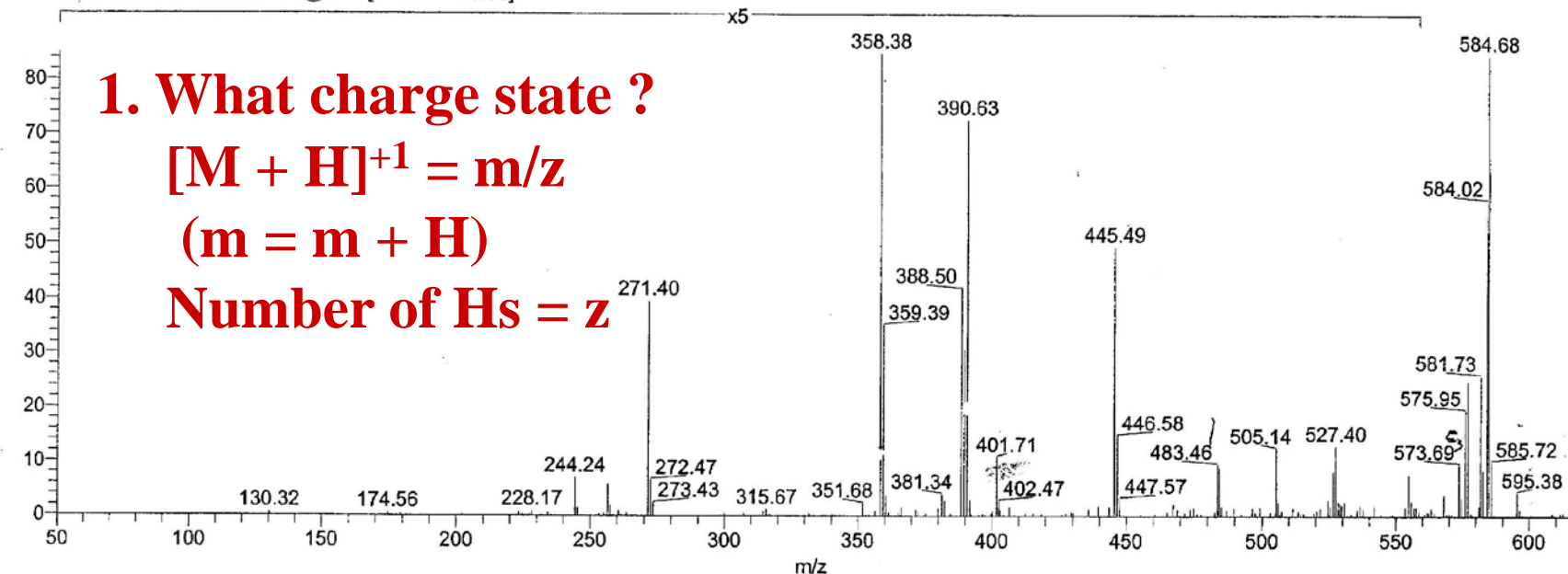
**Ask them if they know how to calculate
a charge state etc.**

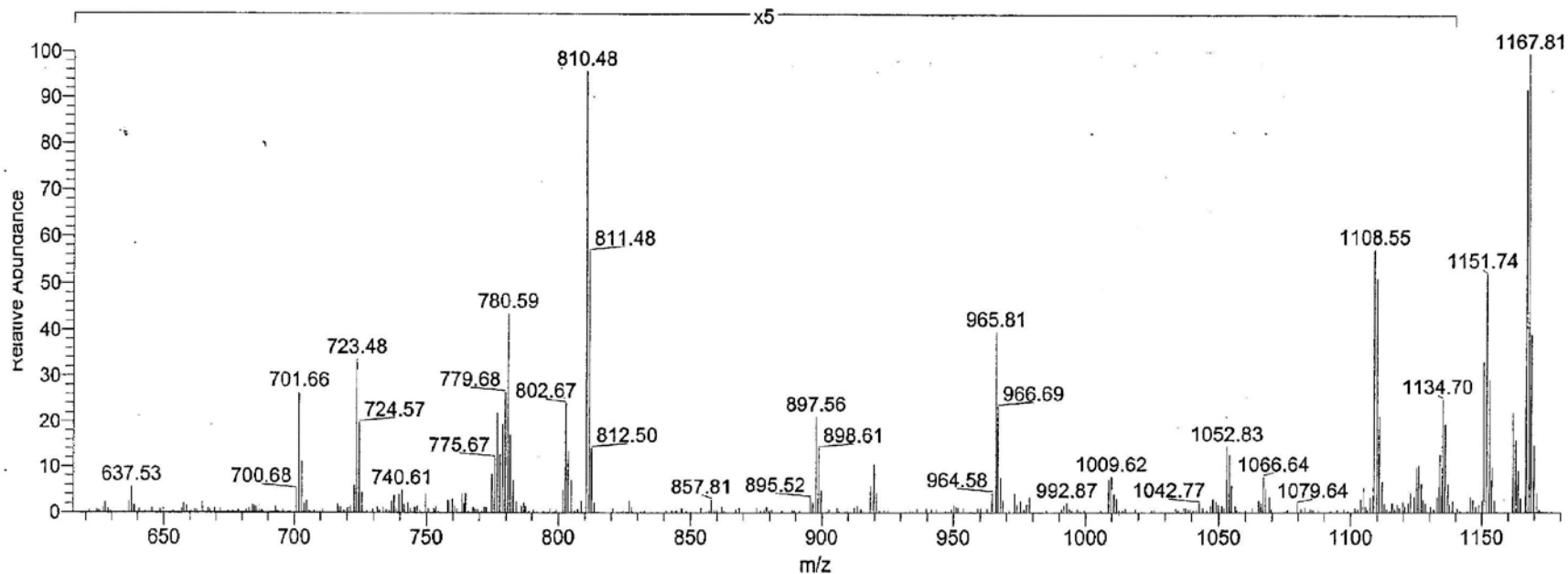
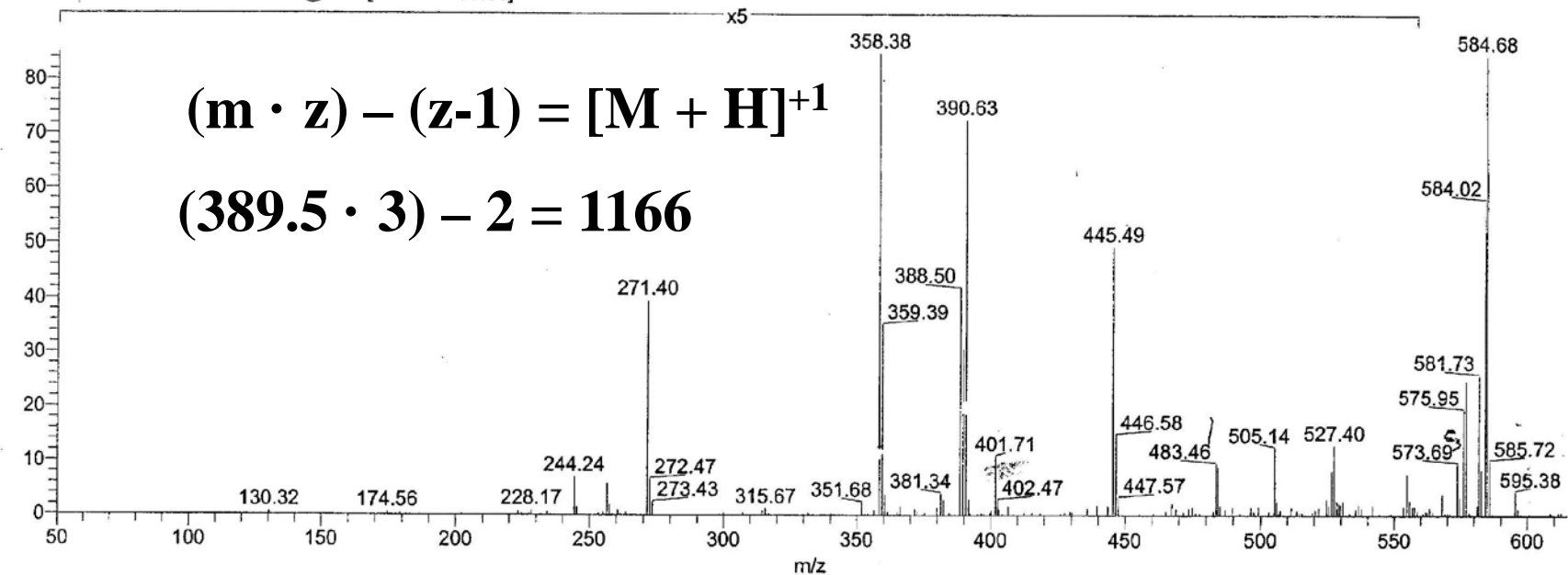
Characteristics of CAD and ETD spectra

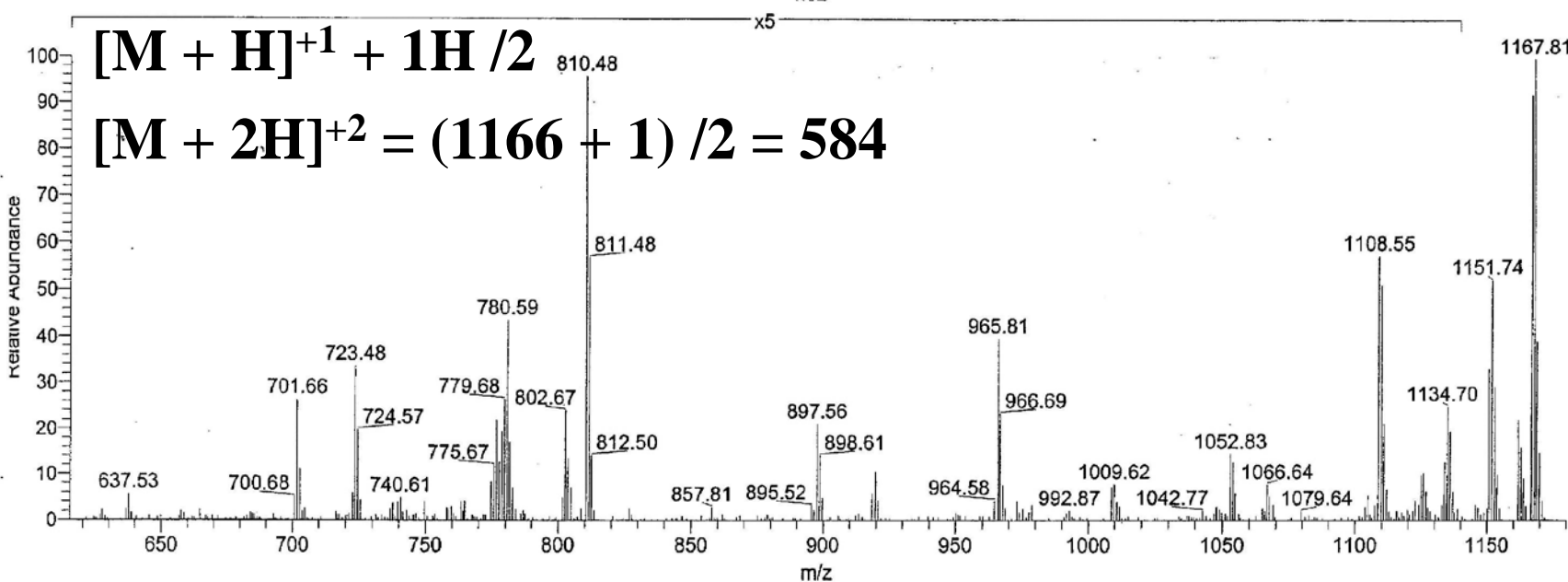
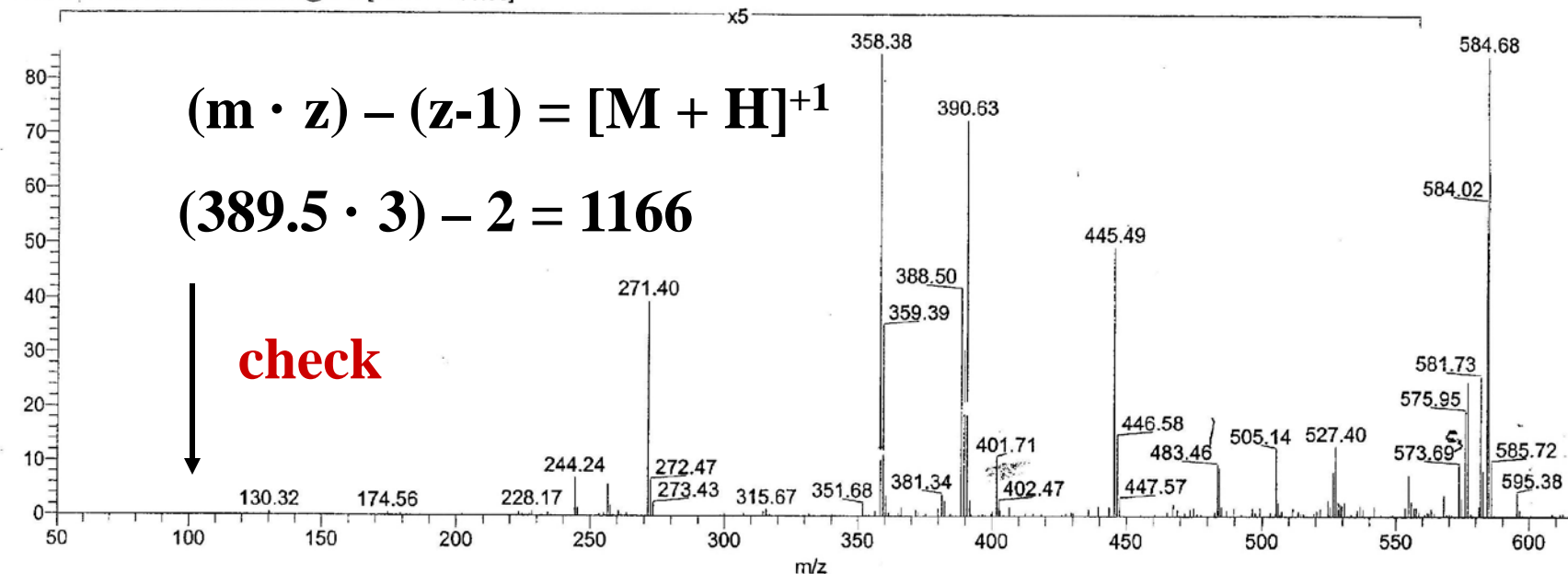
**Show the precursor
ion, show the
neutral loss from the
parent, show the
neutral loss from
peptides**

Sample:
Antigens from MHC molecules:
Proline in the 2nd position!

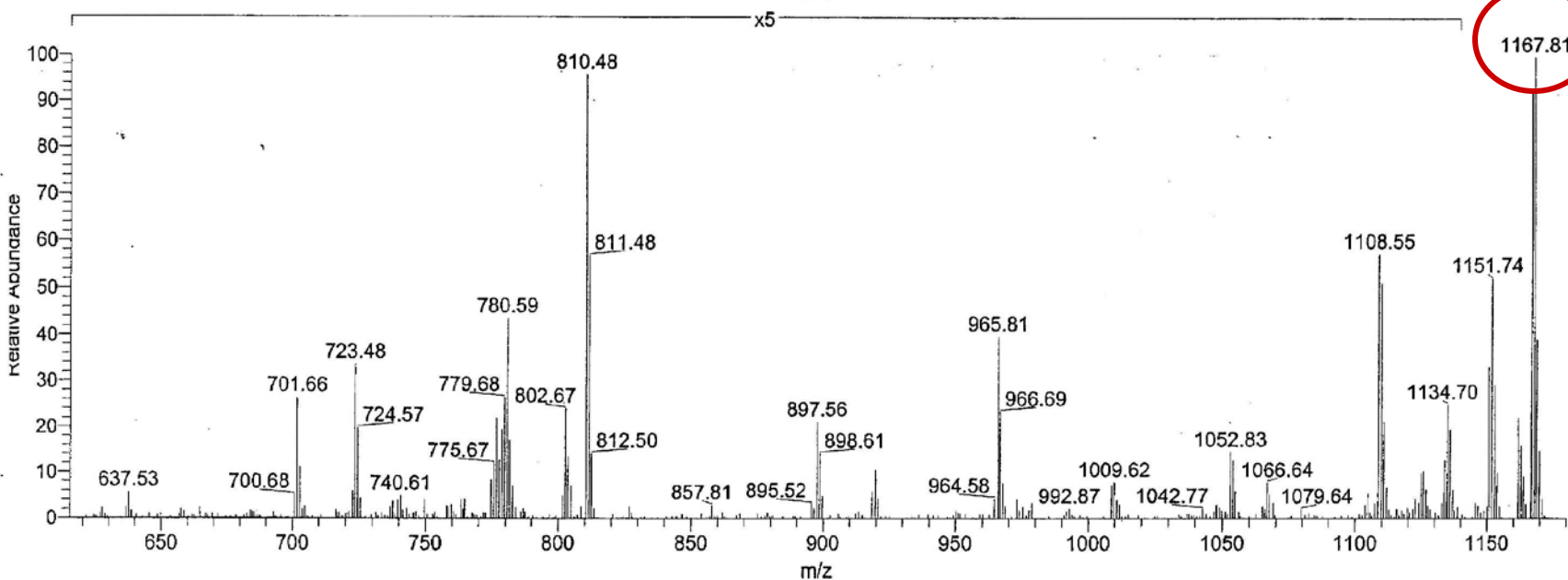
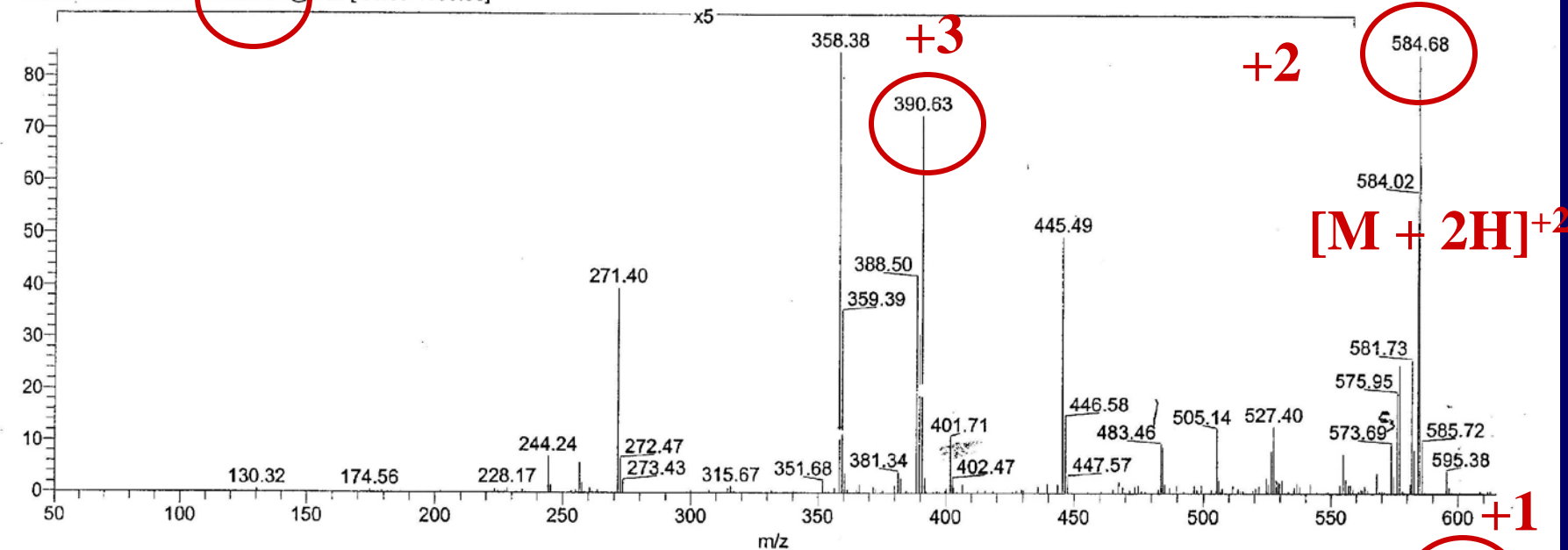


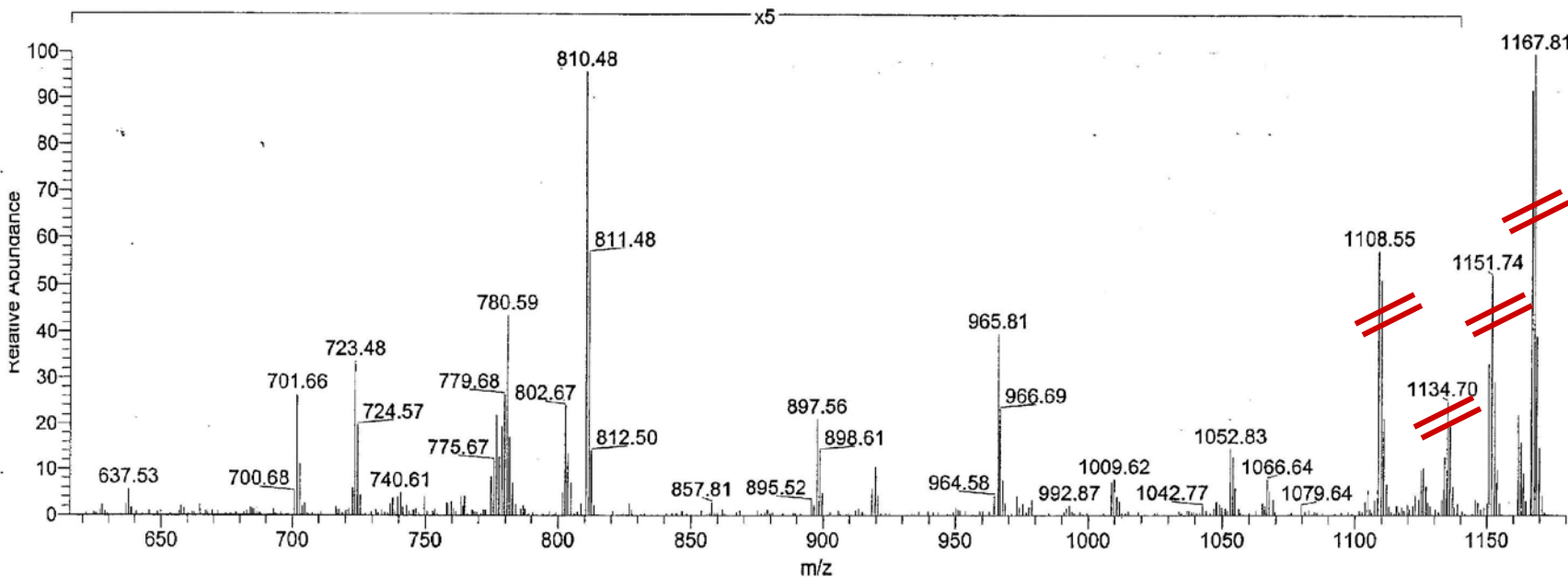
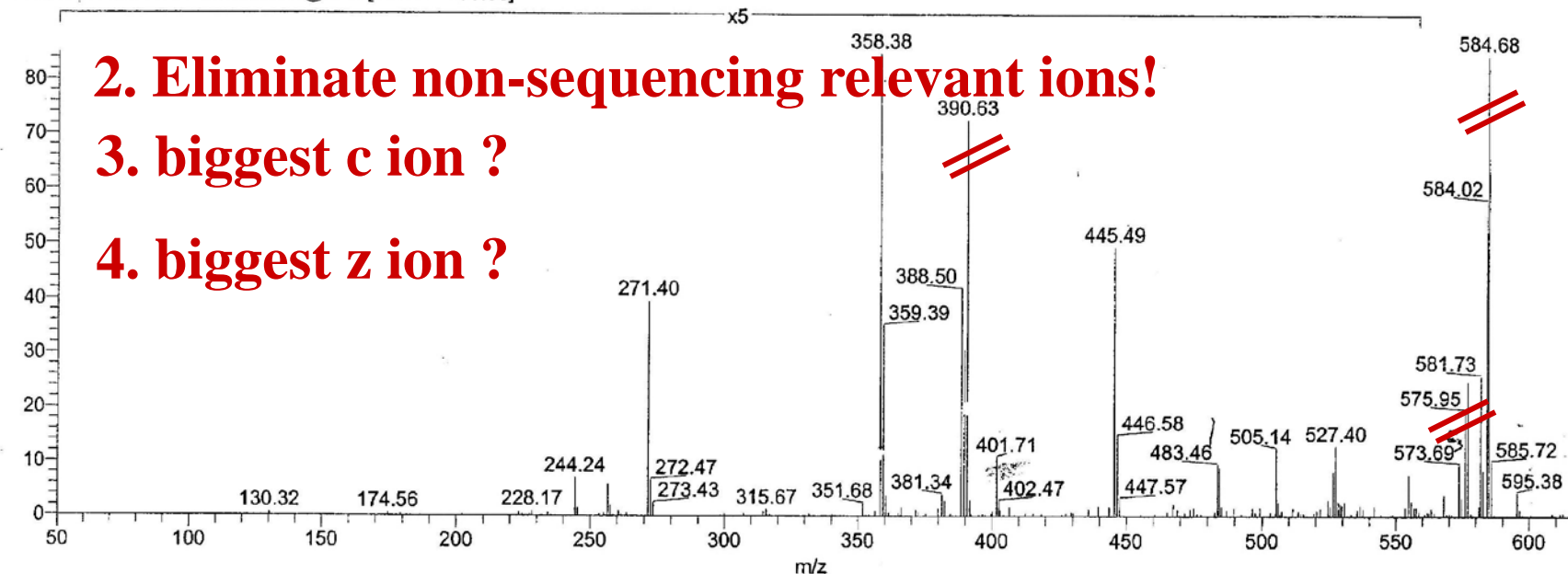


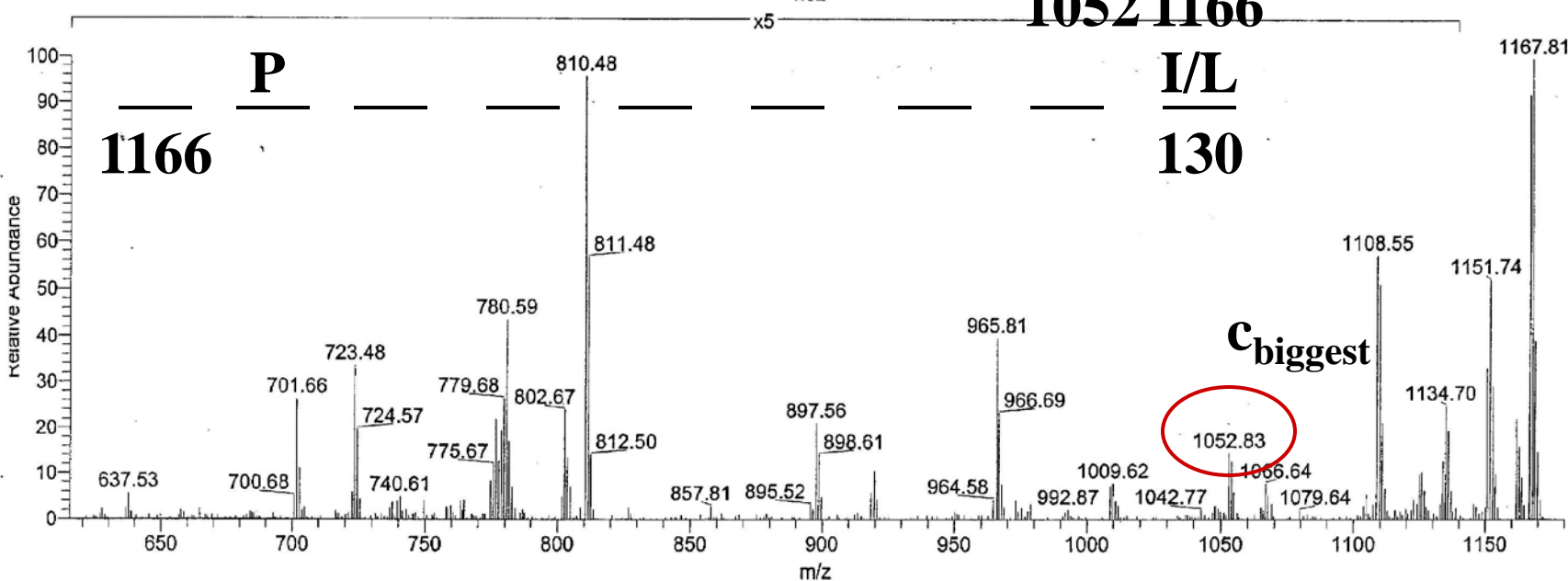
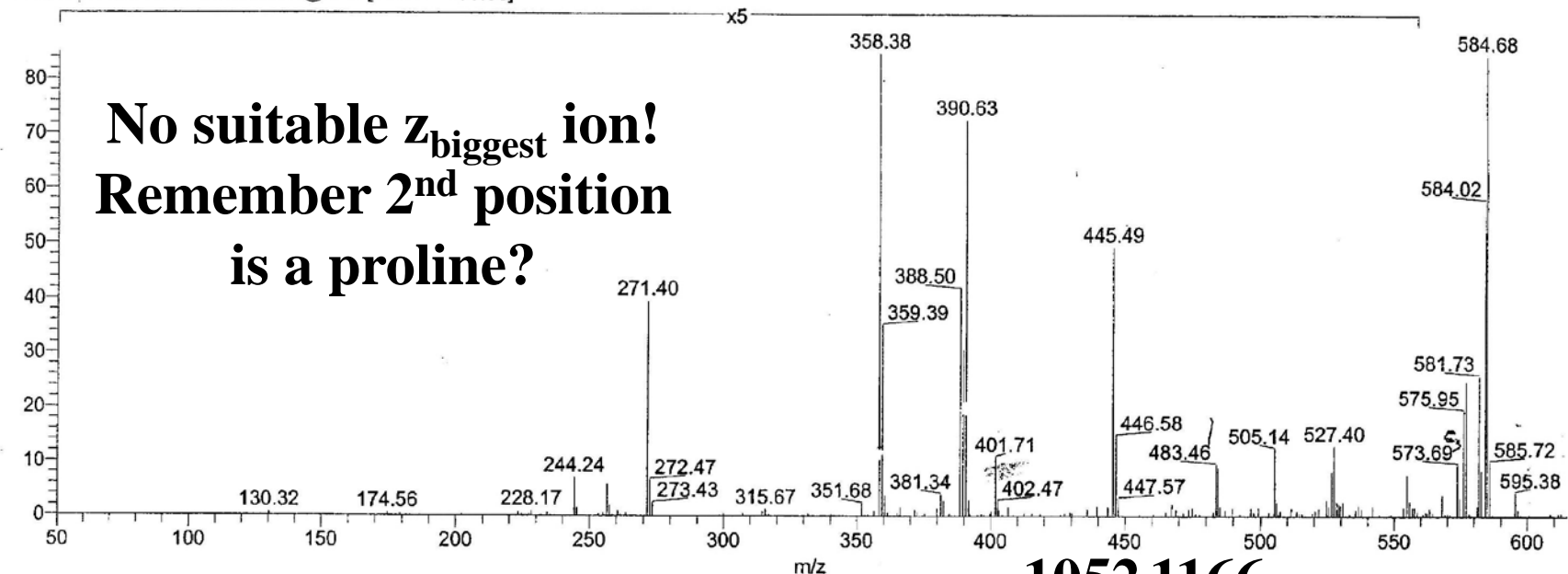


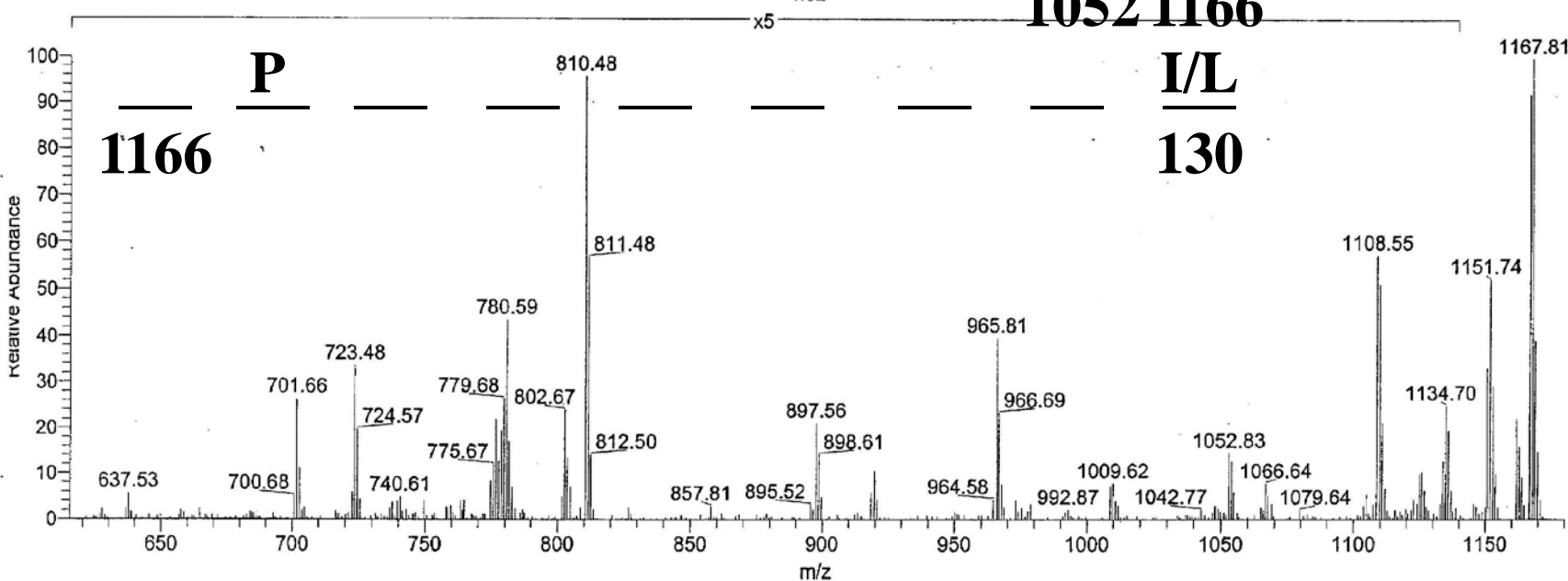
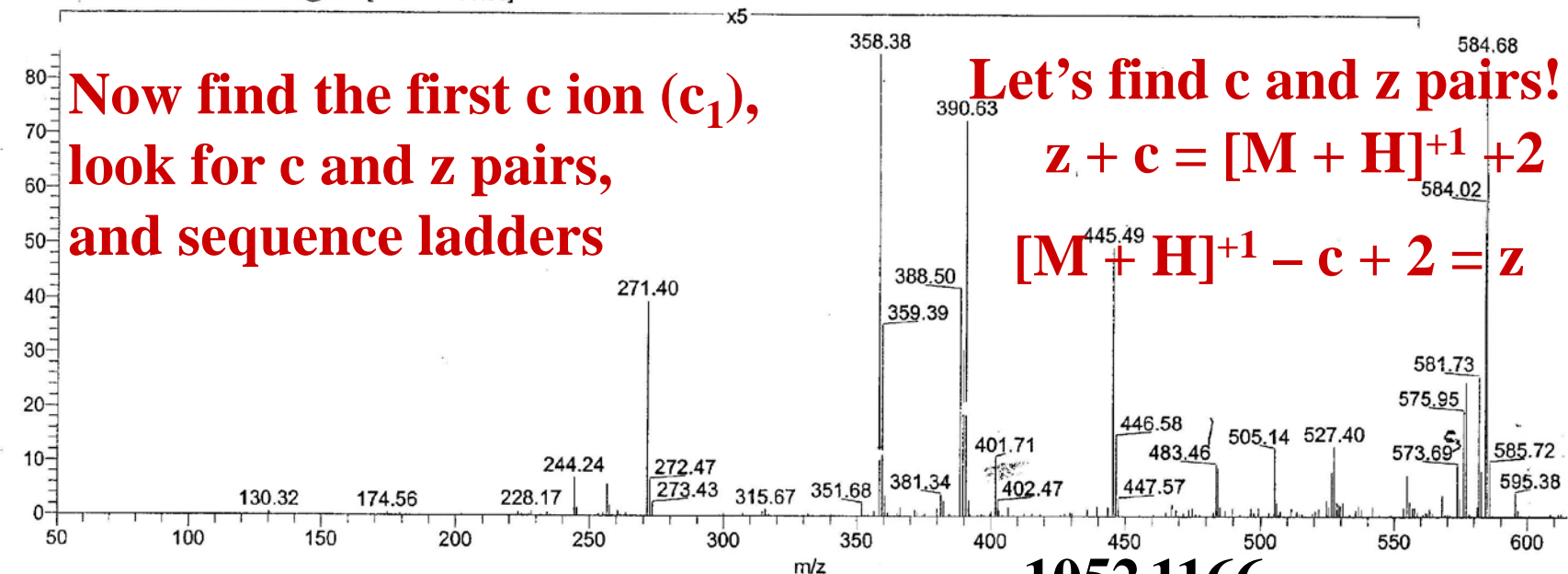


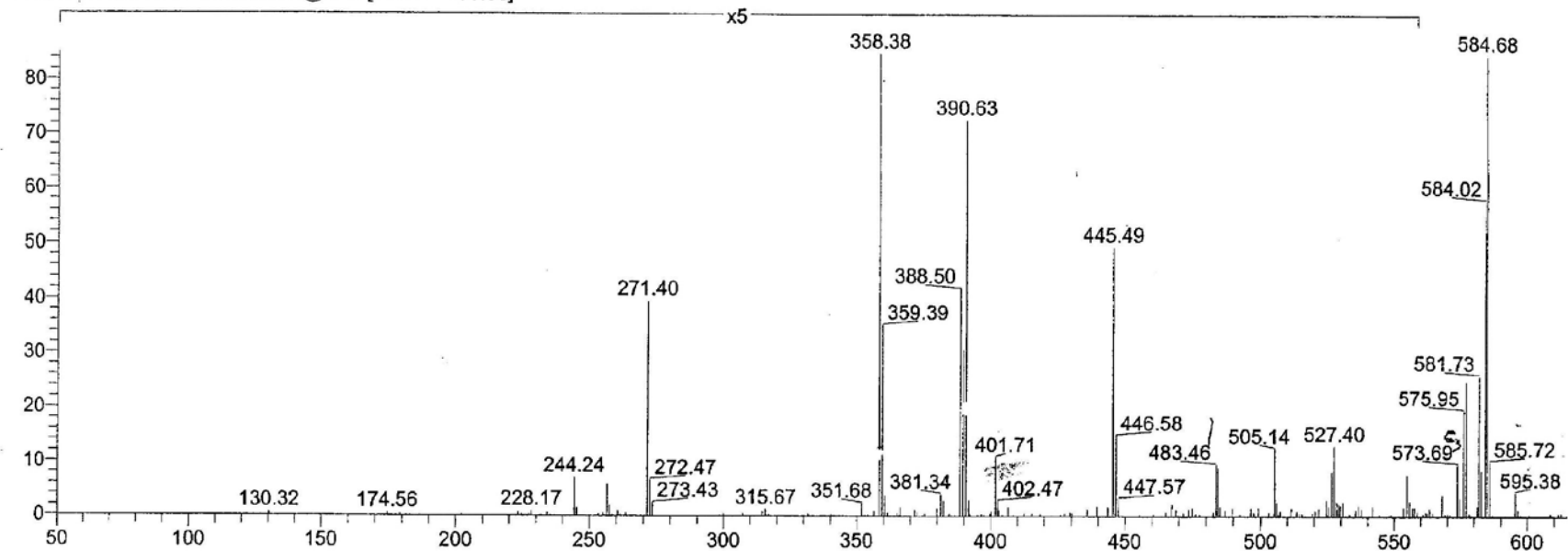
$[M + 3H]^{+3}$







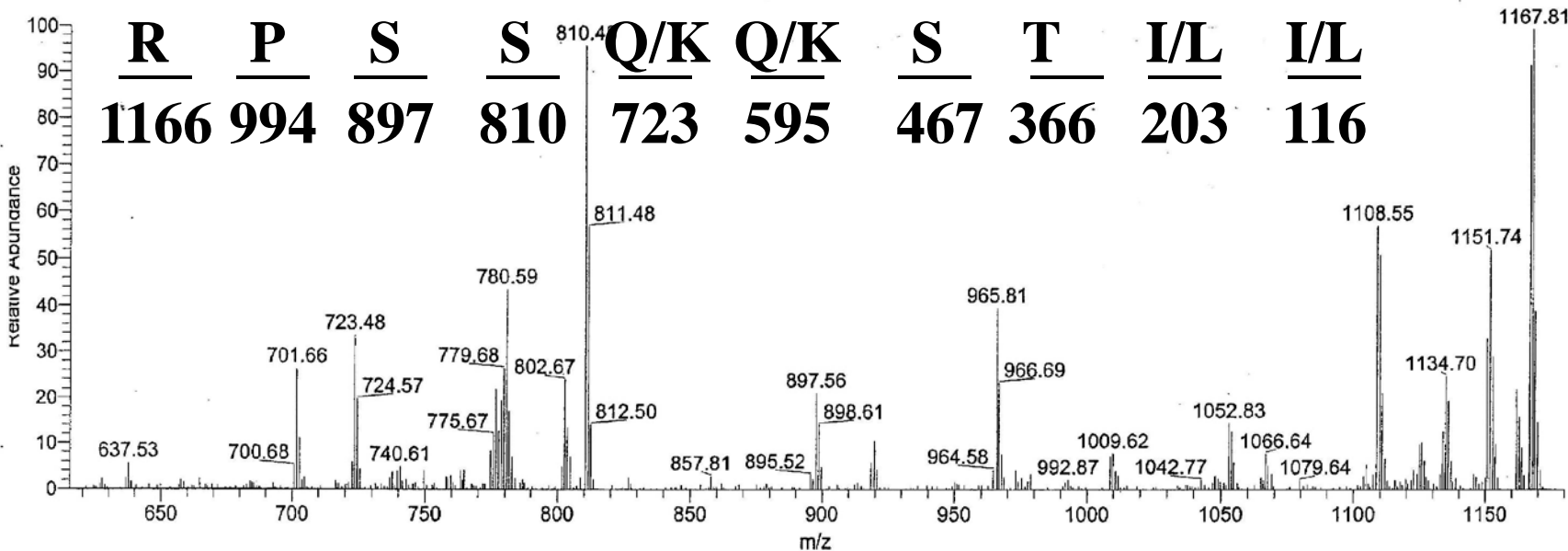




174 271 358 445 573 701 802 965 1052 1166

R P S S Q/K Q/K S T I/L I/L

1166 994 897 810 723 595 467 366 203 116



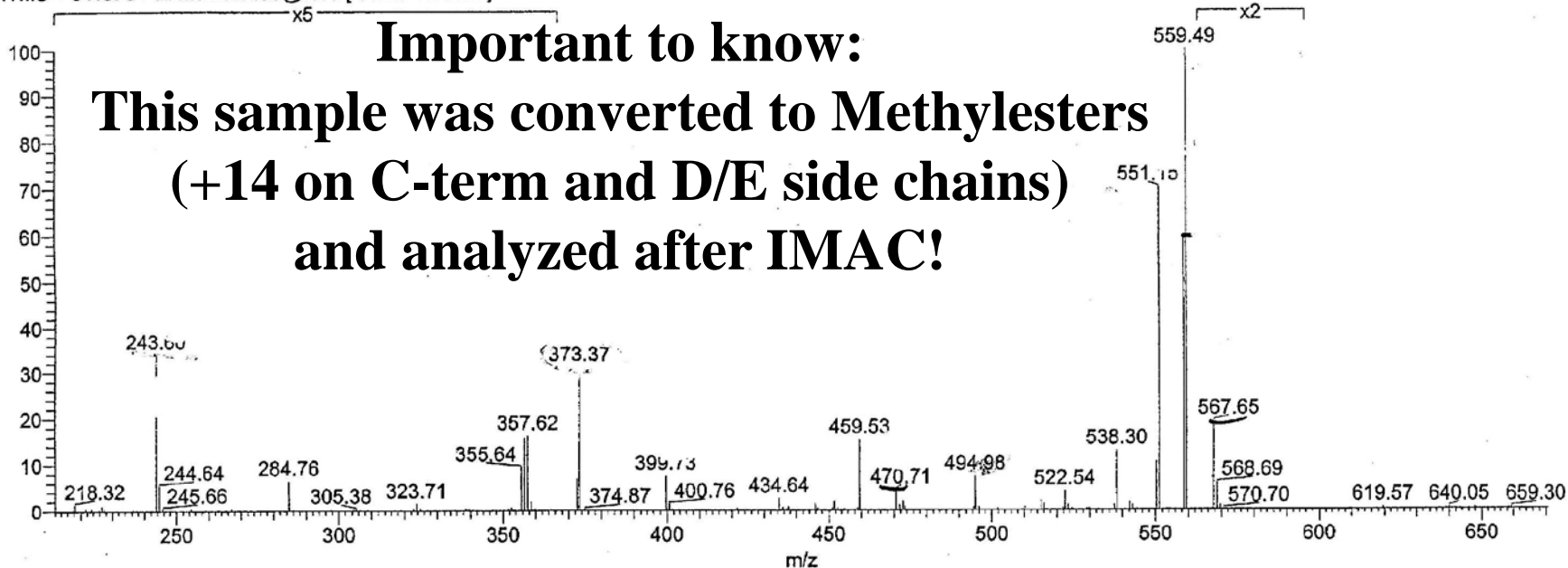
2nd Example

Methylesters

jysJYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6
: ITMS + c NSI d Full ms2 372.85@0.00 [90.00-1130.00]

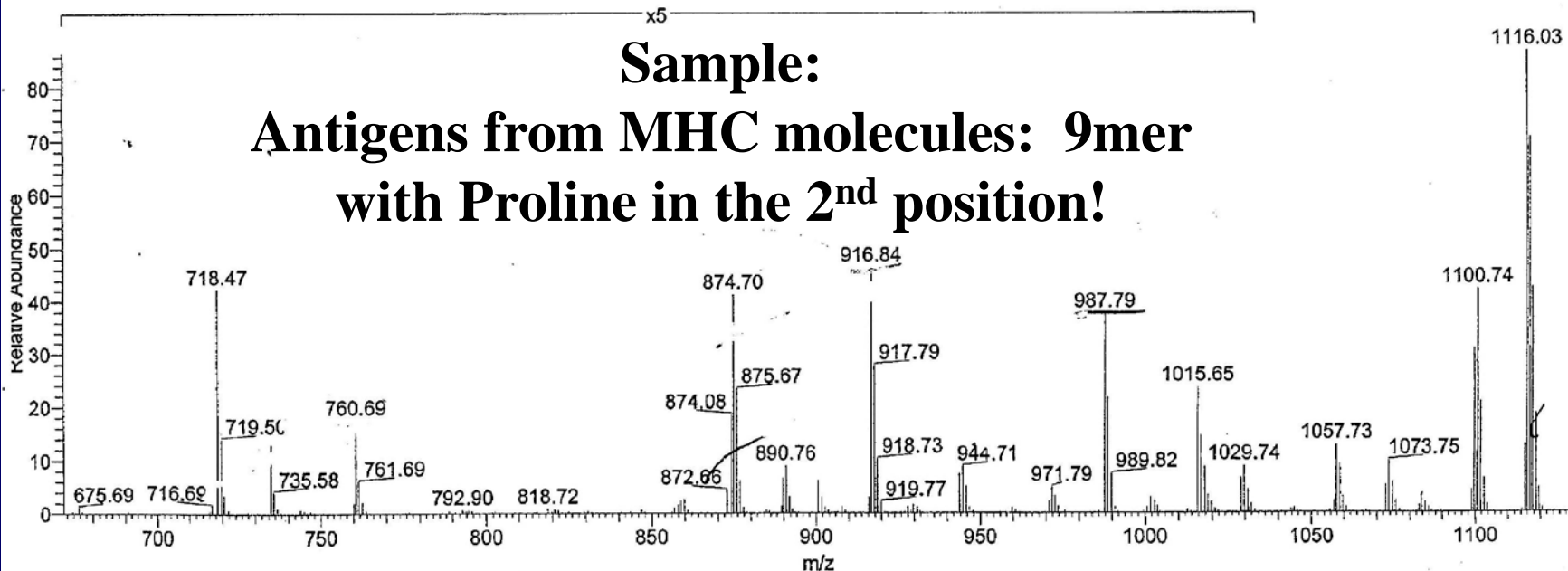
Important to know:

**This sample was converted to Methylesters
(+14 on C-term and D/E side chains)
and analyzed after IMAC!**



Sample:

**Antigens from MHC molecules: 9mer
with Proline in the 2nd position!**



td test

ijsJYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms2 372.85@0.00 [90.00-1130.00]

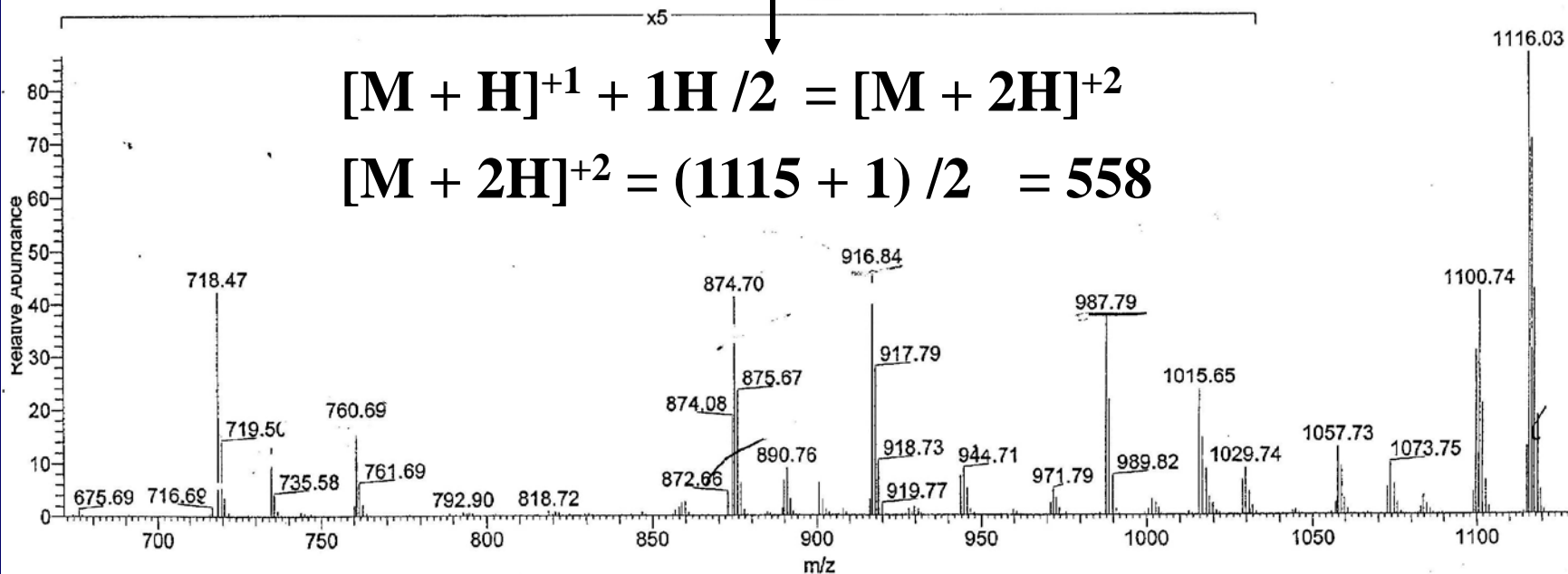
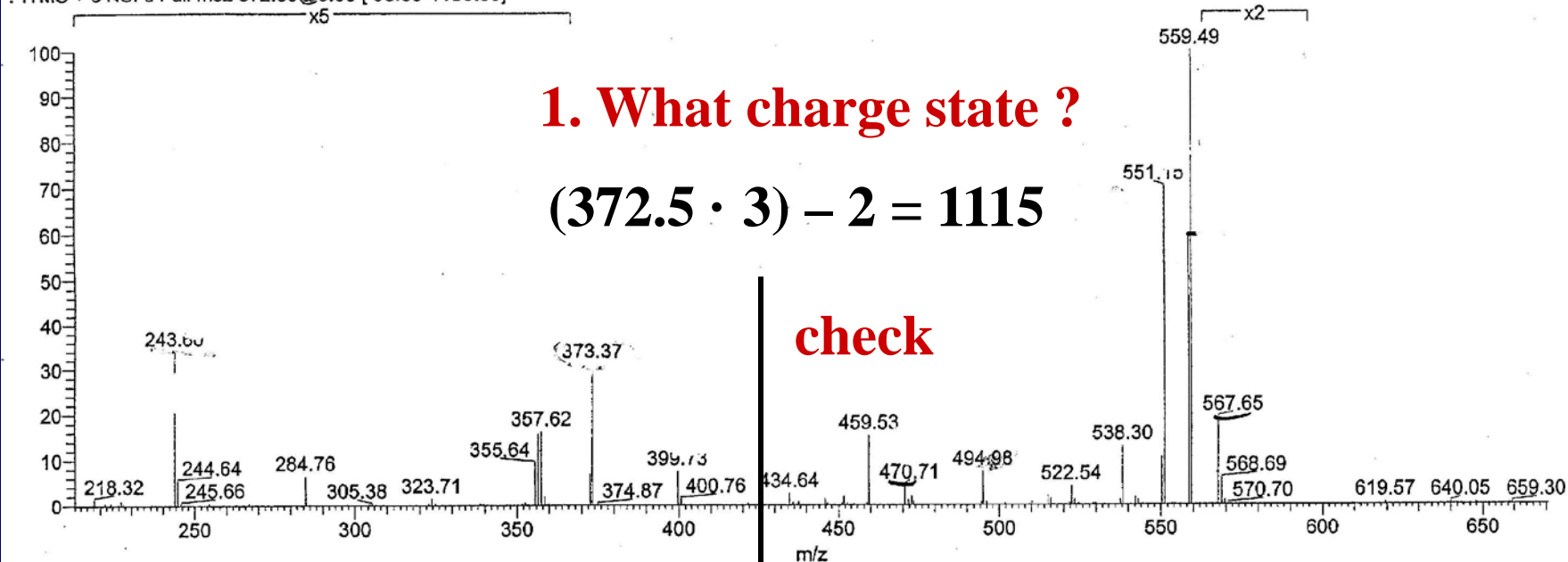
1. What charge state ?

$$(372.5 \cdot 3) - 2 = 1115$$

check

$$[M + H]^{+1} + 1H / 2 = [M + 2H]^{+2}$$

$$[M + 2H]^{+2} = (1115 + 1) / 2 = 558$$

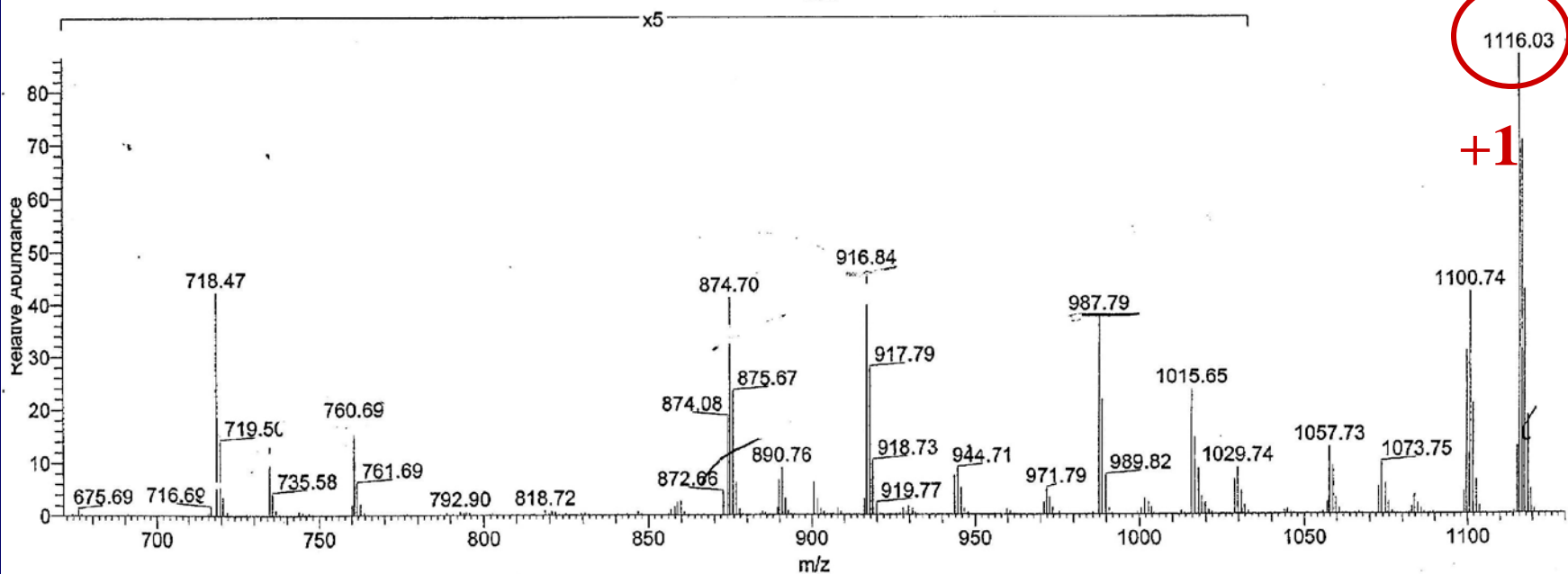
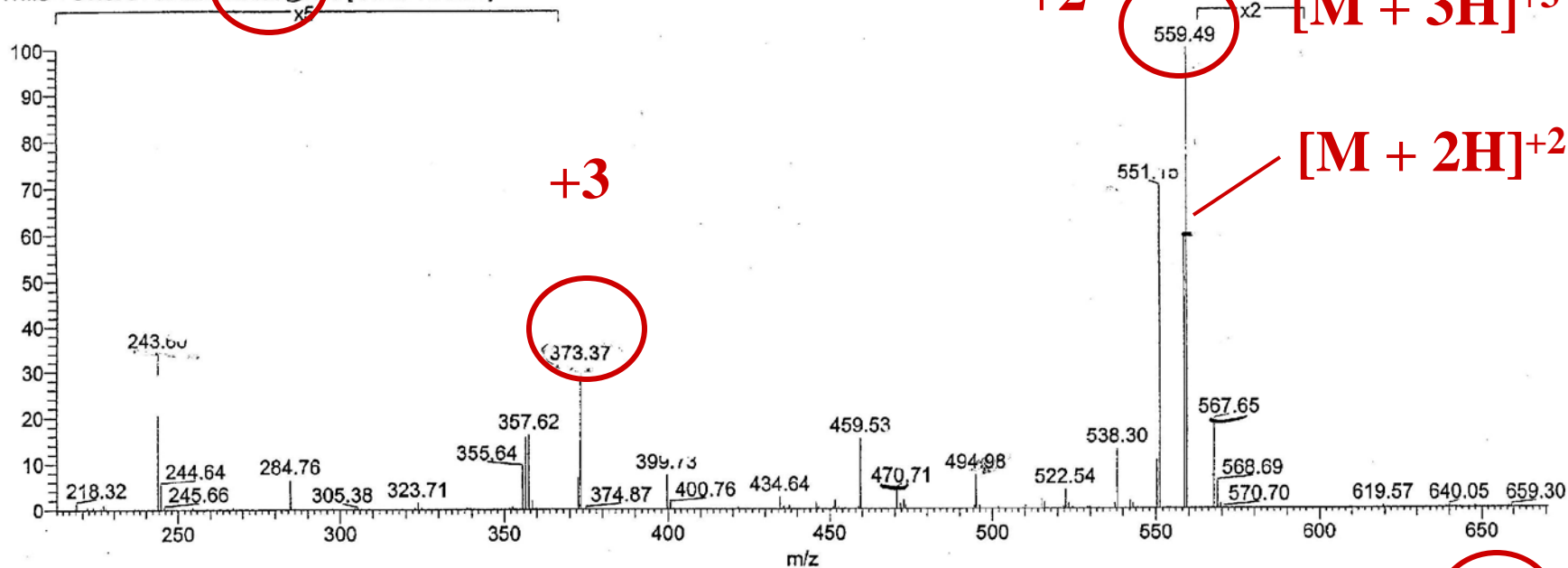


Peptides

td test

jys\JYB7\IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms 372.85@0.00 [90.00-1130.00]



Peptide

td test

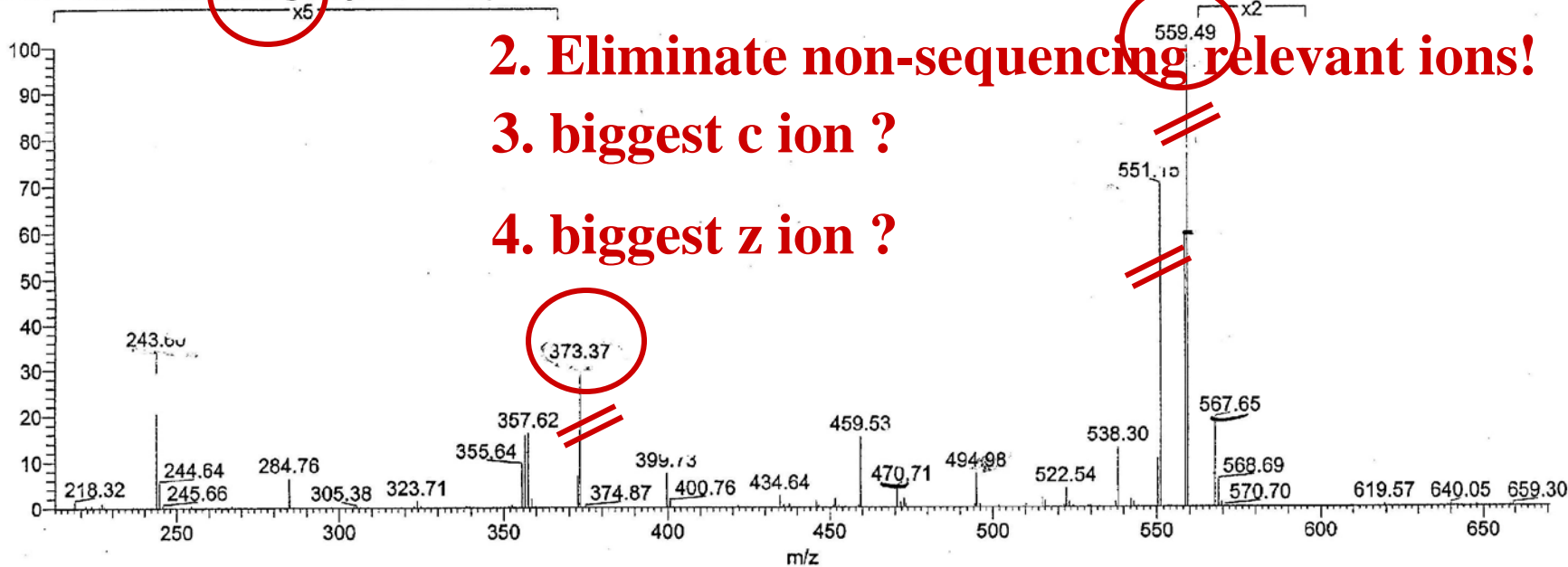
ijsJYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms: 372.85@0.00 [90.00-1130.00]

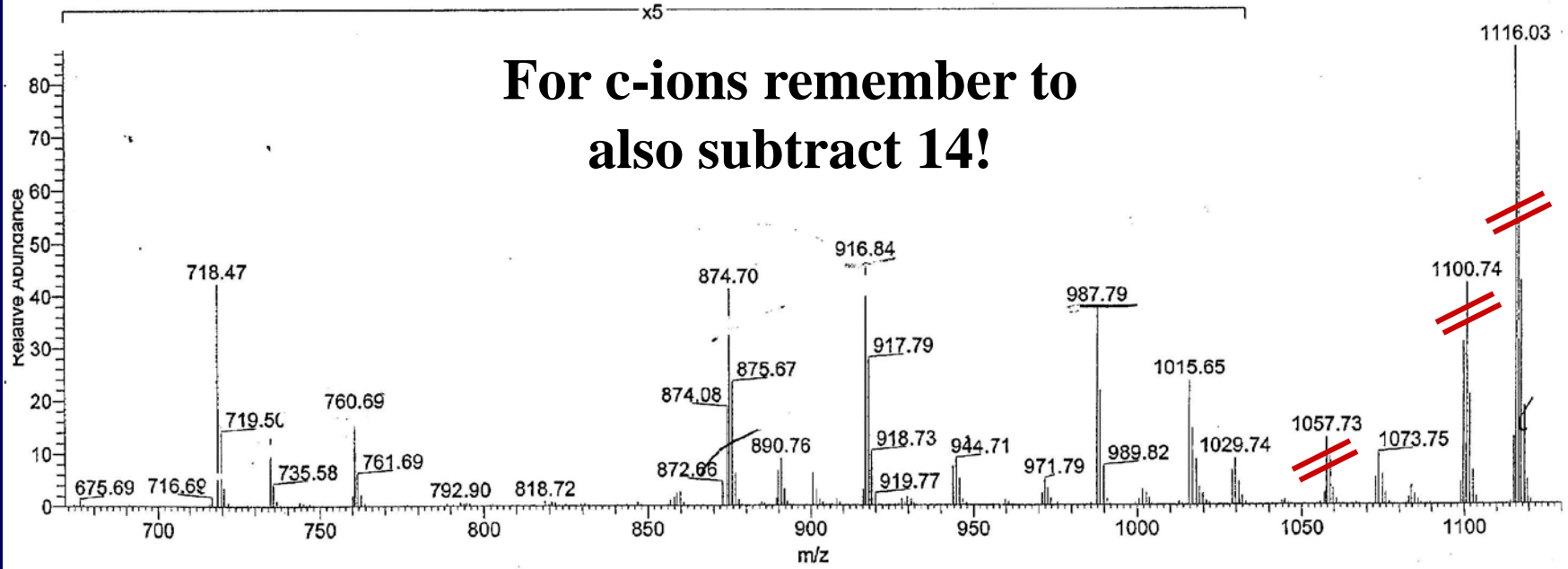
2. Eliminate non-sequencing relevant ions!

3. biggest c ion ?

4. biggest z ion ?



For c-ions remember to also subtract 14!

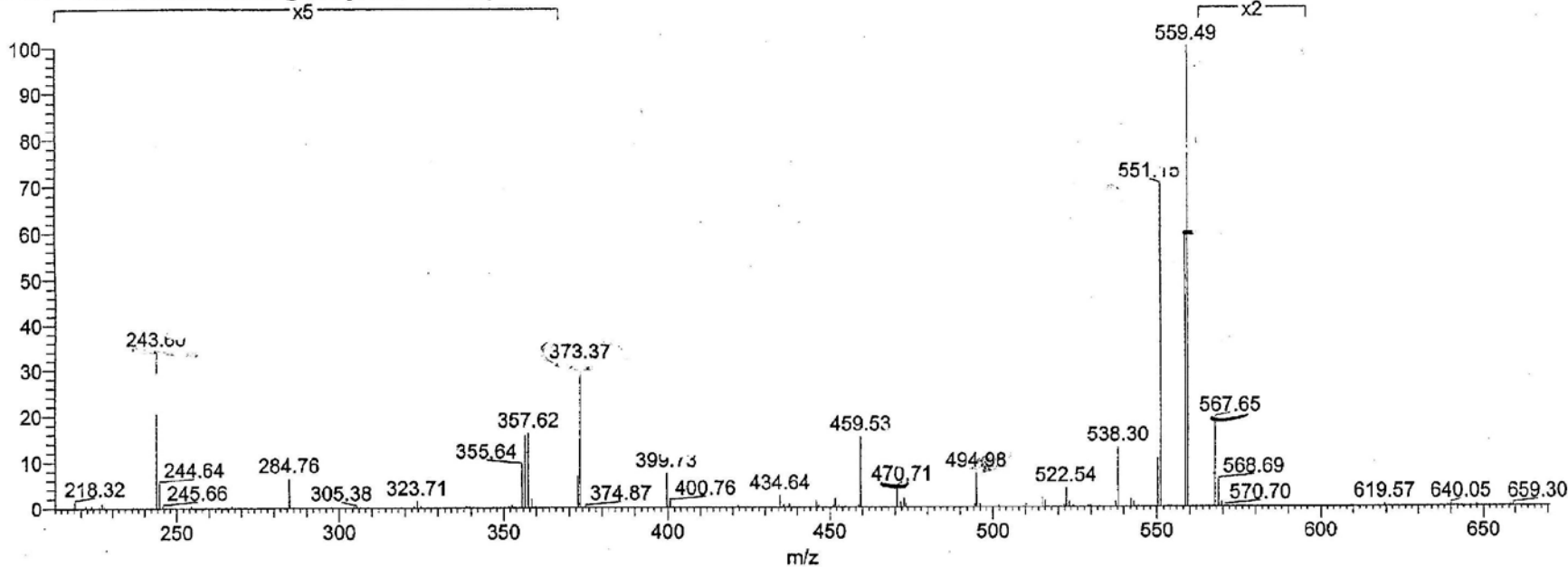


Peptide

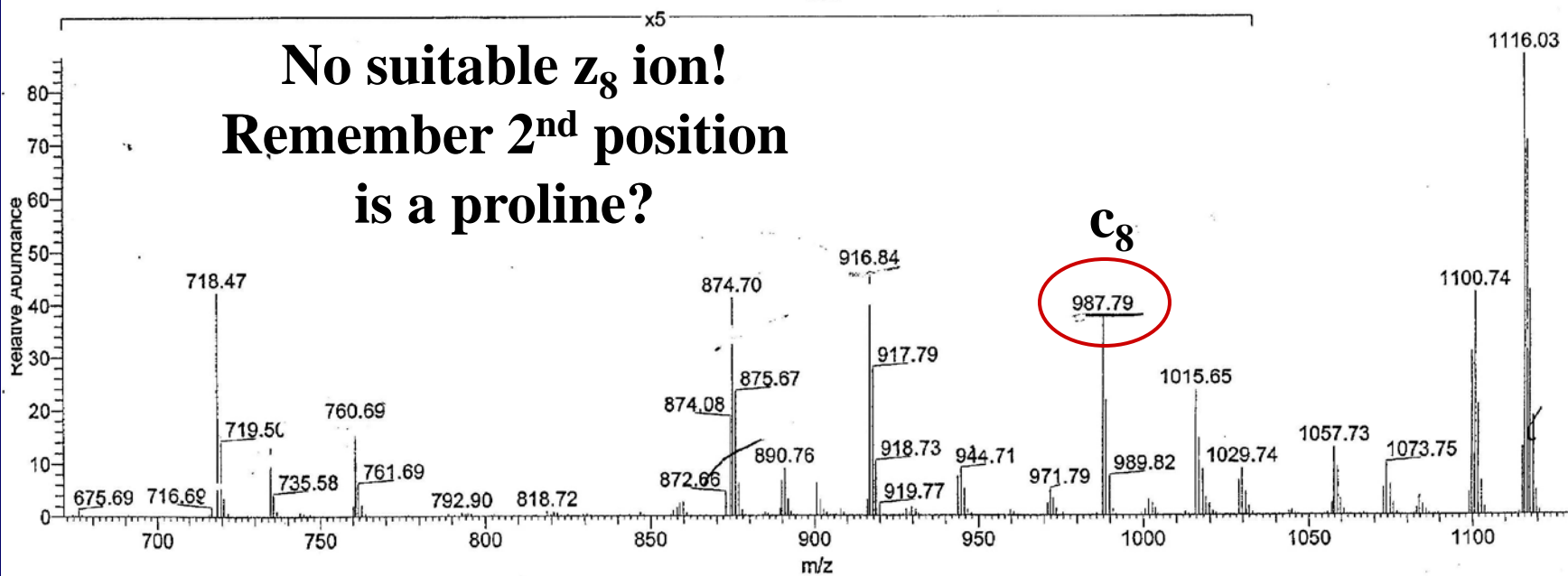
td test

ijsJYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms2 372.85@0.00 [90.00-1130.00]



No suitable z₈ ion!
Remember 2nd position
is a proline?

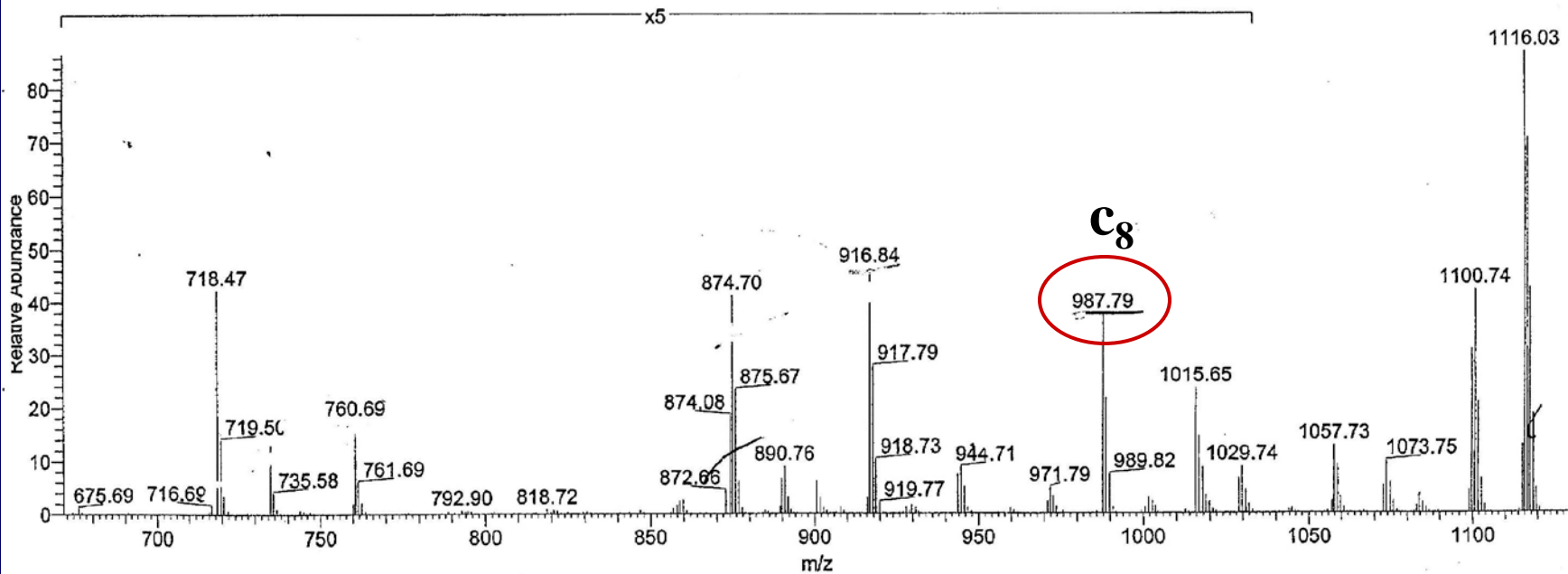
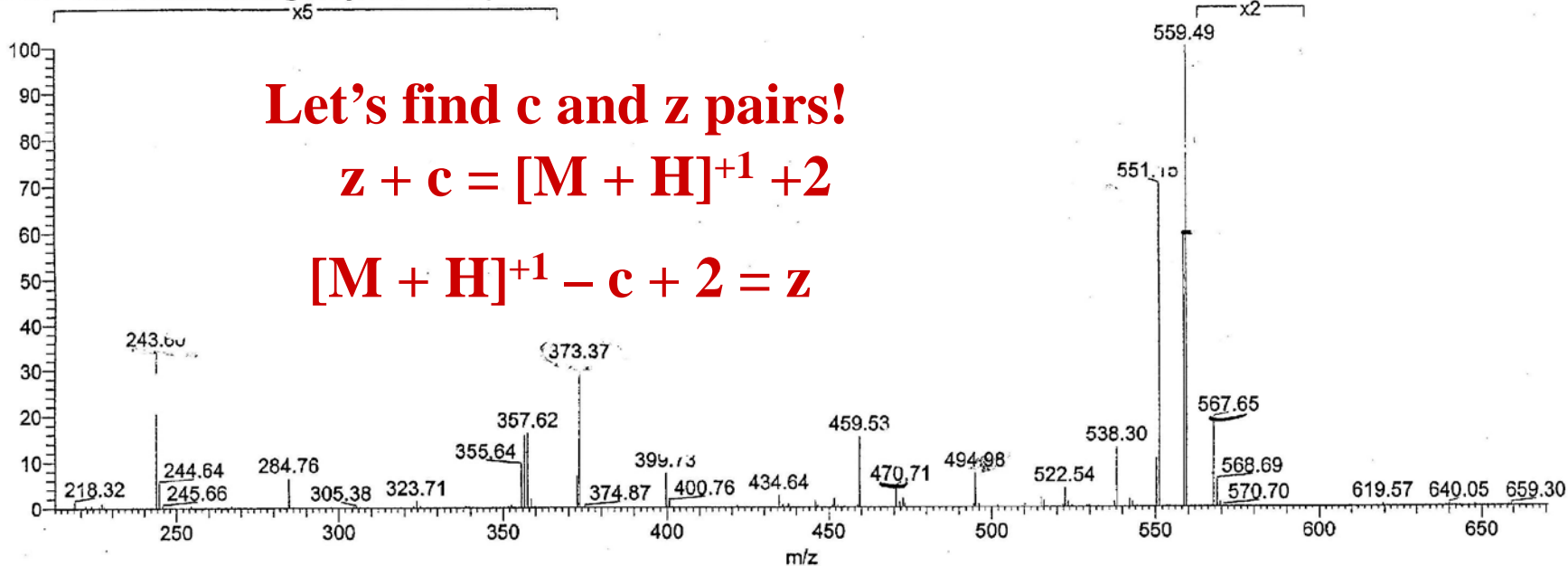


Peptide

td test

ijsJYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms2 372.85@0.00 [90.00-1130.00]

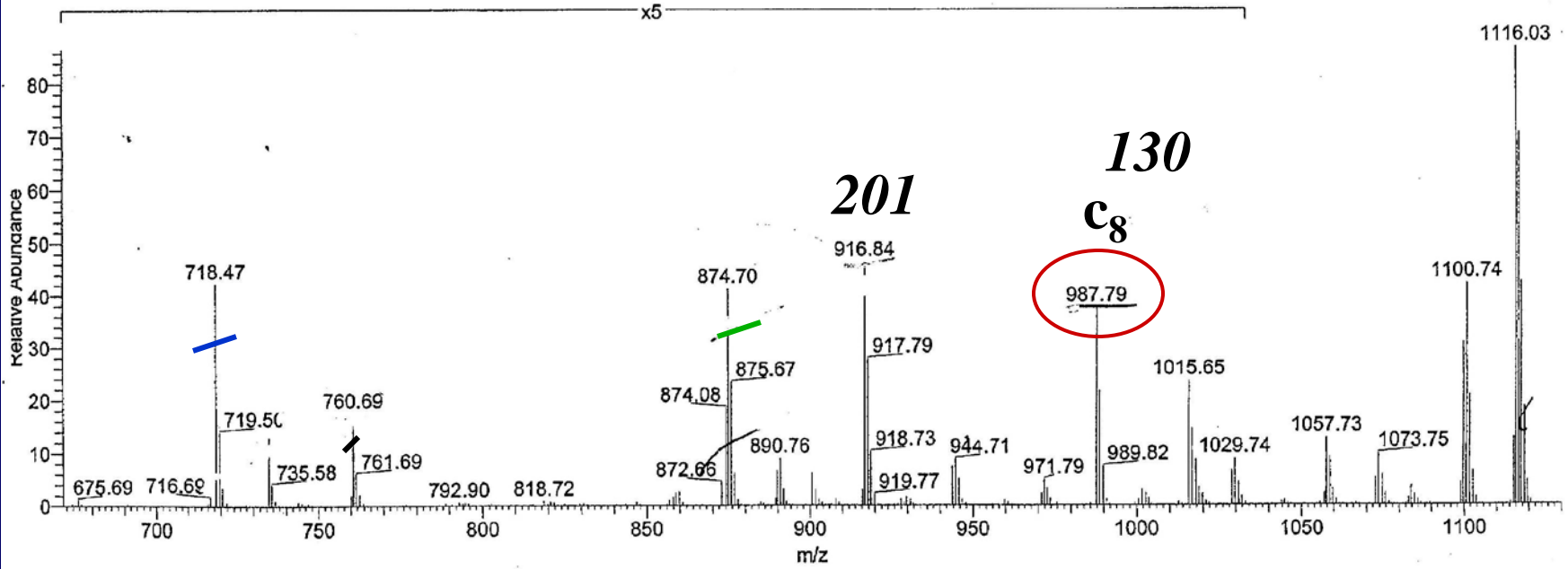
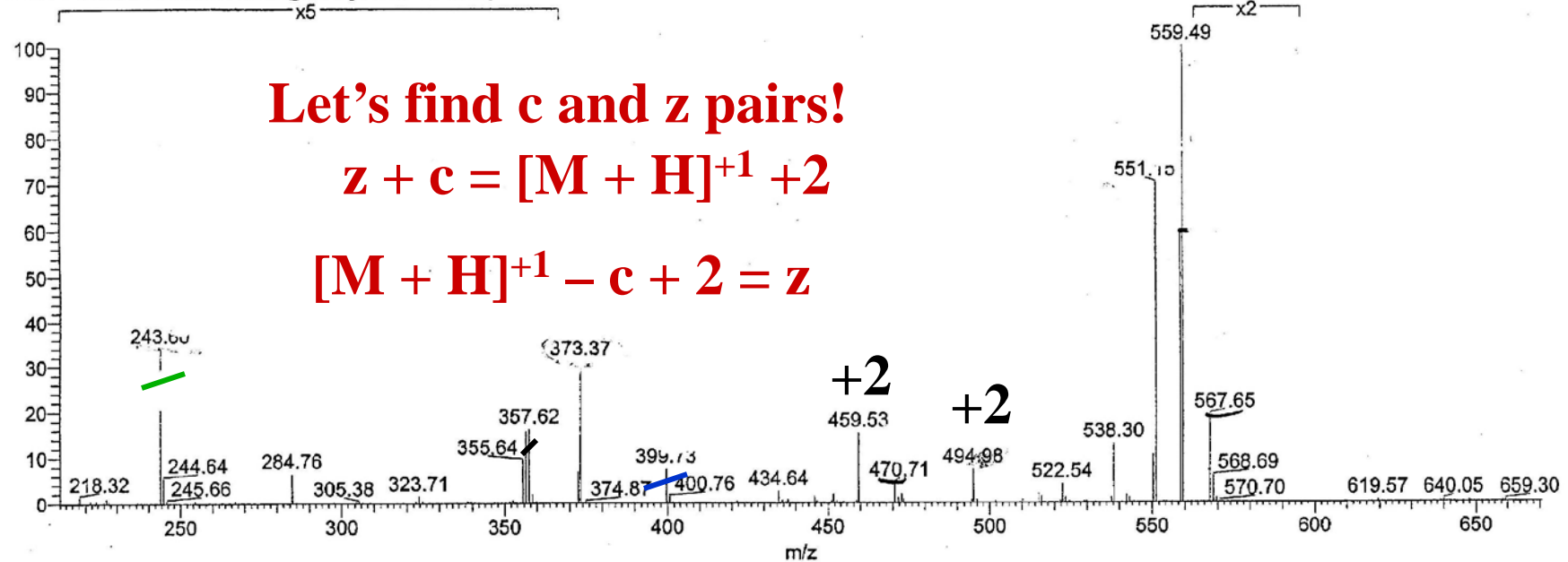


Peptide

td test

ijsJYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms2 372.85@0.00 [90.00-1130.00]

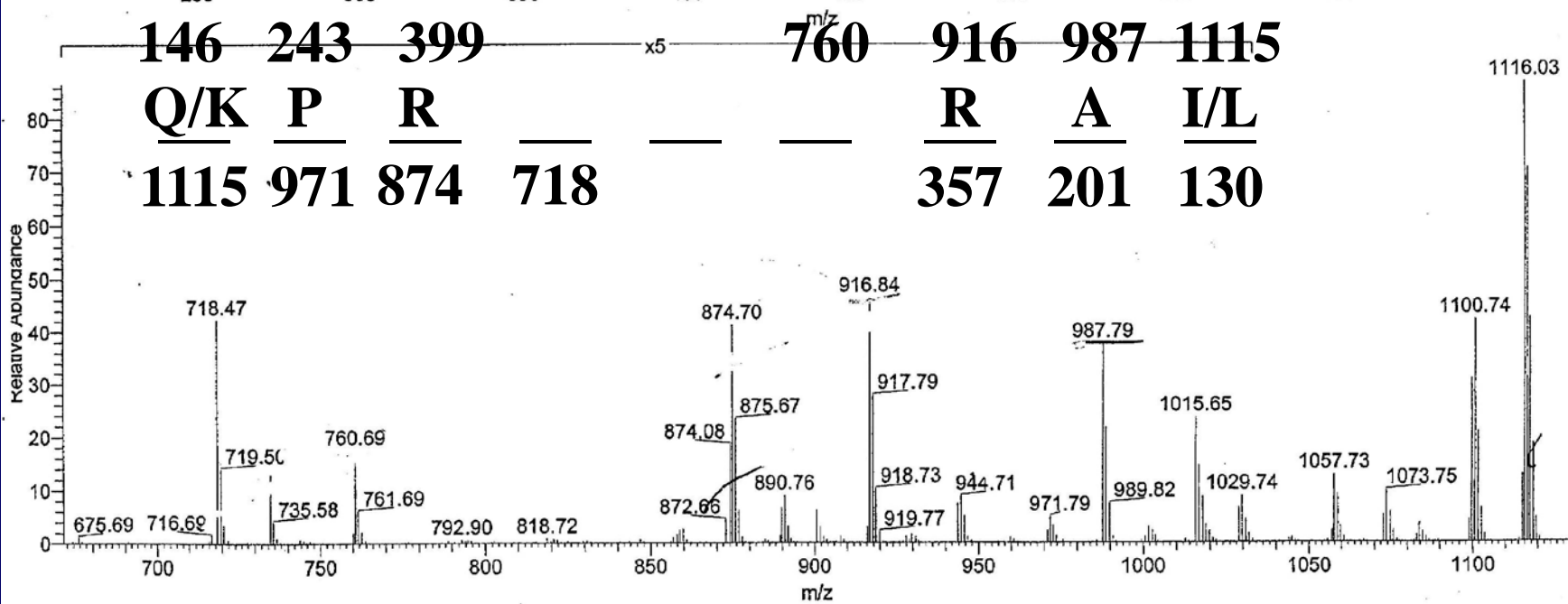
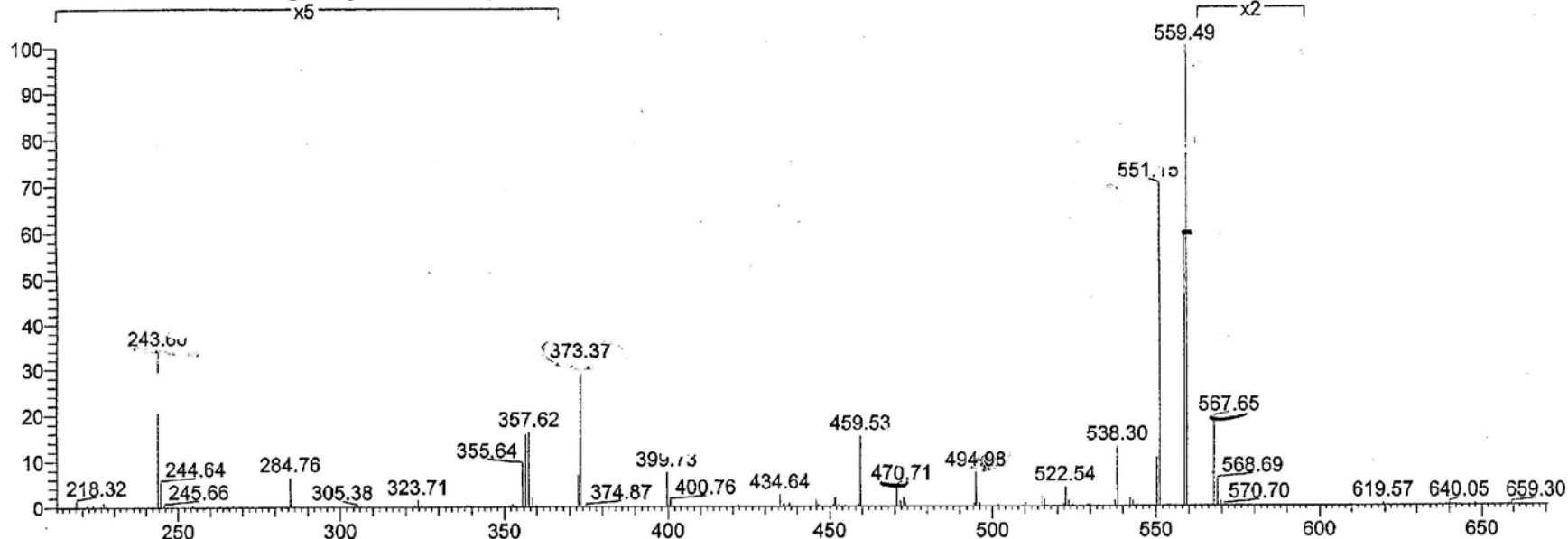


Peptide

td test

ijsJYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms2 372.85@0.00 [90.00-1130.00]

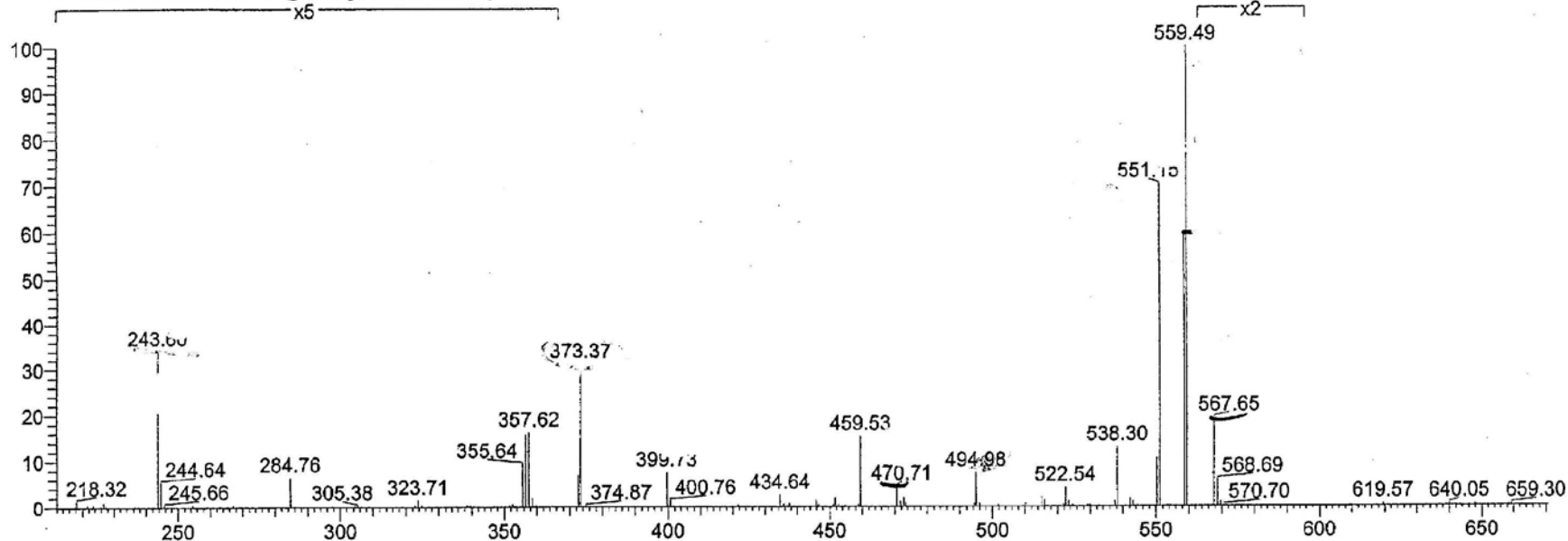


Peptide

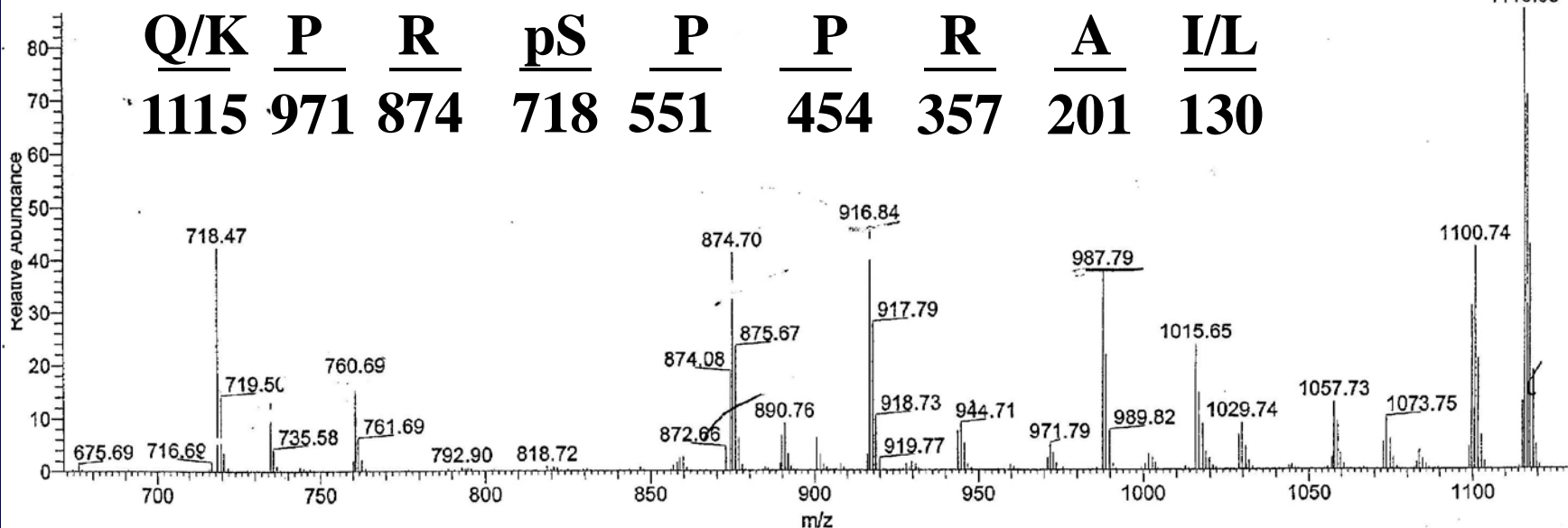
td test

ijs\JYB7IMAC_030304 #2144 RT: 26.21 AV: 1 NL: 2.12E6

: ITMS + c NSI d Full ms2 372.85@0.00 [90.00-1130.00]



146 **243** **399** **566** **663** **760** **916** **987** **1115**
Q/K **P** **R** **pS** **P** **P** **R** **A** **I/L**
1115 **971** **874** **718** **551** **454** **357** **201** **130**



**Proteomics Informatics -
Protein identification III:
de novo sequencing (Week 6)**
